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(54) Title: PHYTASE VARIANTS

(57) Abstract

Phytase variants, their preparation and uses, which phytase variants, when aligned according to Fig. 1, are amended as compared to a model phytase in at least one of a number of positions. Preferred model phytases are basidiomycete and ascomycete phytases, such as improved specific activity and/or improved thermostability.

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Phytase variants

FIELD OF THE INVENTION

This invention relates to variants of phytases, in particular variants of ascomycete phytases and variants of basidiomycete phytases, the corresponding cloned DNA sequences, a method of producing such phytase variants, and the use thereof for a number of industrial applications.

BACKGROUND OF THE INVENTION

- Phytic acid or myo-inositol 1,2,3,4,5,6-hexakis dihydrogen phosphate (or for short myo-inositol hexakisphosphate) is the primary source of inositol and the primary storage form of phosphate in plant seeds. Phytin is a mixed potassium, magnesium and calcium salt of inositol.
- The phosphate moieties of phytic acid chelates divalent and trivalent cations such as metal ions, i.a. the nutritionally essential ions of calcium, iron, zinc and magnesium as well as the trace minerals manganese, copper and molybdenum.
- Phytic acid and its salts, phytates, are often not 20 metabolized, i.e. neither the phosphorous thereof, nor the chelated metal ions are nutritionally available.

Accordingly, food and feed preparations need to be supplemented with inorganic phosphate and often also the nutritionally essential ions such as iron and calcium, must be supplemented.

Still further, the phytate phosphorus passes through the gastrointestinal tract of such animals and is excreted with the manure, resulting in an undesirable phosphate pollution of the environment resulting e.g. in eutrophication of the water 30 environment and extensive growth of algae.

Phytic acid or phytates, said terms being, unless otherwise indicated, in the present context used synonymously or at random, are degradable by phytases.

The production of phytases by plants as well as by 5 microorganisms has been reported. Amongst the microorganisms, phytase producing bacteria as well as phytase producing fungiare known.

phytase producing descriptions of There are several filamentous fungi belonging to the fungal phylum of Ascomycota 10 (ascomycetes). In particular, there are several references to phytase producing ascomycetes of the Aspergillus genus such as Aspergillus terreus (Yamada et al., 1986, Agric. Biol. Chem. 322:1275-1282). Also, the cloning and expression of the phytase gene from Aspergillus niger var. awamori has been described 15 (Piddington et al., 1993, Gene 133:55-62). EP 0420358 describes the cloning and expression of a phytase of Aspergillus ficuum (niger). EP 0684313 describes the cloning and expression of phytases of the ascomycetes Aspergillus niger, Myceliophthora thermophila, Aspergillus terreus. Still further, some partial 20 sequences of phytases of Aspergillus nidulans, Talaromyces thermophilus, Aspergillus fumigatus and another strain of Aspergillus terreus are given.

The cloning and expression of a phytase of Thermomyces lanuginosus is described in WO 97/35017.

There is a current need for phytases of amended properties or characteristics, e.g. phytases of increased thermostability, altered pH optimum (a high pH optimum being desirable for invitro processing, a low for in-vivo processing in the gastro-intestinal tract), and/or of a higher specific activity.

SUMMARY OF THE INVENTION

In a first aspect, the invention provides phytase variants, the characteristics of which are amended - as compared to a so-called model phytase.

Any model phytase, which is of a certain similarity to thirteen herein specifically disclosed model phytases, can be made the model of such variants.

In another aspect, the invention relates to a novel phytase derived from Cladorrhinum foecundissimum.

In still another aspect, the invention provides DNA sequences encoding these phytase variants and this phytase, and methods of their production.

Finally, the invention also relates generally to the use of the phytase and the phytase variants for liberating phosphorous from any phytase substrate, in particular inorganic phosphate from phytate or phytic acid.

BRIEF DESCRIPTION OF THE DRAWINGS

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In the detailed description of the invention below, 20 reference is made to the drawings, of which

- Fig. 1 is an alignment of thirteen specific phytase sequences (a multiple sequence alignment according to the program PileUp; GapWeight: 3.000; GapLengthWeight: 0.100);
- Fig. 2 this figure shows the amino acid and DNA sequence of a first phytase ("P_involtus-A1") derived from strain CBS 100231 of Paxillus involutus which was deposited on 28.11.97; the expression plasmid pYES 2.0 comprising the full length cDNA sequence was

transformed into E. coli strain DSM 11842 which was deposited on 12.11.97 (see WO 98/28409);

- Fig. 3 this figure shows the amino acid and DNA sequence of a second phytase ("P_involtus-A2") derived from strain CBS 100231 of Paxillus involutus which was deposited on 28.11.97; the expression plasmid pYES 2.0 comprising the full length cDNA sequence was transformed into E. coli strain DSM 11843 which was deposited on 12.11.97 (see WO 98/28409);
- this figure shows the amino acid and DNA sequence of Fig. 4 from strain ("T pubescens") derived phytase which pubescens, Trametes CBS 100232 of deposited on 28.11.97; the expression plasmid pYES 15 2.0 comprising the full length cDNA sequence was transformed into E. coli strain DSM 11844 which was deposited on 12.11.97 (see WO 98/28409);
- this figure shows the amino acid and DNA sequence of a phytase ("A_pediades") derived from strain CBS 900.96 of Agrocybe pediades deposited on 04.12.96; the expression plasmid pYES 2.0 comprising the full length cDNA sequence was transformed into E. colistrain DSM 11313 which was deposited on 02.12.96 (see WO 98/28409);
- Fig. 6 this figure shows the amino acid and DNA sequence of a phytase ("P_lycii") derived from strain CBS 686.96 of Peniophora lycii which was deposited on 04.12.96; the expression plasmid pYES 2.0 comprising the full

length cDNA sequence was transformed into E. coli strain DSM 11312 which was deposited on 02.12.96 (see WO 98/28409);

this figure equals figure 2 of EP 0684313 and shows the amino acid and DNA sequence of a phytase ("M_thermophila") derived from strain ATCC 48102 (=ATCC 74340) of Myceliophthora thermophila which was re-deposited on 14.03.97;

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- Fig. 8 this figure shows the amino acid and DNA sequence of a phytase ("A_fumigatus") derived from strain ATCC 13073 of Aspergillus fumigatus (see EP 0897985);
- this figure shows the amino acid ("Conphys") and DNA sequence of an ascomycete consensus phytase (in the present context called "consphyA") (see EP 0897985);
- Fig. 10 this figure shows the amino acid and DNA sequence of

 a phytase ("A_nidulans") derived from strain

 DSM 9743 of Aspergillus nidulans (see EP 0897985);
- Fig. 11 this figure equals figure 8 of EP 0420358 and shows the amino acid and DNA sequence of a phytase ("A_ficuum") derived from Aspergillus ficuum strain NRRL-3135;
- Fig. 12 this figure equals figure 1 of EP 0684313 and shows the amino acid and DNA sequence of a phytase ("A_terreus") derived from strain CBS 220.95 of Aspergillus terreus;

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- Fig. 13 this figure shows the amino acid and DNA sequence of a phytase ("T_thermo") derived from strain ATCC 20186 (=ATCC 74338) of Talaromyces thermophilus which was redeposited on 14.03.97 (see EP 0897985);
- Fig. 14 this figure equals figure 2 of WO 97/35017 and shows the amino acid and DNA sequence of a phytase ("T_lanuginosa") derived from strain CBS 586.94 of Thermomyces lanuginosus; a plasmid comprising the full length cDNA sequence was transformed into E. coli DH5α (pMWR46) strain B-21527 which was deposited with NRRL on 23.02.96;
- this figure shows the amino acid and DNA sequence of a phytase ("C_foecundissimum") derived from strain CBS 427.97 of Cladorrhinum foecundissimum which was deposited on 23 January 1997; the expression plasmid pYES 2.0 comprising the full length cDNA sequence was transformed into E. coli strain DSM 12742 which was deposited on 17 March 1999;
- Fig. 16 this figure shows an alignment of the phytase C_foecundissimum with the model phytase M_thermophila, using the program GAP gcg (Gap Weight 3.000; Length Weight 0.100); and
 - Fig. 17 shows how the C_foecundissimum phytase can be pasted onto the alignment of Fig. 1.

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DETAILED DISCLOSURE OF THE INVENTION

Phytase

In the present context a phytase is an enzyme which catalyzes the hydrolysis of phytate (myo-inositol 5 hexakisphosphate) to (1) myo-inositol and/or (2) mono-, di-, tri-, tetra- and/or penta-phosphates thereof and (3) inorganic phosphate. In the following, for short, the above compounds are sometimes referred to as IP6, I, IP1, IP2, IP3, IP4, IP5 and P, respectively. This means that by action of a phytase, IP6 is 10 degraded into P + one or more of the components IP5, IP4, IP3, IP2, IP1 and I. Alternatively, myo-inositol carrying in total n phosphate groups attached to positions p, q, r,.. is denoted Ins(p,q,r,..)Pn. For convenience Ins(1,2,3,4,5,6)P6(phytic acid) is abbreviated PA.

According to the Enzyme nomenclature database ExPASy (a 15 repository of information relative to the nomenclature of enzymes primarily based on the recommendations of the Nomenclature Committee of the International Union Biochemistry and Molecular Biology (IUBMB) describing each type 20 of characterized enzyme for which an EC (Enzyme Commission) number has been provided), two different types of phytases are known: A so-called 3-phytase (myo-inositol hexaphosphate 3phosphohydrolase, EC 3.1.3.8) and a so-called 6-phytase (myoinositol hexaphosphate 6-phosphohydrolase, EC 3.1.3.26). The 3-25 phytase hydrolyses first the ester bond at the D-3-position, whereas the 6-phytase hydrolyzes first the ester bond at the D-6- or L-6-position.

The expression "phytase" or "polypeptide or enzyme exhibiting phytase activity" is intended to cover any enzyme capable of effecting the liberation of inorganic phosphate or phosphorous from various myo-inositol phosphates. Examples of

such myo-inositol phosphates (phytase substrates) are phytic acid and any salt thereof, e.g. sodium phytate or potassium phytate or mixed salts. Also any stereoisomer of the mono-, di-, tri-, tetra- or penta-phosphates of myo-inositol might serve as a phytase substrate. A preferred phytase substrate is phytic acid and salts thereof.

In accordance with the above definition, the phytase activity can be determined using any assay in which one of these substrates is used. In the present context (unless otherwise specified) the phytase activity is determined in the unit of FYT, one FYT being the amount of enzyme that liberates 1 μ mol inorganic ortho-phosphate per min. under the following conditions: pH 5.5; temperature 37°C; substrate: sodium phytate $(C_6H_6O_{24}P_6Na_{12})$ in a concentration of 0.0050 mol/l. A suitable phytase assay is described in the experimental part.

The present invention provides a genetically engineered phytase as described in the appending claims.

A genetically engineered phytase is a non-naturally occurring phytase which is different from a model phytase, e.g. a wild-type phytase. Genetically engineered phytases include, but are not limited to, phytases prepared by site-directed mutagenesis, gene shuffling, random mutagenesis etc.

The invention also provides DNA constructs, vectors, host cells, and methods of producing these genetically engineered phytases and phytase variants, as well as uses thereof.

A phytase variant is a polypeptide or enzyme or a fragment thereof which exhibits phytase activity and which is amended as compared to a model phytase.

Amended means altered by way of one or more amino acid or 30 peptide substitutions, deletions, insertions and/or additions - in each case by, or of, one or more amino acids. Such

substitutions, deletions, insertions, additions can be achieved by any method known in the art, e.g. gene shuffling, random mutagenesis, site-directed mutagenesis etc.

The model or parent phytase, from which the phytase 5 variant is derived, can be any phytase, e.g. a wild-type phytase or a derivative, mutant or variant thereof, including allelic and species variants, as well as genetically engineered variants thereof, which e.g. can be prepared by site-directed mutagenesis, random mutagenesis, shuffling etc.

Included in the concept of model phytase is also any hybrid or chimeric phytase, i.e. a phytase which comprises a combination of partial amino acid sequences derived from at least two phytases.

The hybrid phytase may comprise a combination of partial amino acid sequences deriving from at least two ascomycete phytases, at least two basidiomycete phytases or from at least one ascomycete and at least one basidiomycete phytase. These ascomycete and basidiomycete phytases from which a partial amino acid sequence derives may, e.g., be any of those specific phytases referred to herein.

the present context, a hybrid, shuffled, mutagenised, site-directed mutagenised or otherwise genetically engineered phytase derived from ascomycete phytases only is also an ascomycete phytase; and a hybrid, shuffled, 25 mutagenised, site-directed mutagenised or otherwise genetically engineered phytase derived from model basidiomycete phytases only is also a basidiomycete phytase. Any hybrid derived from at ascomycete phytase as least one well as basidiomycete phytase is called a mixed ascomycete/basidiomycete 30 phytase and such phytase is also a model phytase in the present context.

Analogously, a hybrid, shuffled, random mutagenised, sitedirected mutagenised or otherwise genetically engineered phytase
derived from one or more Aspergillus phytases is also an
Aspergillus derived phytase; and a hybrid, shuffled, random
mutagenised, site-directed mutagenised or otherwise genetically
engineered phytase derived from any other taxonomic sub-grouping
mentioned herein is also to be designated a phytase derived from
this taxonomic sub-grouping.

Still further, in the present context, "derived from" is intended to indicate a phytase produced or producible by a strain of the organism in question, but also a phytase encoded by a DNA sequence isolated from such strain and produced in a host organism transformed with said DNA sequence. Finally, the term is intended to indicate a phytase which is encoded by a DNA sequence of synthetic and/or cDNA origin and which has the identifying characteristics of the phytase in question.

Preferably the model phytase is a phytase which can be aligned as described below to either of the thirteen phytases of Fig. 1 (which are particularly preferred model phytases).

Preferred wild-type model phytases (i.e. neither recombinant, or shuffled or otherwise genetically engineered phytases) have a degree of similarity or homology, preferably identity, to amino acid sequence no. 38-403 (Peniophora numbers) of either of these thirteen phytases of at least 40%, more preferably at least 50%, still more preferably at least 60%, in particular at least 70%, especially at least 80%, and in a most preferred embodiment a degree of similarity of at least 90%.

Preferred recombinant or shuffled or otherwise genetically engineered model phytases have a degree of similarity or homology, preferably identity, to partial sequence no. 38-49, 63-77, 274-291, 281-300 and 389-403 (Peniophora numbers) of

either of these thirteen phytases of at least 60%, more preferably at least 70%, still more preferably at least 80%, in particular at least 90%.

In a preferred embodiment the degree of similarity is 5 based on a comparison with the complete amino acid sequence of either of the thirteen phytases.

The degree of similarity or homology, alternatively identity, can be determined using any alignment programme known in the art. A preferred alignment programme is GAP provided in the GCG version 8 program package (Program Manual for the Wisconsin Package, Version 8, August 1994, Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA 53711) (see also Needleman, S.B. and Wunsch, C.D., (1970), Journal of Molecular Biology, 48, 443-453). Using GAP with the following settings for polypeptide sequence comparison: GAP weight of 3.000 and GAP lengthweight of 0.100.

Also preferred is a wild-type model phytase which comprises an amino acid sequence encoded by a DNA sequence which hybridizes to a DNA sequence encoding amino acid sequence 38-403 (Peniophora numbers) of any of the DNA sequences encoding the thirteen specific phytase sequences of Fig. 1.

A further preferred model phytase is a genetically engineered phytase, which comprises an amino acid sequence encoded by a DNA sequence which hybridizes to a DNA sequence 25 encoding amino acid sequence 38-49, and to a DNA sequence encoding amino acid sequence 63-77, and to a DNA sequence encoding amino acid sequence 274-291, and to a DNA sequence encoding amino acid sequence 281-300, and to a DNA sequence encoding amino acid sequence 281-300, and to a DNA sequence encoding amino acid sequence 389-403 (Peniophora numbers) of any 30 of the DNA sequences encoding the thirteen specific phytase sequences of Fig. 1.

In a preferred embodiment the hybridization is to the complete phytase encoding part of any of the thirteen phytases.

Suitable experimental conditions for determining whether a given DNA or RNA sequence "hybridizes" to a specified nucleotide or oligonucleotide probe involves presoaking of the filter containing the DNA fragments or RNA to examine for hybridization in 5 x SSC (Sodium chloride/Sodium citrate), (J. Sambrook, E.F. Fritsch, and T. Maniatis, 1989, Molecular Cloning, A Laboratory Manual, 2d edition, Cold Spring Harbor, New York) for 10 min, and prehybridization of the filter in a solution of 5 x SSC, 5 x Denhardt's solution (Sambrook et al. 1989), 0.5 % SDS and 100 µg/ml of denatured sonicated salmon sperm DNA (Sambrook et al. 1989), followed by hybridization in the same solution containing a concentration of 10 ng/ml of a random-primed (Feinberg, A. P. and Vogelstein, B. (1983) Anal. Biochem. 132:6-13), 32P-dCTP-labeled (specific activity > 1 x 109 cpm/µg) probe for 12 hours at approximately 45°C.

The filter is then washed twice for 30 minutes in 2 x SSC, 0.5 % SDS at at least 55°C (low stringency), at at least 60°C (medium stringency), at at least 65°C (medium/high stringency), at at least 70°C (high stringency), or at at least 75°C (very high stringency).

Molecules to which the oligonucleotide probe hybridizes under these conditions are detected using an x-ray film.

It should be noted that a certain specific phytase variant need not actually have been prepared from a specific model phytase, for this model phytase to qualify as a "model phytase" in the present context. It is sufficient that the variant exhibits at least one of the herein indicated amendments when it is afterwards compared with the model phytase.

The alignment of Fig. 1 is made using the program PileUp (Program Manual for the Wisconsin Package, Version 8, August 1994, Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA 53711), with a GapWeight of 3.000 and a 5 GapLengthWeight of 0.100. When aligning a new model phytase or a new phytase variant all thirteen sequences can be included together with the new phytase (variant) in a multiple alignment, or, alternatively, at least one of the thirteen sequences of Fig. 1 is included together with the new phytase (variant) in an alignment.

A preferred procedure for aligning according to Fig. 1 a new model phytase (or a phytase variant) is as follows: The new model phytase is aligned with that specific sequence of the thirteen sequences of Fig. 1 to which the new model phytase has 15 the highest degree of homology. For calculating the degree of homology, and for making the "alignment according to Fig. 1" of the two sequences, the program GAP referred to below is preferably used. Having aligned the two sequences, the new model phytase (or phytase variant) is added (pasted) to the alignment 20 at Fig. 1 using the result of the first alignment (placing identical and homologous amino acid residues above each other as prescribed by the alignment), following which corresponding positions are now easily identifiable.

Example 7 shows an example of how to add a new model phytase to the alignment of Fig. 1 and deduce corresponding phytase variants thereof.

Other model phytases can be aligned and variants deduced in analogy with Example 7. This is so in particular for the following model phytases: The phytase of Aspergillus niger var.

30 awamori (US patent no. 5,830,733); the Bacillus phytase of WO 98/06858; the soy bean phytase of WO 98/20139; the maize

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phytase of WO 98/05785; the Aspergillus phytase of WO 97/38096; the phytases of Monascus anka of WO 98/13480; the phytase from Schwanniomyces occidentalis of EP 0699762 etc.

When comparing a model phytase and a proposed phytase 5 variant using the alignment as described herein, corresponding amino acid positions can be identified, viz. a model position of the model phytase and a variant position of the variant — the corresponding model position and variant position are simply placed one above the other in the alignment. An amendment is said to have occurred in a given position if the model amino acid of the model position and the variant amino acid of the variant position are different. Preferred amendments of these positions manifest themselves as amino acid substitutions, deletions or additions.

15 Amended in at least one position means amended in one or more positions, i.e. in one, two, three, four, five, six, seven, eight, nine, ten, eleven, twelve etc. up to all N positions listed. This definition includes any possible sub-combinations thereof, e.g. any set of two substitutions, any set of three, 20 any set of four, etc. - to any set of (N-1) positions.

In the present context all sequences, whatever the model phytase, and including the thirteen sequences of Fig. 1, are numbered using the numbering corresponding to the phytase P_lycii. These "Peniophora numbers" are indicated at Fig. 1, together with the "alignment numbers." The numbering of P_lycii starts at M1 and ends at E439.

As explained above, the alignment reveals which positions in various phytase sequences other than P_lycii are equivalent or corresponding to the given P. lycii position.

A substitution of amino acids is indicated herein as for instance "3S," which indicates, that at position 3 amino acid S

should be substituted for the "original" or model position 3 amino acid, whichever it is. Thus, the substitution should result in an S in the corresponding variant position. Considering now the alignment at Fig. 1, a substitution like 5 e.g. "3S" is to be interpreted as follows, for the respective phytases shown (the amino acid first indicated is the "original" or model amino acid in "Peniophora position" 3):

	P_involtus_A1:	F3S (number 3 F substituted by S)			
	P_involtus_A2:	L3s			
10	T_pubescens:	M1s			
	A_pediades:	M1s			
15 A	P_lycii:	redundant (already an S)			
	A_fumigatus:	T5s			
	consphyA:	V5s			
	A_nidulans:	T5s			
	A_ficuum_NRRL3135:	A5s			
	A_terreus:	A5s			
	T_thermo:	L5s			
	T_lanuginosa:	V11s			
20	M_thermophila:	G5S			

However, in what follows the above specific substitutions will be designated as follows (always using the Peniophora numbering):

25	P_involtus_A1:	F3S
	P_involtus_A2:	L3S
	T_pubescens:	M3S
	A_pediades:	M3S
	P_lycii:	redundant (already an S)
30	A_fumigatus:	T3S
	consphyA:	V3S

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A_nidulans: T3S
A_ficuum_NRRL3135: A3S
A_terreus: A3S
T_thermo: L3S
T_lanuginosa: V3S
M_thermophila: G3S

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Still further, denotations like e.g. "3S,F,G" means that the amino acid in position 3 (Peniophora numbers) of the model 10 phytase in question is substituted with either of S, F or G, i.e. e.g. the designation "3S,F,G" is considered fully equivalent to the designation "3S, 3F, 3G".

A denotation like ()3S means that amino acid S is added to the sequence in question (at a gap in the actual sequence), in a position corresponding to Peniophora number 3 - and vice versa for deletions (S3()).

In case of regions in which the Peniophora phytase sequence has larger deletions than some of the other phytases in Fig. 1, for instance in the region between position 201 and 202 (Peniophora numbers), intermediate positions (amino acid residues in other sequences) are numbered by adding a,b,c,d, etc, in lower-case letters, to the last Peniophora position number, e.g. for the phytase M_thermophila: E201; G201a; P201b; Y201c; S201d; T201e; I201f; G202; D203 etc.

In one of the priority applications of the present application there are two minor position numbering errors:

According to the above definitions, the positions referred to in first priority application as 204 and 205 (Peniophora ers) are wrongly designated; they should have been numbered and 204, respectively. Therefore, 204 has been substituted and 205 by 204 throughout the present application.

A preferred phytase variant of the invention comprises an amino acid sequence which comprises, preferably contains, one or more of the following amino acid substitutions:

24C; 27P: 31Y; 33C; 39H, S, Q; 40L,N; 42S,G; 5 43A,C,D,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y; 44N; 45D,S; 47Y, F; 49P; 51E, A, R; 56P; 58D, K, A; 59G; 61R; 62V, I; 69Q; 75W, F; 78D, S; 79G; 80K, A; 81A, G, Q, E; 82T; 83A, I, K, R, Q; 84I, Y, Q, V; 88I; 90R, A; 102Y; 115N; 116S; 118V,L; 119E; 120L; 122A; 123N,Q,T; 125M,S; 126H, S, V; 127Q, E, N; 128A, S, T; 132F, I, L; 143N; 148V, I; 151A, S; 10 152G; 153D,Y; 154D,Q,S,G; 157V; 158D,A; 159T; 160A,S; 161T,N; 162N; 163W; 170fH; 170gA; 171N; 172P; 173Q,S; 184Q,S,P; 185S; 186A, E, P; 187A; 187aS; 190A, P; 193S; 194S, T; 195T, V, L; 198A, N, V; 200G, V; 201D, E; a deletion of at least one of 201a, 201b, 201c, 201d, 201e, 201f, preferably all; 201eT; 202S,A; 203R,K,S; 15 203aV,T; 204Q,E,S,A,V; 205E; 211L,V; 215A,P; 220L,N; 223H,D; 228N; 232T; 233E; 235Y,L,T; 236Y,N; 237F; 238L,M; 242P,S; 244D; 246V; 251eE,Q; 253P; 256D; 260A,H; 264R,I; 265A,Q; 267D; 270Y, A, L, G; 271D, N; 273D, K; 275F, Y; 278T,H; 280A, P; 283P; 287A,T; 288L,I,F; 292F,Y; 293A,V; 302R,H; 304P,A; 332F; 336S; 20 337T,G,Q,S; 338I; 339V,I; 340P,A; 343A,S,F,I,L; 348Y; 349P; 352K; 360R; 362P; 364W,F; 365V,L,A,S; 366D,S,V; 367A,K; 368K; 369I,L; 370V; 373A,S; 374S,A; 375H; 376M; 383kQ,E; 387P; 393V; 396R; 404A,G; 409R; 411K,T; 412R; 417E,R; 421F,Y; 431E.

In a preferred embodiment this is with the proviso that
the model phytase does not already comprise the above suggested amino acid substitution or addition or deletion at the position indicated. Or, with the proviso that, for each position, the model amino acid is not already the variant amino acid hereby proposed. But these provisos can be said to be in fact already inherent in the above wording, because of the expression "amended."

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The various preferred phytase variants of claims 16-34 comprises, preferably contains or have, amino acid sequences which comprise or contain one or more of the amino acid substitutions, additions, or deletions listed in the respective 5 claims.

In a preferred embodiment the various phytase variants comprise 1, 2, 3, 4, 5, 6, 7, 8, 9 or even 10 of these substitutions; or a number of substitutions of 10-15, 15-20, 20-30 or even 30-50; eventually up to 60, 70, 80 or 90 substitutions.

In another preferred embodiment, the amino acid sequence of the various phytase variants comprise one or more substitutions of the substitution sub-groupings listed hereinbelow; or combinations of substitutions classified in two or more sub-groupings.

Generally, instead of "comprise," "contain" or "have," the amino acid sequences of preferred variants "consist essentially of" or "consist of" the specific model phytases of fig. 1, as modified by one or more of the substitutions described herein.

In the present context a basidiomycete means a microorganism of the phylum Basidiomycota. This phylum of Basidiomycota is comprised in the fungal kingdom together with e.g. the phylum Ascomycota ("ascomycetes").

Taxonomical questions can be clarified by consulting the references listed below or by consulting a fungal taxonomy database (NIH Data Base (Entrez)) which is available via the Internet on World Wide Web at the following address: http://www3.ncbi.nlm.nih.gov/Taxonomy/tax.html.

For a definition of basidiomycetes, reference is made to 30 either Julich, 1981, Higher Taxa of Basidiomycetes; Ainsworth & Bisby's (eds.) Dictionary of the Fungi, 1995, Hawksworth, D.L.,

P.M. Kirk, B.C. Sutton & D.N. Pegler; or Hansen & Knudsen (Eds.), Nordic Macromycetes, vol. 2 (1992) and 3 (1997). A preferred reference is Hansen & Knudsen.

For a definition of ascomycetes, reference is made to 5 either of Ainsworth & Brisby cited above or Systema Ascomycetum by Eriksson, O.E. & D. L. Hawksworth, Vol. 16, 1998. A preferred reference is Eriksson et al.

Generally, a microorganism which is classified as a basidiomycete/ascomycete in either of the references listed 10 above, including the database, is a basidiomycete/ascomycete in the present context.

Some Aspergillus strains are difficult to classify because they are anamorphous, and therefore they might be classified in Fungi Imperfecti. However, once the teleomorphous counterpart is found, it is re-classified taxonomically. For instance, the teleomorph of A. nidulans is Emericella nidulans (of the family Trichocomaceae, the order Eurotiales, the class Plectomycetes of the phylum Ascomycota). These subgroupings of Ascomycota are preferred, together with the family Lasiosphaeriaceae, the order Sordariales, the class Pyrenomycetes of the phylum Ascomycota.

The wording "ascomycetes" and analogues as used herein includes any strains of Aspergillus, Thermomyces, Myceliophthora, Talaromyces which are anamorphous and thus would be classified in Fungi Imperfecti.

Preferred basidiomycete phytases are those listed in WO 98/28409, in the very beginning of the section headed "Detailed description of the invention".

DNA sequences encoding the thirteen specifically listed model phytases and other model phytases can be prepared according to the teachings of each of the documents listed under the brief description of the drawings.

A DNA sequence encoding a model phytase may be isolated from any cell or microorganism producing the phytase in question, using various methods well known in the art. First, a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that produces the phytase. Then, if the amino acid sequence of the phytase is known, homologous, labelled oligonucleotide probes may be synthesized and used to identify phytase-encoding clones from a genomic library prepared from the organism in question.

10 Alternatively, a labelled oligonucleotide probe containing sequences homologous to a known phytase gene could be used as a probe to identify phytase-encoding clones, using hybridization and washing conditions of lower stringency.

Yet another method for identifying phytaseencoding clones
15 would involve inserting fragments of genomic DNA into an
expression vector, such as a plasmid, transforming phytasenegative bacteria with the resulting genomic DNA library, and
then plating the transformed bacteria onto agar containing a
substrate for phytase thereby allowing clones expressing the
20 phytase to be identified.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established standard methods, e.g. the phosphoroamidite method described by S.L. Beaucage and M.H. Caruthers (1981) or the method described by Matthes et al. (1984). In the phosphoroamidite method, oligonucleotides are synthesized, e.g. in an automatic DNA synthesizer, purified, annealed, ligated and cloned in appropriate vectors.

Finally, the DNA sequence may be of mixed genomic and synthetic origin, mixed synthetic and cDNA origin or mixed genomic and cDNA origin, prepared by ligating fragments of synthetic, genomic or cDNA origin (as appropriate, the fragments

corresponding to various parts of the entire DNA sequence), in accordance with standard techniques. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific primers, for instance as described in US 4,683,202 or R.K. Saiki 5 et al. (1988).

DNA encoding the phytase variants of the present invention can be prepared by methods known in the art, such as Sitedirected Mutagenesis. Once a DNA sequence encoding a model phytase of interest has been isolated, and desirable sites for 10 mutation identified, mutations may be introduced using synthetic oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the desired mutation sites: mutant nucleotides are inserted during oligonucleotide synthesis. specific method, a single-stranded gap of DNA, bridging the 15 phytase-encoding sequence, is created in a vector carrying the phytase-encoding gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) and the construct is ligated 20 using T4 ligase. A specific example of this method is described Morinaga et al. (1984). 4,760,025 discloses US introduction of oligonucleotides encoding multiple mutations by performing minor alterations of the cassette. However, an even greater variety of mutations can be introduced at any one time 25 by the Morinaga method because a multitude of oligonucleotides, of various lengths, can be introduced.

Another method of introducing mutations into DNA sequences encoding a desired model phytase is described in Nelson and Long (1989). It involves a 3-step generation of a PCR fragment containing the desired mutation introduced by using a chemically synthesized DNA strand as one of the primers in the PCR

reactions. From the PCR-generated fragment, a DNA fragment carrying the mutation may be isolated by cleavage with restriction endonucleases and reinserted into an expression plasmid.

Yet another method of mutating DNA sequences encoding a model phytase is Random Mutagenesis. Random mutagenesis is suitably performed either as localised or region-specific random mutagenesis in at least three parts of the gene translating to the amino acid sequence shown in question, or within the whole gene.

The random mutagenesis of a DNA sequence encoding a model phytase may be conveniently performed by use of any method known in the art.

In relation to the above, further aspects of the present invention relates to a method for generating a variant of a model phytase, wherein the variant preferably exhibits amended characteristics as described below, the method comprising:

- (a) subjecting a DNA sequence encoding the model phytase to Site-directed Mutagenesis, or the Nelson and Long PCR 20 mutagenesis method or to Random Mutagenesis,
 - (b) expressing the mutated DNA sequence obtained in step(a) in a host cell, and
- (c) screening for host cells expressing a phytase variant which has an altered property relative to the model 25 phytase.

When using Random Mutagenesis, step (a) of the above method of the invention is preferably performed using doped primers.

For instance, the random mutagenesis may be performed by 30 use of a suitable physical or chemical mutagenizing agent, by use of a suitable oligonucleotide, or by subjecting the DNA

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sequence to PCR generated mutagenesis. Furthermore, the random mutagenesis may be performed by use of any combination of these mutagenizing agents. The mutagenizing agent may, e.g., be one which induces transitions, transversions, inversions, 5 scrambling, deletions, and/or insertions.

Examples of a physical or chemical mutagenizing agent suitable for the present purpose include ultraviolet (UV) irradiation, hydroxylamine, N-methyl-N'-nitro-N-nitrosoguanidine (MNNG), O-methyl hydroxylamine, nitrous acid, ethyl methane sulphonate (EMS), sodium bisulphite, formic acid, and nucleotide analogues. When such agents are used, the mutagenesis is typically performed by incubating the DNA sequence encoding the parent enzyme to be mutagenized in the presence of the mutagenizing agent of choice under suitable conditions for the mutagenesis to take place, and selecting for mutated DNA having the desired properties.

When the mutagenesis is performed by the use of an oligonucleotide, the oligonucleotide may be doped or spiked with the three non-parent nucleotides during the synthesis of the 20 oligonucleotide at the positions which are to be changed. The doping or spiking may be done so that codons for unwanted amino acids are avoided. The doped or spiked oligonucleotide can be incorporated into the DNA encoding the phytase enzyme by any published technique, using e.g. PCR, LCR or any DNA polymerase 25 and ligase as deemed appropriate.

Preferably, the doping is carried out using "constant random doping", in which the percentage of wild-type and mutation in each position is predefined. Furthermore, the doping may be directed toward a preference for the introduction of certain nucleotides, and thereby a preference for the introduction of one or more specific amino acid residues. The

doping may be made, e.g., so as to allow for the introduction of 90% wild type and 10% mutations in each position. An additional consideration in the choice of a doping scheme is based on genetic as well as protein-structural constraints. The doping scheme may be made by using the DOPE program which, inter alia, ensures that introduction of stop codons is avoided.

When PCR-generated mutagenesis is used, either a chemically treated or non-treated gene encoding a model phytase is subjected to PCR under conditions that increase the mis10 incorporation of nucleotides (Deshler 1992; Leung et al., Technique, Vol.1, 1989, pp. 11-15).

A mutator strain of E. coli (Fowler et al., Molec. Gen. Genet., 133, 1974, pp. 179-191), S. cereviseae or any other microbial organism may be used for the random mutagenesis of the DNA encoding the model phytase by, e.g., transforming a plasmid containing the parent glycosylase into the mutator strain, growing the mutator strain with the plasmid and isolating the mutated plasmid from the mutator strain. The mutated plasmid may be subsequently transformed into the expression organism.

The DNA sequence to be mutagenized may be conveniently present in a genomic or cDNA library prepared from an organism expressing the model phytase. Alternatively, the DNA sequence may be present on a suitable vector such as a plasmid or a bacteriophage, which as such may be incubated with or otherwise exposed to the mutagenising agent. The DNA to be mutagenized may also be present in a host cell either by being integrated in the genome of said cell or by being present on a vector harboured in the cell. Finally, the DNA to be mutagenized may be in isolated form. It will be understood that the DNA sequence to be subjected to random mutagenesis is pre-ferably a cDNA or a genomic DNA sequence.

In some cases it may be convenient to amplify the mutated DNA sequence prior to performing the expression step b) or the screening step c). Such amplification may be performed in accordance with methods known in the art, the presently preferred method being PCR-generated amplification using oligonucleotide primers prepared on the basis of the DNA or amino acid sequence of the parent enzyme.

Subsequent to the incubation with or exposure to the mutagenising agent, the mutated DNA is expressed by culturing a 10 suitable host cell carrying the DNA sequence under conditions allowing expression to take place. The host cell used for this purpose may be one which has been transformed with the mutated DNA sequence, optionally present on a vector, or one which was carried the DNA sequence encoding the parent enzyme during the 15 mutagenesis treatment. Examples of suitable host cells are the following: gram positive bacteria such as Bacillus subtilis, Bacillus licheniformis, Bacillus lentus, Bacillus brevis, Bacillus stearothermophilus, Bacillus alkalophilus, Bacillus amyloliquefaciens, Bacillus coagulans, Bacillus circulans, 20 Bacillus lautus, Bacillus megaterium, Bacillus thuringiensis, Streptomyces lividans or Streptomyces murinus; and negative bacteria such as E. coli.

The mutated DNA sequence may further comprise a DNA sequence encoding functions permitting expression of the mutated 25 DNA sequence.

The random mutagenesis may be advantageously localised to a part of the model phytase in question using Localized random mutagenesis. This may, e.g., be advantageous when certain regions of the enzyme have been identified to be of particular importance for a given property of the enzyme, and when modified are expected to result in a variant having improved properties.

Such regions may normally be identified when the tertiary structure of the parent enzyme has been elucidated and related to the function of the enzyme.

The localized, or region-specific, random mutagenesis is conveniently performed by use of PCR generated mutagenesis techniques as described above or any other suitable technique known in the art. Alternatively, the DNA sequence encoding the part of the DNA sequence to be modified may be isolated, e.g., by insertion into a suitable vector, and said part may be subsequently subjected to mutagenesis by use of any of the mutagenesis methods discussed above.

For region-specific random mutagenesis with a view to amending e.g. the specific activity of a model phytase, codon positions corresponding to the following amino acid residues from the amino acid sequences set forth in Fig. 1 may appropriately be targeted:

Residues: 41-47, 68-80, 83-84, 115-118, 120-126, 128, 149-163, 184-185, 191-193, 198-201e, 202-203, 205, 235-236, 238-239, 242-243, 270-279, 285, 288, 332-343, 364-367, 369-375, 394.

Regions: 41-47, 68-80, 120-128, 149-163, 270-279, 332-343, 364-375.

The random mutagenesis may be carried out by the following steps:

- Select regions of interest for modification in the
 parent enzyme
 - 2. Decide on mutation sites and non-mutated sites in the selected region
- 3. Decide on which kind of mutations should be carried out, e.g. with respect to the desired stability and/or 30 performance of the variant to be constructed
 - Select structurally reasonable mutations

- 5. Adjust the residues selected by step 3 with regard to step 4.
- 6. Analyse by use of a suitable dope algorithm the nucleotide distribution.
- 7. If necessary, adjust the wanted residues to genetic code realism, e.g. taking into account constraints resulting from the genetic code, e.g. in order to avoid introduction of stop codons; the skilled person will be aware that some codon combinations cannot be used in practice and will need to be adapted
 - Make primers
 - 9. Perform random mutagenesis by use of the primers
 - 10. Select resulting phytase variants by screening for the desired improved properties.
- Suitable dope algorithms for use in step 6 are well known in the art. One such algorithm is described by Tomandl, D. et al., 1997, Journal of Computer-Aided Molecular Design 11:29-38. Another algorithm is DOPE (Jensen, LJ, Andersen, KV, Svendsen, A, and Kretzschmar, T (1998) Nucleic Acids Research 26:697-702).
- A DNA sequence encoding a model phytase or a phytase variant of the invention can be expressed using an expression vector, a recombinant expression vector, which typically includes control sequences encoding a promoter, operator, ribosome binding site, translation initiation signal, and, optionally, a repressor gene or various activator genes.

The recombinant expression vector may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, e.g. a plasmid, a bacteriophage or an extra-chromosomal element. Alternatively, the vector may

be one which, when introduced into a host cell, is integrated into the host cell genome and replicated together with the chromosome(s) into which it has been integrated.

In the vector, the DNA sequence should be operably connected to a suitable promoter sequence. The promoter may be any DNA sequence which shows transcriptional activity in the host cell of choice and may be derived from genes encoding proteins either homologous or heterologous to the host cell. An example of a suitable promoter for directing the transcription of the DNA sequence encoding a phytase variant of the invention, especially in a bacterial host, is the promoter of the lac operon of E.coli. For transcription in a fungal host, examples of useful promoters are those derived from the gene encoding A. oryzae TAKA amylase.

The expression vector of the invention may also comprise a suitable transcription terminator and, in eukaryotes, polyadenylation sequences operably connected to the DNA sequence encoding the phytase variant of the invention. Termination and polyadenylation sequences may suitably be derived from the same sources as the promoter.

The vector may further comprise a DNA sequence enabling the vector to replicate in the host cell in question. Examples of such sequences are the origins of replication of plasmids pUC19, pACYC177, pUB110, pE194, pAMB1 and pIJ702.

gene the product of which complements a defect in the host cell, such as the dal genes from B. subtilis or B. licheniformis, or one which confers antibiotic resistance such as ampicillin resistance. Furthermore, the vector may comprise Aspergillus selection markers such as amdS, argB, niaD and sC, or the

selection may be accomplished by co-transformation, e.g. as described in WO 91/17243.

The procedures used to ligate the DNA construct of the invention encoding a phytase variant, the promoter, terminator 5 and other elements, respectively, and to insert them into suitable vectors containing the information necessary for replication, are well known to persons skilled in the art (cf., for instance, Sambrook et al. (1989)).

The cell of the invention, either comprising a DNA construct or an expression vector of the invention as defined above, is advantageously used as a host cell in the recombinant production of a phytase variant of the invention. The cell may be transformed with the DNA construct of the invention encoding the variant, conveniently by integrating the DNA construct (in one or more copies) in the host chromosome. This integration is generally considered to be an advantage as the DNA sequence is more likely to be stably maintained in the cell. Integration of the DNA constructs into the host chromosome may be performed according to conventional methods, e.g. by homologous or heterologous recombination. Alternatively, the cell may be transformed with an expression vector as described above in connection with the different types of host cells.

An isolated DNA molecule or, alternatively, a "cloned DNA sequence" "a DNA construct," "a DNA segment" or "an isolated DNA sequence" refers to a DNA molecule or sequence which can be cloned in accordance with standard cloning procedures used in genetic engineering to relocate the DNA segment from its natural location to a different site where it will be replicated. The term refers generally to a nucleic acid sequence which is essentially free of other nucleic acid sequences, e.g., at least about 20% pure, preferably at least about 40% pure, more

preferably about 60% pure, even more preferably about 80% pure, most preferably about 90% pure, and even most preferably about 95% pure, as determined by agarose gel electrophoresis. The cloning procedures may involve excision and isolation of a 5 desired nucleic acid fragment comprising the nucleic acid sequence encoding the polypeptide, insertion of the fragment into a vector molecule, and incorporation of the recombinant vector into a host cell where multiple copies or clones of the nucleic acid sequence will be replicated. The nucleic acid sequence may be of genomic, cDNA, RNA, semisynthetic, synthetic origin, or any combinations thereof.

The term "vector" is intended to include such terms/objects as "nucleic acid constructs," "DNA constructs," expression vectors" or "recombinant vectors."

15 The nucleic acid construct comprises a nucleic acid sequence of the present invention operably linked to one or more control sequences capable of directing the expression of the coding sequence in a suitable host cell under conditions compatible with the control sequences.

"Nucleic acid construct" is defined herein as a nucleic acid molecule, either single or double-stranded, which is isolated from a naturally occurring gene or which has been modified to contain segments of nucleic acid which are combined and juxtaposed in a manner which would not otherwise exist in nature.

The term nucleic acid construct may be synonymous with the term expression cassette when the nucleic acid construct contains all the control sequences required for expression of a coding sequence of the present invention.

The term "coding sequence" as defined herein primarily comprises a sequence which is transcribed into mRNA and

translated into a polypeptide of the present invention when placed under the control of the above mentioned control sequences. The boundaries of the coding sequence are generally determined by a translation start codon ATG at the 5'-terminus and a translation stop codon at the 3'-terminus. A coding sequence can include, but is not limited to, DNA, cDNA, and recombinant nucleic acid sequences.

The term "control sequences" is defined herein to include all components which are necessary or advantageous for 10 expression of the coding sequence of the nucleic acid sequence. Each control sequence may be native or foreign to the nucleic acid sequence encoding the polypeptide. Such control sequences include, but are not limited to, a leader, a polyadenylation sequence, a propeptide sequence, a promoter, a signal sequence, 15 and a transcription terminator. At a minimum, the control sequences include a promoter, and transcriptional and translational stop signals. The control sequences may be provided with linkers for the purpose of introducing specific restriction sites facilitating ligation of the control sequences 20 with the coding region of the nucleic acid sequence encoding a polypeptide.

A "host cell" or "recombinant host cell" encompasses any progeny of a parent cell which is not identical to the parent cell due to mutations that occur during replication.

The cell is preferably transformed with a vector comprising a nucleic acid sequence of the invention followed by integration of the vector into the host chromosome.

"Transformation" means introducing a vector comprising a nucleic acid sequence of the present invention into a host cell so that the vector is maintained as a chromosomal integrant or as a self-replicating extra-chromosomal vector. Integration is

generally considered to be an advantage as the nucleic acid sequence is more likely to be stably maintained in the cell. Integration of the vector into the host chromosome may occur by homologous or non-homologous recombination as described above.

5 The host cell may be a unicellular microorganism, e.g., a prokaryote, or a non-unicellular microorganism, e.g., a eukaryote. Examples of a eukaryote cell is a mammalian cell, an insect cell, a plant cell or a fungal cell. Useful mammalian cells include Chinese hamster ovary (CHO) cells, HeLa cells, baby hamster kidney (BHK) cells, COS cells, or any number of other immortalized cell lines available, e.g., from the American Type Culture Collection.

In a preferred embodiment, the host cell is a fungal cell.

Fungal cells may be transformed by a process involving 15 protoplast formation, transformation of the protoplasts, and regeneration of the cell wall in a manner known per se.

The present invention also relates to a transgenic plant, plant part, such as a plant seed, or plant cell, which has been transformed with a DNA sequence encoding the phytase of the invention so as to express or produce this enzyme. Also compositions and uses of such plant or plant part are within the scope of the invention, especially its use as feed and food or additives therefore, along the lines of the present use and food/feed claims.

25 The transgenic plant can be dicotyledonous or monocotyledonous, for short a dicot or a monocot. Of primary interest are such plants which are potential food or feed components and which comprise phytic acid. A normal phytic acid level of feed components is 0.1-100 g/kg, or more usually 0.5-50 g/kg, most usually 0.5-20 g/kg. Examples of monocot plants are grasses, such as meadow grass (blue grass, Poa), forage grass

such as festuca, lolium, temperate grass, such as Agrostis, and cereals, e.g. wheat, oats, rye, barley, rice, sorghum and maize (corn).

Examples of dicot plants are legumes, such as lupins, pea, 5 bean and soybean, and cruciferous (family Brassicaceae), such as cauliflower, oil seed rape and the closely related model organism Arabidopsis thaliana.

Such transgenic plant etc. is capable of degrading its own phytic acid, and accordingly the need for adding such enzymes to food or feed comprising such plants is alleviated. Preferably, the plant or plant part, e.g. the seeds, are ground or milled, and possibly also soaked before being added to the food or feed or before the use, e.g. intake, thereof, with a view to adapting the speed of the enzymatic degradation to the actual use.

If desired, the plant produced enzyme can also be recovered from the plant. In certain cases the recovery from the plant is to be preferred with a view to securing a heat stable formulation in a potential subsequent pelleting process.

Examples of plant parts are stem, callus, leaves, root, 20 fruits, seeds, tubers etc. But also any plant tissue is included in this definition.

Any plant cell, whatever the tissue origin, is included in the definition of plant cells above.

Also included within the scope of the invention are the 25 progeny of such plants, plant parts and plant cells.

The skilled man will know how to construct a DNA expression construct for insertion into the plant in question, paying regard i.a. to whether the enzyme should be excreted in a tissue specific way. Of relevance for this evaluation is the stability (pH-stability, degradability by endogenous proteases etc.) of the phytase in the expression compartments of the

plant. He will also be able to select appropriate regulatory sequences such as promoter and terminator sequences, and signal or transit sequences if required (Tague et al, Plant, Phys., 86, 506, 1988).

The plant, plant part etc. can be transformed with this DNA construct using any known method. An example of such method is the transformation by a viral or bacterial vector such as bacterial species of the genus Agrobacterium genetically engineered to comprise the gene encoding the phytase of the invention. Also methods of directly introducing the phytase DNA into the plant cell or plant tissue are known in the art, e.g. micro injection and electroporation (Gasser et al, Science, 244, 1293; Potrykus, Bio/Techn. 8, 535, 1990; Shimamoto et al, Nature, 338, 274, 1989).

Following the transformation, the transformants are screened using any method known to the skilled man, following which they are regenerated into whole plants.

These plants etc. as well as their progeny then carry the phytase encoding DNA as a part of their genetic equipment.

In general, reference is made to WO 9114782A and WO 9114772A.

Agrobacterium tumefaciens mediated gene transfer is the method of choice for generating transgenic dicots (for review Hooykas & Schilperoort, 1992. Plant Mol. Biol. 19: 15-38), 25 however it can also be used for transforming monocots. Due to host range limitations it is generally not possible to transform monocots with the help of A. tumefaciens. Here, other methods have to be employed. The method of choice for generating transgenic monocots is particle bombardment (microscopic gold or tungsten particles coated with the transforming DNA) of embryonic calli or developing embryos (Christou, 1992. Plant J.

2: 275-281; Shimamoto, 1994. Curr. Opin. Biotechnol. 5: 158-162; Vasil et al., 1992. Bio/Technology 10: 667-674).

Also other systems for the delivery of free DNA into these plants, including viral vectors (Joshi & Joshi, 1991. FEBS Lett. 5 281: 1-8), protoplast transformation via polyethylene glycol or electroporation (for review see Potyrkus, 1991. Annu. Rev. Plant Physiol. Plant Mol. Biol. 42: 205-225), microinjection of DNA into mesophyll protoplasts (Crossway et al., 1986. Mol. Gen. Genet. 202: 79-85), and macroinjection of DNA into young floral tillers of cereal plants (de la Pena et al., 1987. Nature 325: 274-276) are preferred methods.

In general, the cDNA or gene encoding the phytase variant of the invention is placed in an expression cassette (e.g. Pietrzak et al., 1986. Nucleic Acids Res. 14: 15 consisting of a suitable promoter active in the target plant and a suitable terminator (termination of transcription). This cassette (of course including a suitable selection marker, see below) will be transformed into the plant as such in case of monocots via particle bombardment. In case of dicots 20 expression cassette is placed first into a suitable vector providing the T-DNA borders and a suitable selection marker which in turn are transformed into Agrobacterium tumefaciens. Dicots will be transformed via the Agrobacterium harbouring the expression cassette and selection marker flanked by T-DNA 25 following standard protocols (e.g. Akama et al., 1992. Plant Cell Reports 12: 7-11). The transfer of T-DNA from Agrobacterium to the Plant cell has been recently reviewed (Zupan & Zambryski, Plant Physiol. 107: 1041-1047). Vectors for plant transformation via Agrobacterium are commercially available or 30 can be obtained from many labs that construct such vectors (e.g. Deblaere et al., 1985. Nucleic Acids Res. 13: 4777-4788; for

review see Klee et al., 1987. Annu. Rev. Plant Physiol. 38: 467-486).

Available plant promoters: Depending on the process under manipulation, organ- and/or cell-specific expression as well as 5 appropriate developmental and environmental control required. For instance, it is desirable to express a phytase cDNA in maize endosperm etc. The most commonly used promoter has been the constitutive 35S-CaMV promoter Franck et al., 1980. Cell 21: 285-294). Expression will be more or less 10 throughout the whole plant. This promoter has been successfully to engineer herbicide- and pathogen-resistant plants (for review see Stitt & Sonnewald, 1995. Annu. Rev. Plant Physiol. Plant Mol. Biol. 46: 341-368). Organ-specific promoters have been reported for storage sink tissues such as seeds, 15 potato tubers, and fruits (Edwards & Coruzzi, 1990. Annu. Rev. Genet. 24: 275-303), and for metabolic sink tissues such as meristems (Ito et al., 1994. Plant Mol. Biol. 24: 863-878).

The medium used to culture the transformed host cells may be any conventional medium suitable for growing the host cells in question. The expressed phytase may conveniently be secreted into the culture medium and may be recovered therefrom by well-known procedures including separating the cells from the medium by centrifugation or filtration, precipitating proteinaceous com-ponents of the medium by means of a salt such as ammonium sulphate, followed by chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

Preferred host cells are a strain of Fusarium, Hansenula, Trichoderma or Aspergillus, in particular a strain of Fusarium graminearum, Fusarium venenatum, Fusarium cerealis, Fusarium sp. having the identifying characteristic of Fusarium ATCC 20334, as further described in PCT/US/95/07743, Hansenula polymorpha,

Trichoderma harzianum or Trichoderma reesei, Aspergillus niger or Aspergillus oryzae.

References for expression in Hansenula polymorpha: Gellissen, G., Piontek, M., Dahlems, U., Jenzelewski, V., 5 Gavagan, J.E., DiCosimo, R., Anton, D.I. & Janowicz, Z.A. (1996) Recombinant Hansenula polymorpha as a biocatalyst: coexpression of the spinach glycolate oxidase (GO) and the S. cerevisiae catalase T (CTT1) gene. Appl. Microbiol. Biotechnol. 46, 46-54.

Some more specific uses of the phytase variants according 10 to the invention appear from PCT/DK97/00568, the last pages of the detailed description of the invention section.

In a preferred embodiment, the phytase variant of the invention is essentially free of other non-phytase polypeptides, e.g., at least about 20% pure, preferably at least about 40% pure, more preferably about 60% pure, even more preferably about 80% pure, most preferably about 90% pure, and even most preferably about 95% pure, as determined by SDS-PAGE. Sometimes such polypeptide is alternatively referred to as a "purified" and/or "isolated" phytase.

20 A phytase polypeptide which comprises a phytase variant of the invention includes fused polypeptides or cleavable fusion polypeptides in which another polypeptide is fused at the Nterminus or the C-terminus of the polypeptide or fragment thereof. A fused polypeptide is produced by fusing a nucleic 25 acid sequence (or a portion thereof) encoding another polypeptide to a nucleic acid sequence (or a portion thereof) encoding a phytase variant of the present invention. Techniques for producing fusion polypeptides are known in the art, and include, ligating the coding sequences encoding the polypeptides 30 so that they are in frame and that expression of the fused

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polypeptide is under control of the same promoter(s) and terminator.

A "feed" and a "food," respectively, means any natural or artificial diet, meal or the like or components of such meals intended or suitable for being eaten, taken in, digested, by an animal and a human being, respectively.

The phytase variant of the invention may exert its effect in vitro or in vivo, i.e. before intake or in the stomach of the individual, respectively. Also a combined action is possible.

A phytase composition according to the invention always comprises at least one phytase of the invention.

Generally, phytase compositions are liquid or dry.

Liquid compositions need not contain anything more than the phytase enzyme, preferably in a highly purified form.

15 Usually, however, a stabilizer such as glycerol, sorbitol or mono propylen glycol is also added. The liquid composition may also comprise other additives, such as salts, sugars, preservatives, pH-adjusting agents, proteins, phytate (a phytase substrate). Typical liquid compositions are aqueous or oil-based slurries. The liquid compositions can be added to a food or feed after an optional pelleting thereof.

Dry compositions may be spray-dried compositions, in which case the composition need not contain anything more than the enzyme in a dry form. Usually, however, dry compositions are so-called granulates which may readily be mixed with e.g. food or feed components, or more preferably, form a component of a premix. The particle size of the enzyme granulates preferably is compatible with that of the other components of the mixture. This provides a safe and convenient means of incorporating on enzymes into e.g. animal feed.

Agglomeration granulates are prepared using agglomeration technique in a high shear mixer (e.g. Lödige) during which a filler material and the enzyme are co-agglomerated to form granules. Absorption granulates are prepared by having cores of a carrier material to absorb/be coated by the enzyme.

Typical filler materials are salts such as disodium sulphate. Other fillers are kaolin, talc, magnesium aluminium silicate and cellulose fibres. Optionally, binders such as dextrins are also included in agglomeration granulates.

Typical carrier materials are starch, e.g. in the form of cassava, corn, potato, rice and wheat. Salts may also be used.

Optionally, the granulates are coated with a coating mixture. Such mixture comprises coating agents, preferably hydrophobic coating agents, such as hydrogenated palm oil and beef tallow, and if desired other additives, such as calcium carbonate or kaolin.

Additionally, phytase compositions may contain other substituents such as colouring agents, aroma compounds, stabilizers, vitamins, minerals, other feed or food enhancing enzymes, i.e. enzymes that enhances the nutritional properties of feed/food, etc. This is so in particular for the so-called pre-mixes.

A "food or feed additive" is an essentially pure compound or a multi component composition intended for or suitable for 25 being added to food or feed. In particular it is a substance which by its intended use is becoming a component of a food or feed product or affects any characteristics of a food or feed product. It is composed as indicated for phytase compositions above. A typical additive usually comprises one or more 30 compounds such as vitamins, minerals or feed enhancing enzymes and suitable carriers and/or excipients.

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In a preferred embodiment, the phytase compositions of the invention additionally comprises an effective amount of one or more feed enhancing enzymes, in particular feed enhancing enzymes selected from the group consisting of α -galactosidases, 5 β -galactosidases, in particular lactases, other phytases, β in particular endo- β -1,4-glucanases and endo- β glucanases, 1,3(4)-glucanases, cellulases, xylosidases, galactanases, arabinogalactan endo-1, $4-\beta$ -galactosidases and particular arabinogalactan endo-1,3- β -galactosidases, endoglucanases, in 10 particular endo-1,2- β -glucanase, endo-1,3- α -glucanase, and endo-1,3-β-glucanase, pectin degrading enzymes, in particular pectinases, pectinesterases, pectin lyases, polygalacturonases, arabinanases, rhamnogalacturonases, rhamnogalacturonan acetyl esterases, rhamnogalacturonan- α -rhamnosidase, pectate lyases, 15 and α -galacturonisidases, mannanases, β -mannosidases, mannan acetyl esterases, xylan acetyl esterases, proteases, xylanases, and lipolytic enzymes such as lipases, arabinoxylanases phospholipases and cutinases.

The animal feed additive of the invention is supplemented to the mono-gastric animal before or simultaneously with the diet. Preferably, the animal feed additive of the invention is supplemented to the mono-gastric animal simultaneously with the diet. In a more preferred embodiment, the animal feed additive is added to the diet in the form of a granulate or a stabilized liquid.

An effective amount of phytase in food or feed is from about 10-20.000; preferably from about 10 to 15.000, more preferably from about 10 to 10.000, in particular from about 100 to 5.000, especially from about 100 to about 2.000 FYT/kg feed or food.

Examples of other specific uses of the phytase of the invention is in soy processing and in the manufacture of inositol or derivatives thereof.

The invention also relates to a method for reducing 5 phytate levels in animal manure, wherein the animal is fed a feed comprising an effective amount of the phytase of the invention.

Also comprised in this invention is the use of a phytase of the invention during the preparation of food or feed preparations or additives, i.e. the phytase exerts its phytase activity during the manufacture only and is not active in the final food or feed product. This aspect is relevant for instance in dough making and baking.

The invention relates to a phytase variant which, when aligned according to Fig. 1, is amended as compared to a model phytase in at least one of the following positions, using the position numbering corresponding to P_lycii:

24; 27; 31; 33; 39; 40; 41; 42; 43; 44; 45; 46; 47; 49; 51; 56; 58; 59; 61; 62; 68; 69; 70; 71; 72; 73; 74; 75; 76; 77; 78; 79;

20 80; 81; 82; 83; 84; 88; 90; 102; 115; 116; 117; 118; 119; 120; 121; 122; 123; 124; 125; 126; 127; 128; 132; 143; 148; 149; 150; 151; 152; 153; 154; 155; 156; 157; 158; 159; 160; 161; 162; 163; 170f; 170g; 171; 172; 173; 184; 185; 186; 187; 187a; 190; 191; 192; 193; 194; 195; 198; 199; 200; 201; 201a; 201b; 201c; 201d;

25 201e; 201f; 202; 203; 203a; 204; 205; 211; 215; 220; 223; 228; 232; 233; 234; 235; 236; 237; 238; 239; 242; 243; 244; 246; 251e; 253; 256; 260; 264; 265; 267; 270; 271; 272; 273; 274; 275; 276; 277; 278; 279; 280; 283; 285; 287; 288; 292; 293; 302; 304; 332; 333; 334; 335; 336; 337; 338; 339; 340; 341; 342; 343; 30 348; 349; 352; 360; 362; 364; 365; 366; 367; 368; 369; 370; 371;

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372; 373; 374; 375; 376; 383k; 387; 393; 394; 396; 404; 409; 411; 412; 413; 417; 421; 431.

From these variants we expect amended characteristics, preferably amended activity characteristics. In fact, 5 several variants such amended characteristics have already been shown (see the experimental part). Like above, "amended" means "Amended model phytase. the compared to characteristics" means amended in at least one phytase activity related respect, such as (non-exclusive list): pH stability, 10 temperature stability, pH profile, temperature profile, specific activity (in particular in relation to pH and temperature), substrate specificity, substrate cleavage pattern, substrate binding, position specificity, the velocity and level of release of phosphate from corn, reaction rate, phytate degradation 15 rate), end level of released phosphate reached.

Preferred amended activity characteristics are amended specific activity, preferably increased, and preferably increased at a pH of 3, 4, 5, or 6; amended pH or temperature profile; and/or amended, preferably increased, thermostability, e.g. of an increased melting temperature as measured using DSC.

Preferred phytase variants are: Phytase variants which, when aligned according to Fig. 1, are amended as compared to a model phytase in at least one of the following positions, using the position numbering corresponding to P_lycii:

- 25 43; 44; 47; 51; 58; 62; 78; 80; 83; 88; 90; 102; 143; 148; 153; 154; 186; 187a; 195; 198; 201e; 204; 205; 211; 215; 220; 242; 244; 251e; 260; 264; 265; 267; 270; 273; 278; 302; 336; 337; 339; 352; 365; 373; 383k; 404; 417.
- The following variants of A_fumigatus constitute a 30 subgroup: Q43L; Q270L; G273D,K; N336S; A205E; Y278H; Q43L+Q270L;

Q43L+Q270L+G273D; Q43L+Q270L+G273D+N336S; G273K+A205E; G273K+A205E+Y278H (see EP 0897010).

Generally, variants of the invention can be deduced or identified as follows: Looking at the alignment according to 5 Fig. 1, comparing two sequences, one of which is a model phytase with improved properties, identifying amino acid differences in relevant positions/areas, and transferring (substituting with) from the model to the other phytase sequence the amino acid in a relevant position.

- The invention also relates to a process for preparing a phytase variant which includes the above method, and further includes the deducement and synthesis of the corresponding DNA sequence, the transformation of a host cell, the cultivation of the host cell and the recovery of the phytase variant.
- Relevant positions/areas include those mentioned below in relation to important phytase activity characteristics such as specific activity, thermostability, pH activity/stability.

The present invention also relates to phytase variants (varied according to a model phytase as defined herein) which 20 are obtainable, preferably obtained, by the process outlined above and which are expected to exhibit an amended characteristic/property, preferably does exhibit such amended characteristic, e.g. an improved specific activity.

At least the basidiomycete model phytases P_lycii and 25 T_pubescens exhibit a high specific activity (as determined using the method of Example 2 herein).

This is an example of a desired property which can be transferred to other phytases, e.g. the other phytases listed in Fig. 1, in particular to the A_pediades and the ascomycete 30 phytases such as A_fumigatus, A-ficuum, consphyA, by a deducement process such as the one mentioned above.

Thus, amended specific activity, in particular an improved specific activity, in particular at low pH and/or high temperature, is expected from variants, which have been amended in relevant areas, viz. (i) in the amino acid residues which point into the active site cleft; or (ii) in the amino acid residues in the close neighbourhood of these active site residues. Preferably, close neighbourhood means within 10Å from the active site residues.

From the pdb file 1IHP (Brookhaven Database entry of 18.03.98 re 1IHP, Structure of Phosphomonoesterase, D.Kostrewa; or as published in Nature Structural Biology, 4, 1997, p. 185-190), active site regions can be identified, using the program INSIGHTII from Molecular Simulations MSI, San Diego, California, and using the subset command, an "active site shell" can be defined comprising those amino acid residues which lie close to the catalytic residues, defined as H59, D339 and R58 in A. ficuum phytase (corresponding to Peniophora numbers H71, D335 and R70, respectively). An "active site shell(10Å)" comprises those residues which lie within 10Å from the above catalytic residues.

The residues within 10Å from H71 and D335 are the following (using Peniophora numbers): 41-47, 68-77, 115-118, 120-126, 128, 149-163, 185, 191-193, 199, 243, 270-271, 273-275, 277-279, 288, 332-343, 364-367, 369-375, 394 ("the active site shell(10Å)").

Preferably, a "substrate binding shell" can also be defined which comprises those residues which are in close proximity to the substrate binding site and which can therefore be expected to be in contact with the substrate.

This information can be deduced as described above, by docking a sugar analogue to phytin into the active site cleft

(the residues making up the surface of the active site). If a sugar without any phosphate groups is docked into the active site cleft, e.g. alpha-D-glucose (chair conformation, structure provided by the INSIGHTII program), using a fixed distance as 5 shown below, the residues pointing towards the active site cleft can be extracted using the subset command and using a distance of 10Å from the substrate analogue. Alternatively, the compound inositol-1,4,5-triphosphate (Brookhaven database file ldix. Inositol-1,4,5-triphosphate) can be docked into the active site 10 cleft. This compound and glucose, however, are more or less superimposable.

The distances in Ångström (Å) are: From oxygen atom in position 6 of the alpha-D-glucose to

atom ND1 of H59: 5.84 15 atom NH2 of R58: 6.77 atom NH2 of R142: 5.09 atom ND2 of N340: 3.00 atom ND1 of H59: 7.76 atom NH2 of R58:

(the Peniophora numbers of the above residues are: H71, 20 R70, R155, N336, H71 and R70, respectively).

8.58.

In this way, the residues in contact with the substrate are identified as follows (Peniophora numbers): 43-44; 70-80; 83-84; 115; 153; 155-156; 184; 191-192; 198-202; 205; 235; 238; 25 242; 270; 272-273; 275-277; 332-336; 338; 369; 371 substrate binding shell(10Å)").

Variants being amended in one or more of (1) the active site shell or (2) the substrate binding shell, are strongly expected to have an amended specific activity. This leads to the 30 following joint grouping of positions (still Peniophora numbers and 10Å shells): 41-47, 68-80, 83-84, 115-118, 120-126, 128,

149-163, 184-185, 191-193, 198-201e, 202-203, 205, 235-236, 238-239, 242-243, 270-279, 285, 288, 332-343, 364-367, 369-375, 394.

Preferably, the active site shell and the substrate binding shell are defined as described above using the basidiomycete model phytases of Fig. 1, the Peniophora phytase being a preferred model. A deducement of corresponding variants of other model phytases is possible using the alignment of Fig. 1.

In a preferred embodiment, a distance of 5Å is used in the subset command, thus defining active site and substrate binding shells of a more limited size, e.g. an active site shell comprising the residues 43-44, 69-74, 117, 125, 155-156, 159, 274, 332-340, 370-374 (5Å from H71 and D335), "active site shell(5Å)".

Generally the active site shell and substrate binding shell regions form the basis for selecting random mutagenesis regions. Examples of preferred random mutagenesis regions are

regions 69-74, 332-340, 370-374, doping to be added (a 5Å approach); and

regions 57-62, 142-146, 337-343, doping to be added (a 10Å approach).

It is presently contemplated that any amendment in either of these positions will lead to a phytase of amended characteristics, e.g. of an amended specific activity.

25 The above expression "any amendment in either of the positions" is considered fully equivalent to listing each position and each substitution, e.g. as follows for the above sub-group 41-47:

41A, C, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, Y;

30 42A,C,D,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y; 43A,C,D,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y; 44A, C, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, Y; 45A, C, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, Y; 46A, C, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, Y; 47A, C, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, Y.

In a preferred embodiment, amended specific activity is expected from the following variants:

42S,G; 43A,C,D,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y; 45D,S; 47Y,F; 51E,A; 75W,F; 78S,D; 79G; 80K,A; 83I,Q; 84Q,V; 116S; 118V,L; 119E; 120L; 122A; 123N,T; 125S; 126H,S; 127Q,E; 128A,T; 10 151A,S; 152G; 153D,Y; 154Q,D,G; 157V; 158D,A; 159T; 160A,S; 161T,N; 162N; 163W; 184Q,S; 186A,E; 198A,N; 200G,V; 201D; deletions of one or more of 201a, 201b, 201c, 201d, 201e, 201f preferably all; 202S; 205Q,E; 235Y,L; 238L,M; 242P; 270Y,A,L; 271D; 273D,K; 275F,Y; 278T,H; 332F; 336S; 337T,Q; 339V; 340P,A; 15 343A,S; 364W,F; 365V,L; 366D,V; 367K; 368K; 369I,L; 370V; 373S; 374A; 375H; 376M; 393V.

Particularly preferred variants are the following: 78S; 79G; 80A; 83I,Q; 84Q,V; 198A,N; 200G,V; 201D; deletions in one or more of 201a, 201b, 201c, 201d, 201e, 201f - preferably all 20 deletions; 202S; 205Q,E; 235Y,L; 238L,M; 242P, 273D; 275F,Y.

Other particularly preferred variants are the following: 43A,C,D,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y; in particular 43M,P; 75W,F; 80K; 153D; 184Q,S; 270Y,A; 332F; 369I,L.

The following variants are especially preferred: 25 43L,G,N,V,A,I,T; 78D; 153Y; 154G; 270L; 273D,K. Double and triple variants (43L/270L); (43L/270L/273D); (43L/78D) and (43L/153Y/154G) are also especially preferred. Other preferred variants are 205E; 278H; 336S.

These especially preferred single, double and triple 30 variants are preferably variants of model phytases which can be

aligned to Fig.1, in particular variants of the specific model phytases listed in Fig. 1.

At least consphyA is known to have a high thermostability. Still further, the thermostability of P_lycii is rather high.

This is an example of a desired property which can be transferred to other phytases, e.g. the other phytases listed in Fig. 1, in particular to the basidiomycete phytases such as P_lycii and A_pediades, by a deducement process such as the one mentioned above.

Amended thermostability, in particular improved thermostability, is expected on this background from the following variants:

39H,S; 40L,N; 43P; 47Y,F; 49P; 51E,A; 56P; 58D; 61R; 62V; 80K; 83A; 84Y; 172P; 184P; 195T; 198A; 204V; 211L; 223D; 236Y; 15 242P; 246V; 253P; 264R; 265Q; 280A,P; 283P; 287A; 292F,Y; 293A; 302R; 304P; 337S; 348Y; 387P; 396R; 409R; 411K; 412R; 417E; 421F,Y.

The following variants of amended thermostability are particularly preferred: 39S; 40N; 47Y,F; 51A; 83A; 195T; 204V; 20 211L; 242P; 265A.

Further variants of amended thermostability are the following: 42G; 43T,L,G; 44N; 58K,A; 59G; 62I; 69Q; 75F; 78D; 79G; 80A; 81A,G; 82T; 83K,R; 84I; 88I; 90R,A; 102Y; 115N; 118V; 122A; 123Q,N; 125M,S; 126V,S; 127N,Q; 128S,A; 143N,K; 148V,I; 154S; 158D; 170fH; 170gA; 171T,N; 172N; 173W, 184S; 186A; 187A; 187aS; 193S; 195V,L; 198V; 201E; 201eT; 202A, 203aT; 204A; 211V, 215P,A; 220L,N; 223H; 228N; 232T; 322E; 235T; 236N; 242S; 244D; 251eQ,E; 256D; 264I; 260A,H; 265A; 267D; 270G; 271D; 273K,D; 278T,H; 287T; 293V; 302H; 337T,G; 338I; 339V,I; 340A; 352K; 365A,S; 366S; 367A; 369L; 373S,A; 374S; 376M; 383kE,Q; 404G,A; 411T; 417R; 431E.

Other concepts of the invention, which can be expected to impart an improved thermostability to a phytase, are as follows - considering the 1IHP structure previously referred to and transferring via an alignment according to Fig. 1 as outlined 5 herein:

- (A) Introduction of prolin residues in spatial positions where the prolin special dihedral angles are satisfied and the hydrogen bonding network are not hampered and no steric clashes are observed.
- 10 (B) Filling up holes: By substitution for bigger residues in internal cavities an improvement in stability can often be obtained.
 - (C) Cystin bridge: Cystin bridges will often make the proteins more rigid and increase the energy of unfolding.
- Further variants from which amended thermostability is expected according to these concepts of (A) to (C) are: 27P, 31Y, 132F, 132I, 132L, 184P, 186P, 190P, 280P, 343F, 343I, 343L, 349P, 362P and (33C and 24C).

Concept (A): 27P, 184P, 186P, 190P, 349P, 362P.

20 Concept (B): 343F,I,L; 31Y; 132F,I,L; 273F.

Concept (C): 33C/24C.

Amended pH activity or stability, preferably stability, in particular at low pH, in particular improved, is another desired property which can be transferred by aligning according to Fig. 25 1 and transferring from models of improved pH profiles to other phytases - as outlined above.

Other concepts of the invention, which can be expected to impart an improved stability at low pH to a phytase, are as follows - considering the 1IHP structure previously referred to and transferring via an alignment according to Fig. 1 as outlined herein:

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- Surface charges: Better distribution at low pH, to avoid (D) cluster of negative or positive, and to avoid too close same charged residues.
- Prevent deamidation: Surface exposed Q or N in close (E) 5 contact to negative charged residues.

Phytase variants having improved pH stability/activity at low pH are expected to be: 39H; 39Q; 80A; 203R; 271N; 51R; 154S; 185S; 194S; 194T; 288L; 288F; 360R; 173Q,S; 204Q,S; 303K,S; 81Q,E.

Concept (D): 203R, 271N, 51R, 185S, 360R; 173Q,S; 204Q,S; 10 303K,S; 81Q,E.

Concept (E): 154S; 194S,T; 288L,I,F.

A preferred model phytase for these concepts of (D) and (E) is P lycii.

Experimentally proven to have a lowered pH optimum is: 15 Variant 80A of ascomycete phytases, in particular of A_fumigatus and consphyA.

Especially preferred single, double and triple variants are 43L; (43L/270L) and (43L/270L/273D). These variants have a 20 changed pH profile. They are preferably variants of the specific model phytases listed in Fig. 1.

For all preferred variants listed above:

the stability is preferably amended at high temperature, viz. in the temperature range of 50-100°C, in particular 60-25 90°C, more preferably in the range of 70-90°C;

the activity is preferably amended in a temperature range relevant for the use in the gastro-intestinal system of animals, e.g. 30-40°C, more preferably 32-38°C, most preferably in the range of 35-38°C;

the stability is preferably amended at low pH, viz. in the 30 pH range of pH 1.5-7, preferably 2-6, more preferably 3-5;

the activity is preferably amended in the pH range of pH 1.5-5.5, more preferably at pH 2.5-4.5, still more preferably 3-5.5

Tests for amended phytase characteristics, such as those 5 mentioned above, are well known in the art and any such test can be used to compare the performance of the phytase variants with the phytase models.

A preferred test for specific activity is given in Example 2. Preferred tests for pH and temperature activity and stability 10 are given in Example 3. An even more preferred test for thermal stability is the DSC method of Example 4.

WO 98/28409 discloses tests for various other parameters, too, such as position specificity. All the tests of WO 98/28409 are preferred tests.

Generally, of course all these tests can be conducted at desired pH values and temperatures.

In the dependent claims, some preferred phytase variants based on five of the thirteen herein specifically disclosed model phytases are specified.

In an analogous way other preferred variants based on the remaining eight specifically disclosed model phytases can easily be deduced by combining the suggested amendments with each of the corresponding sequences of Fig. 1. These preferred variants are specifically included in the present invention, and they are easily deducemed, viz. the following:

Variants of a model phytase derived from Paxillus, preferably Paxillus involutus, preferably derived from strain CBS 100231, preferably variants of P_involtus-A1, the sequence of which is shown at Fig. 2, said variants comprising at least one of the following amendments:

()24C; T27P; F31Y; I33C; R39H,S,Q; N40L; S42G;

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P43A, C, D, E, F, G, H, I, K, L, M, N, Q, R, S, T, V, W, Y; Y44N; S45D; Y47F; A51E,R; A58D;K; Q61R; I62V; F75W; S78D; A80K; T81Q,E,G,A; R83A, I, Q, K; I84Y, Q, V; L88I; K90R, A; F102Y; S115N; D116S; V118L; P119E; F120L; A123N,T,Q; S125M; F126H,S,V; D127Q,E,N; A128T,S; 5 A132F,I,L; I148V; D151A,S; S153D,Y; D154Q,S,G; D158A; S159T; A160S; T161N; ()170fH; ()170gA; S171N; H172P; N173Q,S; P184Q,S; Q185S; T186A,E,P; G187A; ()187aS; T190P,A; D193S; N194S,T; M195T, V, L; A198N, V; G200V; D201E; ()201eT; S202A; D203R, K, S; P203aV,T; Q204E,S,A,V; V205E; V211L; S215A,I; L220N; A223D,H; 10 D233E; F235Y,L,T; N236Y; L237F; V238L,M; A242P,S; M244D; ()251eE,Q; D253P; T256D; P260A,H; E264R,I; A265Q; A267D; G270Y, A, L; D271N; D273K; F275Y; T278H; Y280A, P; E283P; V287A, T; Q288L,I,F; Y292F; V293A; N302R,H; A304P; N336S; L337T,Q,S,G; M 3381; V3391; A340P; S343A,F,I,L; F348Y; R349P; A352K; P360R; 15 R362P; W364F; R365V, L, A, S; T366D, V, S; S367K, A; S368K; L369I; S373A; G374A,S; R375H; ()383kQ,E; T387P; Q396R; G404A; L409R; T411K; L412R; E417R; F421Y.

Variants of a model phytase derived from a species of the genus Paxillus, preferably the species Paxillus involutus, 20 preferably derived from strain CBS 100231, preferably variants of P_involtus-A2, the sequence of which is shown at Fig. 3, said variants comprising at least one of the following amendments: P24C; I27P; F31Y; I33C; R39H,S,Q; N40L; S42G;

P43A,C,D,E,F,G,H,I,K,L,M,N,Q,R,S,T,V,W,Y; Y44N; S45D;
25 Y47F; A51E,R; A58D,K; E61R; I62V; F75W; S78D; A80K; A81Q,E,G;
R83A,I,Q,R,K; I84Y,Q,V; L88I; K90R,A; F102Y; S115N; D116S;
V118L; P119E; F120L; A123N,T,Q; S125M; F126H,S,V; D127Q,E,N;
A128T,S; V132F,I,L; D143N; I148V; D151A,S; S153D,Y; D154Q,S,G;
D158A; A160S; T161N; ()170fH; ()170gA; S171N; R172P; N173Q,S;
30 P184Q,S; Q185S; T186A,E,P; G187A; ()187aS; T190P,A; D193S;
N194S,T; M195T,V,L; A198N,V; G200V; E201D; ()201eT; S202A;

E417R; Y421F.

D203R,K,S; P203aV,T; Q204E,S,A,V; V205E; S211L,V; S215A,P; L220N; A223D,H; A232T; F235Y,L,T; N236Y; L237F; V238L,M; P242S; M244D; ()251eE,Q; D253P; T256D; P260A,H; E264R,I; A265Q; A267D; G270Y,A,L; D271N; D273K; F275Y; T278H; Y280A,P; A283P; V287A,T; Q288L,I,F; Y292F; I293A,V; N302R,H; A304P; N336S; L337T,Q,S,G; M338I; V339I; 340P,A; A343S,F,I,L; F348Y; R349P; A352K; P360R; R362P; W364F; L365V,A,S; T366D,V,S; S367K,A; S368K; V369I,L; S373A; R375H; ()383kQ,E; T387P; Q396R; G404A; L409R; A411K,T; L412R; E417R; Y421F.

- Variants of a model phytase derived from a species of the genus Trametes, preferably the species Trametes pubescens, preferably derived from strain CBS 100232, preferably variants of T_pubescens, the sequence of which is shown at Fig. 4, said variants comprising at least one of the following amendments:
- 15 R24C; T27P; L31Y; V33C; Q39H,S; S40L,N; S42G;

M43A,C,D,E,F,G,H,I,K,L,N,P,Q,R,S,T,V,W,Y; Y44N; S45D; Y47F; A51E,R; A58D,K; S59G; Q61R; I62V; F75W; S78D; A80K; A81Q,E,G; R83A,I,Q,K; I84Y,Q,V; V88I; K90R,A; L102Y; D115N; V118L; T123N,Q; S125M; S126H,V; E127Q,N; A128T,S; A132F,I,L; 20 D143N; V148I; S151A; S153D,Y; D154Q,S,G; A158D; A160S; N161T; ()170fH; ()170gA; S171N; S172P; N173Q,S; S184Q, P; A186E, P; G187A; ()187aS; T190P, A; N194S, T; M195T, V, L; A198N, V; G200V; ()201eT; S202A; D203R,K,S; P203aV,T; Q204E,S,A,V; V205E; Q211L,V; P215A; L220N; G223D,H; D233E; Y235L,T; N236Y; L237F; 25 L238M; P242S; E244D; ()251eE,Q; E253P; Q260A,H; D264R,I; A265Q; A267D; A270Y, L, G; D271N; D273K; F275Y; T278H; Y280A, P; V287A, T; Q288L,I,F; Y292F; I293A,V; A302R,H; N304P,A; N336S; Q337T,S,G; M338I; V339I; A340P; S343A,F,I,L; F348Y; N349P; A352K; P360R; R362P; F364W; L365V,A,S; V366D,S; K367A; I369L; A373S; A374S; 30 R375H; ()383kQ,E; Q387P; A396R; G404A; V409R; T411K;

Variants of a model phytase derived from a species of the genus Aspergillus, preferably the species Aspergillus nidulans, preferably derived from strain DSM 9743, preferably variants of A_nidulans, the sequence of which is shown at Fig. 10, said variants comprising at least one of the following amendments: V24C; A27P; H39S,Q; V40L,N; G42S;

Q43A,C,D,E,F,G,H,I,K,L,M,N,P,R,S,T,V,W,Y; Y44N; S45D; Y47F; S49P; E51A,R; V56P; H58D,K,A; E61R; V62I; S69Q; Y75W,F; E78D,S; S79G; K80A; S81Q,E,A,G; K82T; A83I,Q,K,R; Y84Q,V,I; 10 A90R; D115N; D116S; T118V,L; I119E; F120L; E122A; N123T,Q; M125S; V126H,S; D127Q,E,N; S128A,T; F132I,L; I148V; K143N; S151A; S153D, Y; D154Q, S, G; A158D; S159T; A160S; E161T, N; K162N; F163W; G170fH; S170gA; ()171N; ()172P; K173Q,S; P184Q,S; E185S; I186A, E, P; D187A; G187aS; T190P, A; H193S; S194T; S198A, N, V; 15 E200G, V; N201D, E; D201e(); E201e(), T; R201f() (a deletion of at least one of 201d, 201e, 201f, preferably all); D203R,K,S; E203aV,T; I204Q,E,S,A,V; I211L,V; P215A; L220N; D223H; K228N; E232T; N233E; I235Y,L,T; Y236N; L237F; M238L; S242P; M246V; E251eQ; A256D; E260A,H; L264R,I; Q270Y,A,L,G; 20 S271D,N; S273D,K; Y275F; G278T,H; A280P; A287T; Q288L,I,F; F292Y; T293A,V; Q302R,H; P304A; N336S; S337T,Q,G; M338I; I339V; S340P,A; F343A,S,I,L; N349P; Q352K; S360R; Q362P; Y364W,F; A365V,L,S; A366D,V,S; S367K,A; W368K; T369I,L; G373S,A; A374S; R375H; A376M; E383kQ; A404G; T411K; L412R; E417R; F421Y; K431E.

Variants of a model phytase derived from a species of Aspergillus, preferably Aspergillus terreus, preferably derived from strain CBS 220.95, preferably variants of A_terreus, the sequence of which is shown at Fig. 12, said variants comprising at least one of the following amendments:

30 G24C; V27P; H39S,Q; K40L,N; G42S;

L43A, C, D, E, F, G, H, I, K, M, N, P, Q, R, S, T, V, W, Y; Y44N; A450,S; Y47F; S49P; Q51E,A,R; V56P; P58D,K,A; D59G; H61R; I62V; A69Q; S75W,F; H78D,S; S79G; K80A; T81Q,E,A,G; A83I,Q,K,R; Y84Q,V,I; A90R; E115N; E116S; T118V,L; P119E; F120L; R122A; N123T,Q; 5 L125S,H; R126H,S,V; D127Q,E,N; L128A,T,S; F132I,L; H143N; V148I; T151A,S; D152G; A153D,Y; S154D,Q,G; H157V; E158D,A; A160S; E161T,N; K162N; F163W; H173Q,S; P184Q,S; E185S; G186A, E, P; S187A; A187aS; T190P, A; H193S; S194T; L195T, V; A198N, V; E200G, V; S201D, E; S201d(); T201e(); V201f(); G202S, A; 10 D203R, K, S; D203aV, T; A204Q, E, S, V; V205E; V211L; A215P; L220N; D223H; Q228N; D232T; D233E; V235Y,L,T; N236Y; L237F; M238L; P242S; E244E; T251eE,Q; A260H; T264R,I; Q265A; N267D; L270Y,A,G; S271D,N; K273D; Y275F; H278T; G280A,P; V287A,T; Q288L,I,F; W292F,Y; A293V; Q302H; P304A; N337T,Q,S,G; L338I; V339I; 15 S340P,A; W343A,S,F,I,L; N349P; A352K; S360R; S362P; Y364W,F; A365V,L,S; A366D,V,S; A367K; W368K; T369I,L; A373S; A374S; R375H; A376M; R383kQ,E; P404A,G; K411T; A417E,R; F421Y; A431E.

Variants of a model phytase derived from a species of Talaromyces, preferably the species Talaromyces thermophilus, 20 preferably derived from strain ATCC 20186 or ATCC 74338, preferably variants of T_thermo, the sequence of which is shown at Fig. 13, said variants comprising at least one of the following amendments:

H24C; V27P; H39S,Q; S4OL,N; G42S;

Q43A,C,D,E,F,G,H,I,K,L,M,N,P,R,S,T,V,W,Y; Y44N; S45D; F47Y; S49P; A51E,R; V56P; Q58D,K,A; N59G; K61R; I62V; Y75W,F; S78D; S79G; K80A; T81Q,E,A,G; E82T; L83A,I,Q,R,K; Y84Q,V,I; R90A; D116S; T118V,L; P119E; F120L; E122A; N123T,Q; M125S; I126H,S,V; Q127E,N; L128A,T,S; F132I,L; V148I; S151A; S153D,Y; 30 D154Q,S,G; I157V; A158D; S159T; G160A,S; R161T,N; L162N; F163W; S170gA; D171N; K172P; H173Q,S; E184Q,S,P; E185S; G186A,E,P;

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D187A; T190P,A; T193S; G194S,T; S195T,V,L; V198A,N; E200G,V; D201E; S201d(); S201e(),T; S201f(); G202S,A; H203R,K,S; D203aV,T; A204Q,E,S,V; Q205E; Q211L,V; A215P; I220N,L; H223D; D228N; S232T; D233E; P235Y,L,T; Y236N; M237F; D238L,M; P242S; 5 E244D; L246V; ()251eE,Q; A256D; Q260A,H; Q264R,I; A265Q; Q270Y,A,L,G; S271D,N; G273D,K; Y275F; N278T,H; G280A,P; A287T; Q288L,I,F; F292Y; V293A; H302R; P304A; N336S; T337Q,S,G; M338I; T339V,I; S340P,A; A343S,F,I,L; N349P; A352K; S360R; E362P; Y364W,F; S365V,L,A; A366D,V,S; A367K; W368K; T369I,L; G373S,A; 10 G374A,S; R375H; A376M; D383kQ,E; E404A; K411T; R417E; F421Y.

Variants of a model phytase derived from a species of Thermomyces, preferably the species Thermomyces lanuginosus, preferably derived from strain DBS 586.94, preferably variants of T_lanuginosa, the sequence of which is shown at Fig. 14, said variants comprising at least one of the following amendments: K24C; ()27P; ()31Y; ()33C; R39H,S,Q; H40L,N; G42S;

S45D; Q43A,C,D,E,F,G,H,I,K,L,M,N,P,R,S,T,V,W,Y; Y44N; F47Y; S49P; A51E,R; V56P; K58D,A; V62I; S69Q; Y75W,F; A78D,S; H79G; K80A; S81Q, E, A, G; E82T; V83A, I, Q, K, R; Y84Q, V, I; L88I; 20 R90A; F102Y; D115N; N116S; T118V,L; R119E; F120L; E122A; E123N, T,O; M125S; M126H,S,V; E127Q,N; S128A,T; F132I,L; E143N; V148I; A151S; S153D,Y; A154D,Q,S,G; I157V; A158D; S159T; A160S; E161T,N; F162N; F163W; R170fH; S170gA; K172P; D173Q,S; S184Q,P; E185S; E186A, P; T187A; G187aS; T190P, A; G193S; L194S, T; T195V, L; 25 A198N, V; E200G, V; E201D; A201d(); P201e(), T; D202S, A; P203R, K, S; T203aV; O204E,S,A,V; P205E; V211L; R215A,P; I220L,N; H223D; E232T; D233E; P235Y,L,T; L236Y,N; M238L; P242S; Q251eE; H256D; O260H; M264R,I; A265Q; Y270A,L,G; T271D,N; D273K; Y275F; H278T; G280A,P; A283P; S287A; R288L,I,F; F292Y; V293A; G302R,H; P304A; 30 N336S; T337Q,S,G; M338I; T339V,I; G340P,A; S343A,F,I,L; N349P; P360R; T362P; Y364W,F; A365V,L,S; A366D,V,S; S367K,A; W368K; T369I,L; A373S; A374S; R375H; A376M; E383kQ; R404A,G; R411K,T; K417E,R; F421Y; D431E.

Variants of a model phytase derived from a species of Myceliophthora, preferably the species Myceliophthora thermophila, preferably derived from strain ATCC 48102 or ATCC 74340, preferably variants of M_thermophila, the sequence of which is shown at Fig. 7, said variants comprising at least one of the following amendments:

S24C; F31Y; H39S,Q; F40L,N; G42S;

Q43A,C,D,E,F,G,H,I,K,L,M,N,P,R,S,T,V,W,Y; Y44N; S45D; 10 Y47F; S49P; P51E,A,R; I56P; D58K,A; D59G; E61R; V62I; S69Q; A75W, F; L78D, S; K79G; R80K, A; A81Q, E, G; A82T; S83A, I, Q, K, R; Y84Q, V, I; R90A; D115N; E116S; T118V, L; R119E; T120L; Q122A; Q123N,T; M125S; V126H,S; N127Q,E; S128A,T; F132I,L; K143N; 15 V148I; A151S; Q153D,Y; D154Q,S,G; H158D,A; S159T; A160S; E161T,N; G170fH; S170gA; T171N; F163W; V172P; R173Q,S; P184Q,S; E185S; T186A, E, P; G187aS; T190P, A; N193S; D194S, T; L195T, V; A198N,V; E200G,V; E201D; G201a(); P201b(); Y201c(); S201d(); T201e(); I201f(); G202S,A; D203R,K,S; D203aV,T; A204Q,E,S,V; 20 Q205E; T211L,V; P215A; V220N,L; N223D,H; A232T; D233E; V235Y,L,T; A236Y,N; L237F; M238L; P242S; E244D; A251eE,Q; R256D; E260A, H; R264I; A265Q; Q270Y, A, L, G; S271D, N; K273D; Y275F; Y278T,H; P280A; T287A; Q288L,I,F; F292Y; V293A; ()302R,H; P304A; N336S; D337T,Q,S,G; M338I; M339V,I; G340P,A; G343A,S,F,I,L; 25 D349P; P352K; D360R; E362P; Y364W,F; A365V,L,S; A366D,V,S; S367K,A; W368K; A369I,L; A373S; A374S; R375H; I376M; E383kQ; E387P; G404A; M409R; T411K; L412R; E417R; F421Y; D431E.

This invention also provides a new phytase which has been derived from a strain of Cladorrhinum, viz. C. foecundissimum.

30 Accordingly, the invention also relates to a polypeptide having phytase acitivity and which comprises SEQ ID NO:2 or the mature

part (amino acids nos 16-495) thereof; or a polypeptide being at least 70, more preferably 75, 80, 85, 90, 95% homologous thereto; homology meaning similarity, preferably identity, and being determined using the program GAP and the settings as 5 defined hereinabove. And the invention relates to DNA construct which encodes a polypeptide having phytase activity, said DNA construct comprising a DNA molecule which comprises SEQ ID NO:1 or nucleotides nos. 20-70 and 207-1560 thereof; nucleotides nos. 20-70 and 207-1563 thereof; or nucleotides nos. 10 65-70 and 207-1560 thereof; or nucleotides nos. 65-70 and 207-1563 thereof; or a DNA construct or molecule which is at least 70, 75, 80, 85, 90, 95 % homologous to either of these nucleotide sequences; homology meaning similarity, preferably identity, and being determined using computer programs known in 15 the art such as GAP provided in the GCG program package (Program Manual for the Wisconsin Package, Version 8, August 1996, Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, 53711) (Needleman, S.B. and Wunsch, C.D., (1970), Journal of Molecular Biology, 48, 443-453). Using GAP with the following 20 settings for DNA sequence comparison: GAP creation penalty of 5.0 and GAP extension penalty of 0.3. The invention also relates to a DNA construct which hybridizes with any of the above DNA sequences under the conditions mentioned hereinabove.

25 EXAMPLES

Example 1

Phytase activity assay (FYT)

Phytase activity can be measured using the following assay:

30 10 μ l diluted enzyme samples (diluted in 0.1 M sodium acetate, 0.01 % Tween20, pH 5.5) are added into 250 μ l 5 mM sodium

phytate (Sigma) in 0.1 M sodium acetate, 0.01 % Tween20, pH 5.5 (pH adjusted after dissolving the sodium phytate; the substrate is preheated) and incubated for 30 minutes at 37°C. The reaction is stopped by adding 250 µl 10 % TCA and free phosphate is 5 measured by adding 500 µl 7.3 g FeSO4 in 100 ml molybdate reagent (2.5 g (NH₄) 6MO₇O₂₄.4H₂O in 8 ml H₂SO₄ diluted to 250 ml). The absorbance at 750 nm is measured on 200 µl samples in 96 well microtiter plates. Substrate and enzyme blanks are included. A phosphate standard curve is also included (0-2 mM phosphate). 1 FYT equals the amount of enzyme that releases 1 µmol phosphate/min at the given conditions.

Example 2

Test for specific activity

The specific activity can be determined as follows:

A highly purified sample of the phytase is used (the purity is checked beforehand on an SDS poly acryl amide gel showing the presence of only one component).

The protein concentration in the phytase sample is determined by amino acid analysis as follows: An aliquot of the phytase sample is hydrolyzed in 6N HCl, 0.1% phenol for 16 h at 110 C in an evacuated glass tube. The resulting amino acids are quantified using an Applied Biosystems 420A amino acid analysis system operated according to the manufacturers instructions.

25 From the amounts of the amino acids the total mass - and thus

5 From the amounts of the amino acids the total mass - and thus also the concentration - of protein in the hydrolyzed aliquot can be calculated.

The activity is determined in the units of FYT. One FYT equals the amount of enzyme that liberates 1 micromol inorganic 30 phosphate from phytate (5 mM phytate) per minute at pH 5.5, 37°C; assay described e.g. in example 1.

60

The specific activity is the value of ${\tt FYT/mg}$ enzyme protein.

Example 3

5 Test for temperature and pH activity and stability

Temperature and pH activity and stability can be determined as follows:

Temperature profiles (i.e. temperature activity relationship) by running the FYT assay of Example 1 at various 10 temperatures (preheating the substrate).

Temperature stability by pre-incubating the phytase in 0.1 M sodium phosphate, pH 5.5 at various temperatures before measuring the residual activity.

The pH-stability by incubating the enzyme at pH 3 (25 mM 15 glycine-HCl), pH 4-5 (25 mM sodium acetate), pH 6 (25 mM MES), pH 7-9 (25 mM Tris-HCl) for 1 hour at 40°C, before measuring the residual activity.

The pH-profiles (i.e. pH activity relationship) by running the assay at the various pH using the same buffer-systems (50 mM, pH re-adjusted when dissolving the substrate).

Example 4

DSC as a preferred test for thermostability

The thermostability or melting temperature, Tm, can be 25 determined as follows:

In DSC the heat consumed to keep a constant temperature increase in the sample-cell is measured relative to a reference cell. A constant heating rate is kept (e.g. 90°C/hour). An endothermal process (heat consuming process - e.g. the unfolding of an enzyme/protein) is observed as an increase in the heat

transferred to the cell in order to keep the constant temperature increase.

DSC can be performed using the MC2-apparatus from MicroCal. Cells are equilibrated 20 minutes at 20°C before scanning to 90°C at a scan rate of 90°/h. Samples of e.g. around 2.5 mg/ml phytase in 0.1 M sodium acetate, pH 5.5 are loaded.

Example 5

Phytase variants of amended activity characteristics

Variants of an Aspergillus fumigatus model phytase (a wild type phytase derived from strain ATCC 13073) were prepared as described in EP 98104858.0 (EP-A-0897010), examples 2-3 and 5, and the phytase activity was determined as described in example 7 thereof. pH- and temperature optimum and melting point was determined as described in examples 9 and 10 of EP 98113176.6 (EP-A-0897985).

In Table 1, variants of improved specific activity at pH 5.0 are listed. Table 2 lists variants of improved relative activity at pH 3.0, and Table 3 lists variants of improved thermostability (temperature optimum, e.g. determined by DSC).

Table 1

20

Amended in position | Substitution into Specific activity at no. pH 5.0 (U/mg) 43 43L 83.4 43N 45.5 43T 106.9 43I 91.2 43V 35.0 43A 27.3 43G 59.6

43 and 270	43L, 270L	88.7
43 and 270 and 273	43L, 270L, 273D	92.3
43 and 78	43L, 78D	118.5
43 and 153 and 154	43L, 153Y, 154G	193.0
A. fumigatus wild-	-	26.5
type phytase		

Table 2

Amended in position	Substitution into	Relative phytase
no.		activity at pH 3.0
205	205E	41%
273	273K	61%
278	278Н	75%
273 and 205	273K, 205E	65%
273 and 278	273К, 278Н	100%
273 and 205 and 278	273K, 205E, 278H	96%
A. fumigatus wild-	-	32%
type phytase		

Table 3

Amended in position	Substitution into	Tempera-	Tm (°C)
no.		ture	(DSC)
	1	optimum	
	to -	(°C)	
43 and 47 and 88 and	43T, 47Y, 88I, 102Y,	60	67
102 and 220 and 242	220L, 242P, 267D		
and 267			
as above plus 51 and	as above plus 51A,	63	_
302 and 337 and 373	302н, 337т, 373А,		
and 115	115N		

A.	fumigatus	wild-	_	55	62.5
type	e phytase				

Example 6

Further phytase variants of amended activity characteristics

Variants of the ascomycete consensus sequence "conphys" of Fig. 9 were prepared as described in EP 98113176.6 (EP-A-5 0897985), examples 4-8. Phytase activity, including pH- and temperature optimum, and melting point was determined as described in examples 9 and 10, respectively, thereof.

The tables below list variants of amended activity characteristics, viz.

Table 4 variants of improved specific activity at pH 6.0;
Table 5 variants of amended pH optimum (the pH-optimum indicated is an approximate value, determined as that pH-value (selected from the group consisting of pH 4.0; 4.5; 5.0; 5.5; 6.0; 6.5; and 7;0) at which the maximum phytase activity was obtained);

Table 6 a variant of improved thermostability (expressed by way of the melting point as determined by differential scanning calorimetry (DSC)); and

Table 7 variants of amended thermostability (temperature 20 optimum); a "+" or "-" indicates a positive or a negative, respectively, effect on temperature optimum of up to 1°C; and a "++" and "--" means a positive or a negative, respectively, effect on temperature optimum of between 1 and 3°C.

25 Table 4

Amended	in	position	Substitution	into	Specific activity at
no.					pH 6.0 (U/mg)
43			43T		130

	43L	205
Conphys	-	62

Table 5

Amended in position	Substitution into	pH optimum
no.		around
43	43T	6.0
,	43L	5.5
	43G	6.5
43 and 44	43L, 44N	6.0
	43T, 44N	5.5
Conphys	-	6.0

Table 6

Amended in position	Substitution into	Tm (°C)
no.		
43	43T	78.9
Conphys	-	78.1

5 <u>Table 7</u>

Amended in position	Substitution into	Temperature optimum
no.		amendment
51	A	+
58	K	+
220	N	+
195	L	++
201e	T	++
244	D	+
264	I	+
302	Н	+

227		
337	Т	++
352	К	+
373	A	++
47	F	-
62	I	
83	K	
90	R	
143	N	_
148		
	V	
186	A	
187a	S	_
198	V	-
204	A	
211	V	- :
215	P	
251e	Q	_
260	A	
265	A	
		-
339	V	
365	A	
383k	E	_
404	Ğ	
417	R	
Conphys	-	0
	I .	

Table 8

Amended in position	Substitution	Tm (°C) (DSC)	Specific
no.	into		activity at
:-			pH 5.0 (U/mg)
43 and 51 and 220	51A, 220N,	84.7	105
and 244 and 264 and	244D, 264I,		
302 and 337 and 352	302н, 337т,	•	
and 373	352K, 373A,		
	43T	·	
as above plus 80	as above plus	85.7	180
	80A	·	
Conphys		78.1	30

Example 7

Cloning of a phytase of Cladorrhinum foecundissimum

DNA encoding a phytase from Cladorrhinum foecundissimum CBS 427.97 has been cloned, and the enzyme isolated and purified, essentially as described in WO 98/28409.

Fig. 15 shows the DNA sequence of the HindIII/XbaI cloned PCR product in pA2phy8. The cloned PCR product is amplified from the genomic region encoding Cladorrhinum foecundissimum CBS 427.97 phyA gene. The putative intron is indicated by double underline of the excision-ligation points in accordance with the GT-AG rule (R. Breathnach et al. Proc. Natl. Acad. Sci. USA 75 (1978) pp4853-4857). The restrictions sites used for cloning are underlined.

According to the SignalP V1.1 prediction (Henrik Nielsen, Jacob Engelbrecht, Stren Brunak and Gunnar von Heijne: 5 "Identification of prokaryotic and eukaryotic signal peptides and prediction of their cleavage sites," Protein Engineering 10, 1-6 (1997)), the signal peptide part of the enzyme corresponds

to amino acids nos. 1-15, accordingly the mature enzyme is amino acids nos. 16-495.

The enzyme exhibits a pH optimum around pH 6 with no activity at the low pH (pH 3), but significant activity up until 5 pH 7.5; thus it is a more alkaline phytase as compared to the Aspergillus ficuum phytase.

A temperature optimum around 60°C was found at pH 5.5. Thus, this phytase is more thermostable than the A. ficuum phytase.

Example 8

10

Alignment of a new model phytase according to Fig. 1

The phytase sequence of Cladorrhinum foecundissimum as disclosed in Example 7 is compared with the 13 model phytases of 15 Fig. 1 using GAP version 8 referred to above with a GAP weight of 3.000 and a GAP lengthweight of 0.100. Complete amino acid sequences are compared. The M_thermophila phytase sequence turns up to be the most homologous sequence, showing a degree of similarity to the C. foecundissimum sequence of 70.86%.

Still using the GAP program and the parameters mentioned above, the phytase sequence "C_foecundissimum" is now aligned to the "M-thermophila" phytase - see Fig. 16. The average match is 0.540;, the average mismatch -0.396; quality 445.2; length 505; ratio 0.914; gaps 9; percent similarity 70.860; percent identity 25 53.878.

In a next step, see Fig. 17, the C_foecundissimum is pasted (or it could simply be written) onto the alignment of Fig. 1 as the bottom row, ensuring that those amino acid residues which according to the alignment at Fig. 16 are identical (indicated by a vertical line) or similar (indicated by one or two dots) are placed above each other. At 5 places along the sequence, the C_foecundissimum sequence comprises

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"excess" amino acid residues, which the alignment of Fig. 1 does not make room for. At Fig. 17, these excess residues are transferred onto a next row (but they can be included in the multiple alignment and numbered as described previously in the position numbering related paragraphs (using the denotations a, b, c etc.).

Corresponding variants of the phytase of C_foecundissimum are then easily deduced on the basis of Fig. 17. Some examples:

The variants generally designated "80K,A" and "43T" in

10 C_foecundissimum correspond to "K80A" and "Q43T," respectively.

CLAIMS

- A phytase variant which, when aligned according to Fig. 1, is amended as compared to a model phytase in at least one of the following positions, using the position numbering corresponding to P_lycii:
 - 24; 27; 31; 33; 39; 40; 41; 42; 43; 44; 45; 46; 47; 49; 51; 56; 58; 59; 61; 62; 68; 69; 70; 71; 72; 73; 74; 75; 76; 77; 78; 79; 80; 81; 82; 83; 84; 88; 90; 102; 115; 116; 117; 118; 119; 120;
- 121; 122; 123; 124; 125; 126; 127; 128; 132; 143; 148; 149; 150;
- 10 151; 152; 153; 154; 155; 156; 157; 158; 159; 160; 161; 162; 163; 170f; 170g; 171; 172; 173; 184; 185; 186; 187; 187a; 190; 191; 192; 193; 194; 195; 198; 199; 200; 201; 201a; 201b; 201c; 201d;
 - 201e; 201f; 202; 203; 203a; 204; 205; 211; 215; 220; 223; 228;
 - 232; 233; 234; 235; 236; 237; 238; 239; 242; 243; 244; 246;
- 15 251e; 253; 256; 260; 264; 265; 267; 270; 271; 272; 273; 274;
 - 275; 276; 277; 278; 279; 280; 283; 285; 287; 288; 292; 293; 302;
 - 304; 332; 333; 334; 335; 336; 337; 338; 339; 340; 341; 342; 343;
 - 348; 349; 352; 360; 362; 364; 365; 366; 367; 368; 369; 370; 371;
- 372; 373; 374; 375; 376; 383k; 387; 393; 394; 396; 404; 409;
- 20 411; 412; 413; 417; 421; 431.
- 2. A phytase variant which, when aligned according to Fig. 1, comprises at least one of the following amendments as compared to a model phytase, using the position numbering corresponding to the phytase of P lycii:
 - 24C; 27P; 31Y; 33C; 39H,S,Q; 40L,N; 42S,G;
 - 43A,C,D,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y; 44N; 45D,S; 47Y,F;
 - 49P; 51E,A,R; 56P; 58D,K,A; 59G; 61R; 62V,I; 69Q; 75W,F; 78D,S;
 - 79G; 80K,A; 81A,G,Q,E; 82T; 83A,I,K,R,Q; 84I,Y,Q,V; 88I; 90R,A;
- 30 102Y; 115N; 116S; 118V,L; 119E; 120L; 122A; 123N,Q,T; 125M,S;

126H,S,V; 127Q,E,N; 128A,S,T; 132F,I,L; 143N; 148V,I; 151A,S; 152G; 153D,Y; 154D,Q,S,G; 157V; 158D,A; 159T; 160A,S; 161T,N; 162N; 163W; 170fH; 170gA; 171N; 172P; 173Q,S; 184Q,S,P; 185S; 186A,E,P; 187A; 187aS; 190A,P; 193S; 194S,T; 195T,V,L; 198A,N,V; 200G,V; 201D,E; 201a(); 201b(); 201c(); 201d(); 201e(); 201f(); 201eT; 202S,A; 203R,K,S; 203aV,T; 204Q,E,S,A,V; 205E; 211L,V; 215A,P; 220L,N; 223H,D; 228N; 232T; 233E; 235Y,L,T; 236Y,N; 237F; 238L,M; 242P,S; 244D; 246V; 251eE,Q; 253P; 256D; 260A,H; 264R,I; 265A,Q; 267D; 270Y,A,L,G; 271D,N; 273D,K; 275F,Y; 10 278T,H; 280A,P; 283P; 287A,T; 288L,I,F; 292F,Y; 293A,V; 302R,H; 304P,A; 332F; 336S; 337T,G,Q,S; 338I; 339V,I; 340P,A; 343A,S,F,I,L; 348Y; 349P; 352K; 360R; 362P; 364W,F; 365V,L,A,S; 366D,S,V; 367A,K; 368K; 369I,L; 370V; 373A,S; 374S,A; 375H; 376M; 383kQ,E; 387P; 393V; 396R; 404A,G; 409R; 411K,T; 412R; 15 417E,R; 421F,Y; 431E.

- 3. The phytase variant of any of claims 1 or 2, which is derived from an ascomycete phytase.
- 20 4. The phytase variant of claim 3 which is derived from an Aspergillus phytase.
- The phytase variant of claim 4, wherein the model phytase is a strain of Aspergillus niger, Aspergillus ficuum,
 Aspergillus nidulans, Aspergillus fumigatus, Aspergillus terreus.
- 6. The phytase variant of claim 5 wherein the model phytase is Aspergillus nidulans DSM 9743; or any of the following 30 strains of Aspergillus terreus: CBS 116.46, DSM 9076, CBS 220.95.

7. The phytase variant of claim 6 wherein the model phytase is the Aspergillus nidulans phytase sequence shown in Fig. 10; or the Aspergillus terreus phytase sequence shown in Fig. 12.

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- 8. The phytase variant of claim 3 wherein the model phytase is a strain of Thermomyces lanuginosus, Talaromyces thermophilus, or Myceliophthora thermophila.
- 10 9. The phytase variant of claim 8 wherein the model phytase is Thermomyces lanuginosus CBS 586.94; or any of the following strains of Talaromyces thermophilus: ATCC 20186, ATCC 74338; or any of the following strains of Myceliophthora thermophila: ATCC 34625, ATCC 74340.

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10. The phytase variant of claim 9 wherein the model phytase is the Thermomyces lanuginosus phytase sequence shown in Fig.14; or the Talaromyces thermophilus sequence shown in Fig.13; or the Myceliophthora thermophila phytase sequence shown in Fig.7.

20

- 11. The phytase variant of claim 3 wherein the model phytase is an ascomycete consensus phytase sequence.
- 12. The phytase variant of any of claims 1 or 2, which is 25 derived from a basidiomycete phytase.
 - 13. The phytase variant of claim 12, wherein the model phytase is a strain of Paxillus involutus, Trametes pubescens, Agrocybe pediades, or Peniophora lycii.

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- 14. The phytase variant of claim 13 wherein the model phytase is Trametes pubescens CBS 100232 or Paxillus involutus CBS 100231.
- 5 15. The phytase variant of claim 14 wherein the model phytase is the Trametes pubescens phytase sequence of Fig. 4 or either of the Paxillus involutus phytase sequences of Figs. 2 and 3.
- 16. The phytase variant according to any of claims 1 or 2, 10 which comprises at least one of the following amendments:

 R24C; V27P; H39Q,S; L40N; G42S;

Q43A,C,D,E,F,G,H,I,K,L,M,N,P,R,S,T,V,W,Y; Y44N; A45D,S; F47Y; S49P; A51E,R; V56P; A58D,K; V62I; S69Q; Y75W,F; D78S; S79G; K80A; G81A,Q,E; K82T; K83A,I,R,Q; Y84Q,I,V; E90R,A; D115N;

- 15 D116S; T118V,L; P119E; F120L; E122A; Q123N,T; L125S,M; V126H,S;
 N127Q,E; S128A,T; F132I,L; I148V; S151A; S153D,Y; S154Q,D,G;
 I157V; A158D; S159T: G160A,S; K161T,N; K162N; F163W; R170fH;
 Q171N; G173Q,S; S184P,Q; E185S; A186E,P; S187A; T190P,A; P193S;
 G194S,T; T195V,L; V198A,N; E200G,V; D201E; S201d(); E201e(),T;
- 20 L201f(); preferably all three deletions; A202S; D203R,K,S;
 D203aV,T; V204Q,E,S,A; T211L,V; S215AP; L220N; D223H; T228N;
 T235Y,L; Y236N; L237F; M238L; S242P; I246V; K251eE,Q; H260A;
 I264R; N265Q,A; Q270Y,A,L,G; S271D,N; K273D; Y275F; H278T;
 A280P; T287A; Q288L,I,F; Y292F; A293V; H302R; P304A; N336S;
- 25 G337S,T,Q; I339V; S340P,A; F343A,S,F,I,L; N349P; N360R; T362P; F364W; S365V,L,A; S366D,V; A367K; W368K; T369I,L; A373S: S374A; R375H; L376M; Q383kE; P404A,G; T411K; R417E; F421Y; A431E.
- 17. The phytase variant of claim 16, the model phytase of 30 which is an Aspergillus derived phytase, preferably derived from Aspergillus ficuum or Aspergillus niger.

- 18. The phytase variant of claim 17, the model phytase of which is a phytase derived from either of Aspergillus ficuum (niger) NRRL 3135, Aspergillus niger ATCC 9142, or Aspergillus 5 niger ATCC 74337.
 - 19. The phytase variant of claim 18, the model phytase of which is the Aspergillus ficuum phytase sequence of Fig. 11.
- 10 20. The phytase variant according to any of claims 1 or 2, which phytase variant comprises at least one of the following amendments:

A24C; V27P; H39,S,Q; L40N; G42S; Q43C,D,E,F,H,K,M,P,R,S,W,Y; Y44N; S45D; F47Y; S49P; E51A,R; L56P; K58D,A; D59G; I62V; S69Q;

- 15 Y75W,F; S78D; S79G; K80A; S81A,G,Q,E; K82T; K83A,I,Q,R; Y84Q,V,I; V88K; A90R; F102Y; D115N; D116S; T118V,L; P119E; F120L; E122A; Q123N,T; L125S,M; V126H,S; N127Q,E; S128A,T; F132,I,L; S143N; I148V; S151A; S153D,Y; D154Q,S,G; I157V; A158D; S159T; G160A,S; E161T,N; K162N; F163W; G170fH; ()171N; N173Q,S;
- 20 T172P; P184Q,S; E185S; S186A,E,P; E187A; T187aS; T190P,A;
 G194S,T; V195L,T; K198A,N,V; E200G,V; A201D,E; S201d();
 Q201e(),T; L201f(); preferably all three deletions; G202S,A;
 D203R,K,S; E203aV,T; V204Q,E,S,A; A205E; L211V; A220L,N; H223D;
 T228N; E232T; D233E; V235Y,L,T; V236Y,N; L237F; M238L; C242P,S;
- 25 T246V; Q251eE,Q; Q256D; H260A; K264R,I; K265Q,A; N267D; Q270Y,A,L,G; S271D,N; G273D,K; Y275F; Y278T,H; A280P; A287T; Q288L,I,F; F292Y; T293A,V; R302H; P304A; F332F; N336S; S337T,G,Q; M338I; V339I; S340P,A; F343A,S,I,L; N349P; E352K; S360R; K362P; Y364W,F; S365V,L,A; A366D,V,S; S367A,K; W368K;
- 30 V369I,L; G373S,A; R375H; A376M; K383kQ,E; D404A,G; K411T; I393V; L412R; K417E,R; W421F,Y; G431E.

21. The phytase variant of claim 20, which is derived from an Aspergillus phytase, preferably using a model phytase derived from Aspergillus fumigatus.

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22. The phytase variant of claim 21, the model phytase of which is a phytase derived from either of the following strains of Aspergillus fumigatus: ATCC 13073, ATCC 32722, ATCC 58128, ATCC 26906 or ATCC 32239.

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- 23. The phytase variant of claim 22, the model phytase of which is the Aspergillus fumigatus phytase sequence of Fig. 8.
- 24. The phytase variant according to any of claims 1 or 2, 15 which phytase variant comprises at least one of the following amendments:

G24C; V27P; H39S,Q; L40N; G42S;

Q43A,C,D,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y; Y44N; S45D; Y47F; S49P; E51A,R; V56P; D58K,A; D59G; V62I; S69Q; Y75W,F; S78D;

- 20 S79G; K80A; S81A,G,Q,E; K82T; A83I,Q,K,R; Y84,Q,I,V; A90R;
 D115N; D116S; T118V,L; F119E; P120L; E122A; N123Q,T; M125S;
 V126H,S; N127Q,E; S128A,T; Y132F,I,L; K143N; I148V; S151A;
 S153D,Y; D154Q,S,G; I157V; A158D; S159T; A160S; E161T,N; K162N;
 F163W; G170fH; S170qA; Q171N; H173Q,S; P184Q,S; E185S;
- 25 G186A, E, P; S187A; G187aS; T190P, A; H193S; G194S, T; T195V, L;
 A198N, V; E200G, V; D201E; S201d(); E201e(), T; L201f(); preferably
 all three; G202S, A; D203R, K, S; D203aV, T; V204Q, S, A, E; L211V;
 A215P; L220N; D223H, T228N; E232T; D233E; V235Y, L, T; Y236N;
 L237F; M238L; P242S; E244D; E251e, Q; A256D; H260A; R264I; Q265A;
- 30 Q270Y,A,L,G; S271D,N; G273D,K; Y275F; Y278T,H; A280P; A287T; Q288L,I,F; F292Y; A293V; R302H; P304A; N336S; S337T,Q,G; M338I;

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I339V; S340P,A; F343A,S,I,L; N349P; A352K; S360R; E362P; Y364W,F; S365V,L,A; A366D,V,S; S367K,A; W368K; T369I,L; G373S,A; A374S; R375H; A376M; Q383kE; A404G; K411T; E417R; F421Y; A431E.

- 5 25. The phytase variant of claim 24, the model phytase of which is an ascomycete consensus phytase.
 - 26. The phytase variant of claim 25, the model phytase of which is the ascomycetes consensus sequence "conphys" of Fig. 9.
 - 27. The phytase variant according to any of claims 1 or 2, which phytase variant comprises at least one of the following amendments:

V24C; F27P; ()31Y; F33C; D39H,S,Q; S40L,N; A42S,G;

- 15 A43C, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, Y; Y44N; T45D, S; Y47F; Q51E, A, R; K58D, A; K61R; I62V; F75W; S78D; A80K; G81A, Q, E; R83A, I, Q, K; I84Y, Q, V; V88I; K90R, A; L102Y; D115N; D116S; V118L; P119E; F120L; L123N, T, Q; S125M; S126H, V; Q127E, N; A128S, T; T132F, I, L; E143N; V148I; S151A; S152G; S153D, Y; N154D, Q, S, G;
- 20 D158A; S159T; A160S; T161N; ()170fH; ()170gA; ()171N; H173Q,S;
 H172P; S184Q,P; E185S; S186A,E,P; L187A; ()187aS; T190P,A;
 D193S; A194S,T; M195T,V,L; N198A,V; G200V; S201D,E ()201eT;
 S202A; D203R,K,S; P203aV,T; Q204E,S,A,V; T205E; I211L,V; P215A;
 L220N; Q223D,H; A232T; D233E; S235Y,L,T; N236Y; L237F; I238L,M;
- 25 A242P,S; E244D; I246V; ()251eE,Q; N256D; P260A,H; A264R,I; Q265A; E267D; G270Y,A,L; L332F; D271N; D273K; F275Y; T278H; Y280A,P; Y283P; V287A,T; Q288L,I,F; Y292F; I293A,V; E302R,H; P304A; L332F; N336S; Q337T,S,G; M338I; I339V; A340P; S343A,F,I,L; F348Y; N349P; S352K; P360R; R362P; W364F; 30 V365L,A,S; T366D,V,S; S367K,A; R368K; L369I; T370V; S373A;

- A374S; R375H; S383kQ,E; T387P; A396R; G404A; L409R; T411K; L412R; E417R; Y421F.
- 28. The phytase variant of claim 27, the model phytase of 5 which is a phytase derived from Agrocybe pediades,
 - 29. The phytase variant of claim 27, the model phytase of which is a phytase derived from Agrocybe pediades CBS 900.96.
- 10 30. The phytase variant of claim 29, the model phytase of which is the Agrocybe pediades phytase sequence of Fig. 5.
- 31. The phytase variant according to any of claims 1-2, which phytase variant comprises at least one of the following 15 amendments:
 - F24C; V27P; L31Y; I33C; S39H,Q; N40L; G42S; P43A,C,D,E,F,G,H,I,K,L,M,N,Q,R,S,T,V,W,Y; Y44N; D45S; F47Y; E51A,R; E58D,K,A; T61R; V62I; W75F; S78D; A80K; R81Q,E,G,A; S82T; R83A,I,Q,K; Q84Y,V,I; V88I; K90R,A; A115N; D116S; L118V;
- 20 P119E; F120L; N123T,Q; S125M; H126S,V; Q127E,N; T128A,S;
 M132F,I,L; G143N; V148I; A151S; D153Y; Q154D,S,G; D158A; S159T;
 S160A; T161N; ()170fH; ()170gA; S171NG172P; E173Q,S; Q184S,P;
 E185S; E186A,P; G187A; ()187aS; T190P,A; N193S; N194S,T;
 M195T,V,L; N198A,V; V200G; D201E; ()201eT; G202S,A; D203R,K,S;
- 25 ()203aV,T; E204Q,S,A,V; S205E; V211L; N215A,P; L220N; A223D,H;
 S232T; D233E; L235Y,T; T236Y,N; L237F; M238L; P242S; L246V;
 ()251eE,Q; A260H; V264R,I; S265Q,A; E267D; Y270A,L,G; D271N;
 D273K; F275Y; G278T,H; P280A; A283P; T287A; Q288L,I,F; Y292F;
 V293A; G302R,H; A304P; N336S; T337Q,S,G; M338I; V339I; P340A;

A352K; E360R;

R362P;

W364F;

30 A343S, F, I, L; F348Y; N349P;

15

V365L,A,S; D366V,S; S367K,A; L369I; S373A; G374A,S; ()383kQ,E; E387P; A396R; G404A; V409R; E411K,T; L412R; E417R; Y421F; A431E.

- 32. The phytase variant of claim 31, the model phytase of 5 which is a phytase derived from Peniophora lycii.
 - 33. The phytase variant of claim 32, the model phytase of which is a phytase derived from Peniophora lycii CBS 686.96.
- 10 34. The phytase variant of claim 33, the model phytase of which is the Peniophora lycii phytase sequence of Fig. 6.
 - 35. A phytase polypeptide which comprises a phytase variant according to any of the previous claims.
 - 36. A DNA construct comprising a DNA sequence encoding a phytase variant according to any one of claims 1-34.
- 37. A recombinant expression vector which comprises a DNA 20 construct according to claim 36.
 - 38. A host cell which is transformed with a DNA construct according to claim 36 or a vector according to claim 37.
- 25 39. A process for preparing a phytase variant, the process comprising culturing the host cell according to claim 38 under conditions permitting the production of the phytase variant, and recovering the phytase from the culture broth.
- 30 40. A feed or food comprising at least one phytase variant of any of claims 1-34.

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41. A process for preparing a feed or food according to claim 40, wherein the at least one phytase variant is added to the food or feed components.

5

- 42. A composition comprising at least one phytase variant of any of claims 1-34.
- 43. The composition according to claim 42 suitable for use in 10 food or feed preparations.
 - 44. The composition according to any of claims 42-43 which is an animal feed additive.
- 15 45. A process for reducing phytate levels in animal manure comprising feeding an animal with an effective amount of the feed according to claim 40 or obtainable according to claim 41.
- 46. Use of the phytase variant of any of claims 1-34; or the 20 composition of any of claims 42-43 for liberating phosphorous from a phytase substrate.
- 47. A transgenic plant or plant part which is capable of expressing a phytase variant according to any one of claims 1-25 34.

	Peniophora number	's 1				
	Alignment numbers	1				37
	P_involtus Al		I. ECHINA A	T 0107772		50
	5 P_involtus_A2		H T.GEVALACI	L SLSEVLATS	V PKN	r aptfpiæse
	T_pubescens		MARCHIACI	I HLSEVFAAS	V PRN	I APKFSIESE
	A pediades		MST.ETCCCT	L FVCYAYARA	V PRAHIPLED	r sacldvirdy
	P_lycii		V SSAFADETT	L VFLQASAYG L SLMSSLALS	VVQATEVQP	FPQI
	A_fumigatus		. TELLCARVI	L .SGRVSAAP	r Qrsr	AAQLPIPAQN
10	consphyA		TELLISARII	F GSTSGTALG	SAGSKSCDT	DLGYQCSPAT
•	A_nidulans		TVALCIVVI	T. CDVCACAL	RGNSHSCDT	DGGYQCFPEI
	A_ficuum_NRRL3135	MCSVS	: IVAUSUIIU	LSRVSAQAI S GVTSGLAVPI	VVQNHSCNTA	DGGYQCFPNV
	A_terreus	MGFI	ATVICTATION	F RSTSGTPLG	A SKNOSSCOT	DOGYOCKSET
	T_thermo	MST	TIMECULA	r kologirldi	RGRHSDCNSV	DHGYQCFPEL
15	T_lanuginosa	MAGTGT.GSET	. VILLOPEAT	A LYVSRNI	HVDSHSCNTV	EGGYQCRPEI
	M_thermophila	MTGI	. COMMONICE	L TASPAIPPFV	RKKHPNVD	I
			GVMVVMVGE.	L AIASL	QSESRPCDTF	DLGFQCGTAI
		38				
		51				83
20	P_involtus A1	QRNWSPYSPY	FPLAEVKA	PPAGCQIN		100
	P_involtus_A2	QRNWSPYSPY	FPLAEVKA	PPAGCEIN	OWITCHER	REPTSGATTR
	T_pubescens	QOSWSMYSPY	FPAATYVA	PPASCOIN	WEITAGET TOTAL	REPUSCANTE
	A pediades	ODSWAAYTPY	VPVOAVTD	PPKDCKIT	OMITTORNO	REPTSGAAKR
	P lycii	TSNWGPYDPF	FPVEDVAA	PPEGCTVT	OWILTORIGA	REPTSGAGTR
25		SHLWGOYSPF	FSLEDELSVS	S SKLPKDCRIT	T.VOVI. CDUCA	RWPTSGARSR
	consphyA	SHLWGOYSPY	FSLEDESAT	PDVPDDCRVT	ENOM: CORGA	RIPISSKSKK
	A_nidulans	SHVWGOYSPY	FSIEGESATS	EDVPHGCEVT	FUOUL SPUCA	RIPISSKSKA
	A_ficuum_NRRL3135	SHLWGOYAPF	FSLANESVIS	PEVPAGCRVT	FAOVI.SPHGA	RIPIESASAA
	A_terreus	SHKWGLYAPY	FSLODESPFE	LDVPEDCHIT	FVOVI.APHGA	DCDTECTOR
30	T_thermo	SHSWGQYSPF	FSLADOSEIS	PDVPQNCKIT	FVOLLSPHGA	DVDTCCVTCT
	T_lanuginosa	ARHWGQYSPF	FSLAEVSEIS	PAVPKGCRVE	FVOVISRHGA	PVDTAUKEET
•	$M_{thermophila}$	SHFWGQYSPY	FSVPSELI	ASIPDDCEVT	FAOVLSRHGA	RAPTLKRAAS
		84				133
35	•	101				150
	P_involtus_A1	IKAGLTKLQG	VQNFTDAKFN	FIKSFKYDLG	NSDLVPFGAA	
	P_involtus_A2	IKAGLSKLQS	VQNFTDPKFD	FIKSFTYDLG	TSDLVPFGAA	OSFDAGLEVF
	${ t T}$ pubescens	IQTAVAKLKA	ASNYTDPLLA	FVTNYTYSLG	QDSLVELGAT	OSSEAGORAF
	A_pediades	IQAAVKKLQS	AKTYTOPRLD	FLTNYTYTLG	HDDLVPFGAL	QSSOAGEETF
40	P_lycii	QVAAVAKIQM	ARPFTDPKYE	FLNDFVYKFG	VADLLPFGAN	OSHOTGTDMY
	${ t A_fumigatus}$	YKKLVTAIQA	NATDFKGKFA	FLKTYNYTLG	ADDLTPFGEQ	OLVNSGIKFY
	consphyA	YSALIEAIQK	NATAFKGKYA	FLKTYNYTLG	ADDLTPFGEN	OMVNSGIRFY
	A_nidulans	YSGLIEAIQK	NATSFWGQYA	FLESYNYTLG	ADDLTIFGEN	OMVDSGAKFY
	A_ficuum_NRRL3135	YSALIEEIQQ	NATTFDGKYA	FLKTYNYSLG	ADDLTPFGEO	ELVNSGIKFY
45	A_terreus	YAATIAAIQK	SATAFPGKYA	FLQSYNYSLD	SEELTPFGRN	OLRDLGAOFY
	$\mathtt{T_thermo}$	YSQLISRIQK	TATAYKGYYA	FLKDYRYQLG	ANDLTPFGEN	OMIOLGIKEY
	T_lanuginosa	YAELLQRIQD	TATEFKGDFA	FLRDYAYHLG	ADNLTRFGEE	OMMESGROFY
	$M_{thermophila}$	YVDLIDRIHH	GAISYGPGYE	FLRTYDYTLG	ADELTRTGOO	OMVNSGIKFY
50		134				176
		151.				200
	P_involtus_A1	arysklvskn	NLPFIRADGS	DRVVDSATNW	TAGFASA	SHNTVO
	P_involtus_A2	ARYSKL v ssd	NLPFIRSDGS	DRVVDTATNW	TAGFASA	SRNAIO

	\mathtt{T} _pubescens	TRYSSLVSAD	ELPFVRASGS	DRVVATANNW	TAGFALA	SSNSIT
	A_pediades	QRYSFLVSKE	NLPFVRASSS	NRVVDSATNW	TEGFSAA	SHHVLN
	P_lycii	TRYSTLFEGG	DVPFVRAAGD	QRVVDSSTNW	TAGFGDA	SGETVL
	A_fumigatus	QRYKAL.ARS	VVPFIRASGS	DRVIASGEKF	IEGFQQAKLA	DPGA.TNRAA
5	consphyA	RRYKAL.ARK	IVPFIRASGS	DRVIASAEKF	IEGFQSAKLA	DPGSQPHQAS
	A_nidulans	RRYKNL.ARK	NTPFIRASGS	DRVVASAEKF	INGFRKAQLH	DHGSKRAT
	A_ficuum_NRRL3135	QRYESL.TRN	IVPFIRSSGS	SRVIASGKKF	IEGFQSTKLK	DPRAQPGQSS
	A_terreus	ERYNAL.TRH	INPFVRATDA	SRVHESAEKF	VEGFQTARQD	DHHANPHQPS
	T thermo	NHYKSL.ARN	AVPFVRCSGS	DRVIASGRLF	IEGFQSAKVL	DPHSDKHDAP
10	T_lanuginosa	HRYREQ.ARE	IVPFVRAAGS	ARVIASAEFF	NRGFQDAKDR	DPRSNKDQAE
	M thermophila				TQGFHSALLA	
			* **			
15	•	177				217
		201			•	250
	P involtus A1		G. NOTLEDN	MCPAAGD	SDPQVNA	
	P involtus A2	-		MCPAAGE		WLASAFPSVT
	T_pubescens	- · -		MCPAAGD		WLAQFAPPMT
20	A pediades		LNDTLDDA			WTSIYGTPIA
	P_lycii			MCPNEVD		WLGVFAPNIT
	A_fumigatus			VCTKFEA		
	consphyA				SELGDDVEAN	
	A nidulans				DERADEIEAN	
25	A_ficuum_NRRL3135				SELADIVEAN	
	A_ficadam_NRCH3133				STVGDDAVAN	
	T thermo				SSGGHDAQEK	
	T lanuginosa			TCPAAEE		
	M_thermophila				STIGDDAQDT	
30	w_cnermoburia	PIDMA ATERI	AGAINTIIAND	ICIAPEEGF I	BIIODDAQDI	1201110111
30		218				252
		251				300
	P_involtus_A1		NIT PERSONNEL CONTR.	VCI.CAPI.TVS	KEKK	
	P involtus A2	ARLINAMAPSV	MITIDIDAENT	VOLCAPILIVE	KEQK	
35	T pubescens	AQLINAAAPGA	MUTDADATNI	ASHCELMIAN	TERR	S
23	A pediades	ARLMAGAPGA	METOTOLIME	THUCFFELVA	KETP	S
•		NRLINQQAPGA	MITAADVSML	THICARETTA	SGNA	
	P_lycii A fumigatus				RTSDASQ.	
		ARAEKHLPGV	TETDEDVVSL	MDMCSEDIVA	RTSDATE.	T.C
40	consphyA				RTAHGTE.	
40	A_nidulans					
	A_ficuum_NRRL3135	ÖKTENDIZGA	TETTTEVIXL	MDMCSEDIIS	TSTVDTK.	
	A_terreus	QRLEADLPGV	OTS. LDDA ANT	MAMCPRETVS	LTDDAHT.	
	T_thermo				RNHTDT	
	T_lanuginosa				SDPVLFPRQ.	
45	M_thermophila	ARVNANLPGA	NLTDADTVAL	MDLCPFETVA	SSSSDPATAD	AGGGNGRPLS
		_				200
		253		•		300
		301				350
-	P_involtus_A1	DFCTLFEGIP	GSFEAFAYGG	DLDKFYGTGY	GQELGPVQGV	GYVNELLARL
50	P_involtus_A2	DFCTLFEGIP	GSFEAFAYAG	DLDKFYGTGY	GQALGPVQGV	GYINELLARL
	T_pubescens	EFCDIYEELQ	AE.DAFAYNA	DLDKFYGTGY	GQPLGPVQGV	GYINELIARL
	A_pediades	PFCNLFTP	EEFAQFEYFG	DLDKFYGTGY	GQPLGPVQGV	GYINELLARL
	P_lycii	PFCDLFTA	EEYVSYEYYY	DLDKYYGTGP	GNALGPVQGV	GYVNELLARL
	${ t A_fumigatus}$	PFCQLFTH	NEMKKANATÖ	SLGKYYGYGA	GNPLGPAQGI	GFTNELIARL
55	consphyA	PFCALFTH	DEWRQYDYLQ	SLGKYYGYGA	GNPLGPAQGV	GFANELIARL
	A_nidulans	PFCAIFTE	KEMIOADAIO	SLSKYYGYGA	GSPLGPAQGI	GFTNELIARL

	A_ficuum_NRRL3135	PECDLET U DEVILORE CONTROL OF THE PERSON OF
	A_terreus	
	T_thermo	
	T_lanuginosa	
	5 M_thermophila	TOWNS TO THE TENED TO SERVICE TO THE TRANSPORT OF THE TRA
	crrcrmoburts	PFCRLFSE SEWRAYDYLQ SVGKWYGYGP GNPLGPTQGV GFVNELLARL
		301
		351 349
	P_involtus A1	400
10	P_involtus_A2	ATTEMMENT ATTEMMENT AND PRODUCT OF
	T_pubescens	INS.AVNONT QINRILDAAP DIFPLNKIMY ADFSHONLMV AVESAMGIED
	A_pediades	THE STATE OF EIFFLINKILLY ADESHONOMY ATESAMOTEM
	P_lycii	ATTENDED BITTELDESIY ADLISHDOMI ATTENDED
	A_fumigatus	A THE PROPERTY AND SHOWING PINALITY AND SHOWING PINALITY AND SHOWING
15	consphyA	TRS.PVQDHT STNSTLVSNP ATFPLNATMY VDFSHDNSMV STFFALGLYN
	A_nidulans	TRS.PVQDHT STNHTLDSNP ATFPLNATLY ADFSHDNSMI SIFFALGLYN
	A_ficuum_NRRL3135	TQS.PVQDNT STNHTLDSNP ATFPLDRKLY ADFSHDNSMI SIFFAMGLYN
		THS.PVHDDT SSNHTLDSSP ATFPLKSTLY ADFSHDNGIT STLEALGLYN
	A_terreus	TRA. PVHDHT CVNNTLDASP ATFPLNATLY ADFSHDSNLV STFWALGLVN
20	T_thermo	THS.PVQDYT TVNHTLDSNP ATFPLNATLY ADFSHONTMT STEALGLYN
		TGNLPVKDHT TVNHTLDDNP ETFPLDAVLY ADFSHDNTMT GIFSAMGLVN
	M_thermophila	A.GVPVRDGT STNRTLDGDP RTFPLGRPLY ADFSHDNDMM GVLGALGAYD
		•
25		350 383
		401 450
	P_involtus_A1	QPAPLSTSVP NPWRT WRTSSLVPFS GRMVVERLSC
	P_involtus_A2	QSAPLSTSTP DPNRT WLTSSVVPFS ARMAVERLSC
	T_pubescens	QSAPLDPTTP DPART FLVKKIVPFS ARMVVERLDC
30	A_pediades	QSSPLDPSFP NPKRT WVTSRLTPFS ARMVTERLLC QRDGTGSGGP
	P_lycii	ATA.LDPLKP DENRL WVDSKLVPFS GHMTVEKLAC
	A_fumigatus	GTEPLSRTSV ESAKELDG YSASWVVPFG ARAYFETMQC
	consphyA	GTAPLSTTSV ESIEETDG YSASWTVPFG ARAYVEMMQC
	A_nidulans A_ficuum_NRRL3135	GTQPLSMDSV ESIQEMDG YAASWTVPFG ARAYFELMQC
35		GTKPLSTTTV ENITQTDG FSSAWTVPFA SRLYVEMMQC
~~	A_terreus	GTAPLSQTSV ESVSQTDG YAAAWTVPFA ARAYVEMMQC
	T_thermo	GTAKLSTTEI KSIEETDG YSAAWTVPFG GRAYIEMMQC
	T_lanuginosa	GTKPLSTSKI QPPTGAAADG YAASWTVPFA ARAYVELLRC ETETSSEEEE
	$M_{thermophila}$	GVPPLDKTAR RDPEELGG YAASWAVPFA ARIYVEKMRC SGGGGGGGGG
40		
- 10		384 425
	B 4	451 500
	P_involtus_A1	FGT TKVRVLVQDQ VQPLEFCGGD RNGLCTLAKF VESQTFARSD
	P_involtus_A2	AGT TKVRVLVQDQ VQPLEFCGGD QDGLCALDKF VESOAYARSG
45	T_pubescens	GGA QSVRLLVNDA VQPLAFCGAD TSGVCTLDAF VESOAYARND
30	A_pediades	SRIMRNGNVQ TFVRILVNDA LQPLKFCGGD MDSLCTLEAF VESOKYARED
	P_lycii	SGK EAVRVLVNDA VQPLEFCGG. VDGVCELSAF VESQTYAREN
	A_fumigatus	KSEKE PLVRALINDR VVPLHGCDVD KLGRCKLNDF VKGLSWARSG
	consphyA	QAEKE PLVRVLVNDR VVPLHGCAVD KLGRCKRDDF VEGLSFARSG
50	A_nidulans	EKKE PLVRVLVNDR VVPLHGCAVD KFGRCTLDDW VEGLNFARSG
JU .	A_ficuum_NRRL3135	QAEQA PLVRVLVNDR VVPLHGCPVD ALGRCTRDSF VRGLSFARSG
	A_terreus	R.AEKE PLVRVLVNDR VMPLHGCPTD KLGRCKRDAF VAGLSFAOAG
	T_thermo	D.DSDE PVVRVLVNDR VVPLHGCEVD SLGRCKRDDF VRGLSFAROG
	1_1anuginosa	EGEDE PFVRVLVNDR VVPLHGCRVD RWGRCRRDEW IKGLTFAROG
	$ exttt{M_thermophila}$	EGRQEKDE EMVRVLVNDR VMTLKGCGAD ERGMCTLERF IESMAFARGN
55		

		501	514
	P_involtus_A1	GAGDFEKCFA	TSA.
	P_involtus_A2	GAGDFEKCLA	TTV.
	T_pubescens	GEGDFEKCFA	т
5	A_pediades	GQGDFEKCFD	
	P_lycii	GQGDFAKCGF	VPSE
	A_fumigatus	GNWGECFS	
	consphyA	GNWAECFA	*
	A_nidulans	GNWKTCFT	L
10	A_ficuum_NRRL3135	GDWAECFA	
	A_terreus	GNWADCF.	
	T_thermo	GNWEGCYA	ASE.
	T_lanuginosa	GHWDRCF.	
	M_thermophila		
15			

(2	2) IR	FOR	MATIC	ON FO	R SE	Q II	NO:	25:									
	((i) :	(A) (B) (C)	ENCE LENG TYPE STRA TOPO	TH: : nu NDEE	1522 clei NESS	bas c ac : si	e pa id ngle	irs								
	(i	i) 1	OLEC	ULE	TYPE	: cD	NA.										
	(₩	i) c	(A)	NAL ORGA STRA	MEIN	: Pa			nvol	utus							
	(i	ж) Е		RE : NAME LOCA				3					-				
٠	(i.	-	EATU (A) (B)	re : Name Loca	/KEY	: ma :115	t_pe;	ptid 83	e	٠							
	(i.		EATU: (A) :	RE :. NAME , LOCA :	/KEY:	siq :58.	_pei	otide	•						•		
	(x:	L) s	EQUE	NCE I	DESCE	RIPT	ON:	SEQ	ID N	10: 2	25:						
GGI	ATCC	TAAE	TCG	CACI	cg 1	ACG	STCC	C C	GTCI	ACCC	TC1	GCT	GCC	TTG	GAAG		57
ATO Met	Let	TTO Pho	C GG1	TTC / Phe -15	. Val	GCC Ala	CTC Lev	GCC Ala	TGI Cys -10	Leu	TTO Lev	TCC Ser	CTC	TCC Sei	GAG Glu		105
GTC Val	CTI Leu	GCC Ala	ACC Thr	Ser	GTG Val	CCC Pro	AAG Lys 5	ne A	ACA Thr	GCG Ala	Pro	ACC Thr	Phe	CCC Pro	ATT Ile		153
CCG Pro	GAG Glu 15	Ser	GAG	CAG Gln	CGG	AAC Asn 20	Trp	TCC	CCG	TAC	TCG Ser 25	Pro	TAC Tyr	TTC Phe	CCT Pro		201
CTT Leu 30	Ala	GAG Glu	TAC	AAG Lys	GCT Ala 35	CCT	CCG Pro	GCG	GGC	TGC Cys 40	CAG Gln	ATC Ile	OAA neA	CAG Gln	GTC Val 45		249
AAC Asn	ATC	ATC Ile	CAA Gln	AGA Arg 50	TAT His	GL Y	GCC Ala	CGG Arg	TTC Phe 55	CCG Pro	ACC Thr	TCT Ser	GGC	GCG Ala 60	ACC Thr		297
ACC Thr	CGT Arg	ATC Ile	AAG Lys 65	GCG Ala	GGT Gly	TTG Leu	ACC Thr	AAG Lys 70	TTG Leu	CAA Gln	GGC Gly	GTC Val	CAG Gln 75	AAC Asn	TTT Phe	•	345
ACC Thr	GAC Asp	GCC Ala 80	AAA Lys	TTC Phe	AAC Asn	TTC Phe	ATC Ile 85	AAG Lys	TCG Ser	TTC Phe	AAG Lys	TAC Tyr 90	GAT Asp	CTC Leu	GGT Gly		393
AAC Asn	TCG	gyc Gyc	CTC Leu	GTT Val	CCG Pro	TTC Phe	GGT Gly	GCA Ala	GCA Ala	CAG Gln	TCC Ser	TTC Phe	GAC Asp	GCT Ala	GGT Gly		441

105

100

CAG Gln 110	GAG Glu	GCC Ala	TTC Phe	GCC Ala	CGC Arg 115	TAC Tyr	TCG Se≍	AAG Lys	CTT Leu	GTC Val 120	AGC Ser	AAG Lys	AAC Asn	AAC Asn	CTG Leu 125	489
CCG Pro	TTC Phe	ATT Ile	CGT Arg	GCC Ala 130	TAƏ Qe <i>A</i>	GGA Gly	AGT Ser	TAD qeA	CGT Arg 135	GTT Val	GTG Val	GAT Asp	TCT Ser	GCT Ala 140	ACA Thr	537
AAC Asn	TGG Trp	ACT Thr	GCG Ala 145	GGT Gly	TTC Phe	GCT Ala	TCG Ser	GCA Ala 150	AGT Ser	CAC EiH	AAC Asn	ACG Thr	GTC Val 155	CAG Gln	CCC Pro	585
AAG Lys	CTG Leu	AAC Asn 160	CTG Leu	ATT Ile	CTC Leu	CCG Pro	CAA Gln 165	ACT	GCC	TAA Taa	ÇAT Asp	ACC Thr 170	CTG Leu	GAA Glu	GAT Asp	633
AAT Asn	ATG Met 175	Суз	CCT Pro	GCT Ala	GCT Ala	GGC Gly 180	TAD QEA	TCT Ser	GAC Asp	CCC	CAG Gln 185	Val	AAC Asn	GCG Ala	TGG Trp	681
TTG Leu 190	Ala	GTT Val	GCT Ala	TTC	CCT Pro 195	TCC Ser	ATC Ile	ACT	GCA Ala	CGG Arg 200	Leu	AAC Asn	GCC	GCC	GCG Ala 205	729
Pro	TCT Ser	GTC Val	AAC Asn	CTC Leu 210	ACC Thr	GAC Asp	ACG Thr	GAC Asp	GCG Ala 215	Phe	AAC Asn	CTC Leu	GTC Val	AGT Ser 220	Leu	777
Суз	Ala	Phe	Leu 225	Thr	Val	Ser	ГЛЗ	Glu 230	Lys	Lys	Ser	qeA :	235	. Cys	Thr	825
Leu	Phe	240	Gly	Ile	Pro	Gly	Ser 245	Phe	Glu	Ala	. Phe	250	Tyr)	. G17	, Gly	873
Asp	255	a Asp	Lys	.Phe	Tyr	G1y 260	Thr	: Gly	Tyr	: Gl	265 265	ı Glu	ı Lev	7 GT?	CCC Pro	921
GT1 Val 270	. Glr	A GGC	GTC Val	GLY	TAC Tyr 275	Val	AAC TEA	GAG Glu	CTC	280	a Ala	C CGC	CTC Lev	Th:	AAC Asn 285	969
Sei	: Ala	a Vai	l Arg	290	Asn	Thi	: Glr	1 Thi	295	Ar	g Th:	r Lei	ı Ası	30°		1017
Pro	C GT	A ACC	r Phe	Pro	TTO Lev	AAG 1 Asi	AAC Ly:	ACC Thi	Phe	TA	c GC	C GA' a As	P Ph	e Se	C CAC	
GA(AA C	C CT	u Met	GT(C GCC	C GTO	2 TTC 1 Pho 32	e Se	C GCC	AT Me	G GG t Gl	C CT y Le 33	u Ph	C CG e Ar	c CAG g Gln	1113
Pr	G GC Al 33	a Pr	G CTO	C AGO	C ACC	TC: Se:	r Va	G CCG	AA E EA c	c CC	A TG	p Ar	C AC g Th	G TG	p Arg	1161

ACG Thr 350	AGC Ser	TCC Ser	CTC Leu	GTC Val	CCC Pro 355	TTC Phe	TCC Ser	GGA Gly	CGC	ATG Met 360	GTC Val	GTG Val	GAA Glu	CGC Arg	CTC Leu 365		1209
AGC Ser	TGT Cys	TTC Phe	GGC	ACG Thr 370	ACC	AAG Lys	GTT Val	CGC Arg	GTC Val 375	CTC Leu	GTG Val	CAG Gln	GAC Asp	CAG Gln 380	GTG Val		1257
CAG Gln	CCG Pro	CTC Leu	GAG Glu 385	TTC Phe	TGC Cys	GGG Gly	GGT Gly	GAT Asp 390	AGG Arg	OAA neA	GGG GLy	CTG Leu	TGC Cya 395	ACG Thr	CTT Leu		1305
GCT Ala	AAG Lya	TTT Phe 400	GTG Val	GAG Glu	AGC Ser	CAG Gln	ACG Thr 405	TTT Phe	GCG Ala	AGG Arg	AGT Ser	GAT Asp 410	GGT Gly	GCG Ala	GGG		1353
GAC Asp	TTT Phe 415	GAG Glu	AAG Lys	TGC . Cys	TTC Phe	GCG Ala 420	ACC Thr	TCG Ser	GCG Ala	TGAG	GATG	GA C	GAAC	AAAA	T		1403
TAAA	TTGG	GG I	'ATTT	TATO	G TA	TAAT	TATG	GTG	TGTG	TAG	AACA	TGGG	CT C	GGGG	TCGAT	•	1463
GGTG	AAAA	GC A	AAGG	TTTA	T CG	TCTA	AAAA	. AAA	AAAA	AAA	AAAA	AATT	CC I	GCGG	CCGC		1522

8/51

(2)	INFORMATION	FOR	SEO	ID	NO:	27:

(2)	INFO	RMAT	NOI	FOR	SEQ	ID N	0: 2	7:								
	(i)	(E	QUENC L) LE L) TY C) SI	ngth Pe: Rand	: 16 nucl EDNE	42 b eic SS:	ase acid sing	pair	s							
	(ii)	MOI	ECUI	E TY	PE:	CDNA										
	(vi)	•		IGANI	SM:	Paxi			rolut	us						
	(ix)	_	LTURE L) NA L) LO	me\k			.373									
	(ix)	FE <i>I</i> (<i>I</i>	ATURE A) NA B) LO	ME/K	EY: ON:1	mat_ .05	pept 1373	ide								
	(ix)		ATURE A) NI B) LO	ME\K				ide:								
	(xi)	SEC	QUENC	CE DE	SCRI	PTIC	on: s	EQ 1	D NO): 27	/·:					
GGA?	rccga	AT 1	CCAC	STCC	C AF	\GCT <i>I</i>	ATC	TCI	rgcto	GCC	TTG	SAAG		CAC His	CTC Leu	56
GLY GGC	TTC Phe -15	GTC Val	ACC Thr	CTC Leu	GCT Ala	TGT Cys ~10	CTC Leu	ATA Ile	CAC His	CTC Leu	TCC Ser -5	GAG Glu	GTC Val	TTC Phe	GCG Ala	104
GCA Ala 1	TCC Ser	GTG Val	CCC Pro	CGG Arg 5	AAT Asn	ATT Ile	GCT Ala	CCG Pro	AAG Lys 10	TTC Phe	TCA Ser	ATT Ile	CCG Pro	GAA Glu 15	AGC Ser	152
GAG Glu	CAG Gln	CGA Arg	AAC Asn 20	TGG Trp	TCG Ser	CCT Pro	TAC Tyr	TCT Ser 25	CCT Pro	TAC Tyr	TTT Phe	CCC Pro	CTA Leu 30	GCC Ala	GAA Glu	200
TAC Tyr	AAG Lys	GCT Ala 35	CCT Pro	CCA Pro	GCA Ala	GCC	TGC Cys 40	GAG Glu	ATT	AAC Asn	CAA Gln	GTC Val 45	ÄAT Asn	ATT Ile	ATC Ile	248
CAA Gln	CGG Arg 50	CAT His	GLY	GCA Ala	CGG Arg	TTC Phe 55	CCA Pro	ACC Thr	TCG Ser	GGT Gly	GCG Ala 60	GCC Ala	ACT Thr	CGC Arg	ATC Ile	296
AAG Lys 65	GCT Ala	GGT Gly	TTA Leu	AGC Ser	AAG Lys 70	CTG Leu	CAA Gln	TCC Ser	GTC Val	CAG Gln 75	AAT Asn	TTC Phe	ACC Thr	GAC qeA	CCC Pro 80	344

392

95

90

AAA TTC GAC TTC ATC AAG TCG TTC ACA TAC GAT CTT GGT ACT TCC GAC

Lys Phe Asp Phe Ile Lys Ser Phe Thr Tyr Asp Leu Gly Thr Ser Asp

85

CTC Leu	GTG Val	CCA Pro	TTC Phe 100	GGC Gly	GCA Ala	GCA Ala	CAA Gln	TCA Ser 105	TTC Phe	GAT Asp	GCC Ala	GGC Gly	CTG Leu 110	GAG Glu	GTC Val	440
TTC Phe	GCT Ala	CGC Arg 115	TAT Tyr	TCG Ser	AAG Lys	CTC Leu	GTC Val 120	AGC Ser	TCG Ser	GAC Asp	AAC Asn	CTG Leu 125	CCT Pro	TTC	ATT	488
CGC Arg	TCA Ser 130	GAT Asp	GGT Gly	AGC Ser	GAT Asp	CGT Arg 135	GTA Val	GTC Val	GAC Asp	ACT Thr	GCT Ala 140	ACG Thr	AAC Asn	TGG Trp	ACT Thr	536
GCA Ala 145	ggt Gly	TTT Phe	GCT Ala	TCC Ser	GCG Ala 150	AGC Ser	CGC Arg	AAC Asn	GCG Ala	ATC Ile 155	CAA Gln	CCC	AAG Lys	CTC Leu	GAC Asp 160	584
											GAG Glu					632
											GCG Ala					680
											GCA Ala					728
											AGC Ser 220					776
											Cys					824
											GCC Ala					872
											GGA Gly					920
											ACC Thr					968
											GCC Ala 300					1016
											TCA Ser					1064
											CGC Arg					1112

Fig 3P

								AAC Asn 345								1160
															GCT Ala	1208
								GTG Val							CTC Leu	1256
								GGG Gly								1304
								AGT Ser								1352
		CTT Leu					TGAC	EATGO	igg 1	'AATO	TACG	G T	BAAGO	CAGCO	3	1403
GAGA	GCCI	CT (CAACO	AATO	C A	AGGA	TAGO	TTC	GAGG	CTT	ACTI	CATO	CAA C	CTAI	CATCAT	1463
CATA	GGAC	CAA C	CCC	CCAP	T AG	CCAC	ACTO	GTC	GTTI	GAC	ATC	etgti	ATG I	TAAA!	'AACCC	1523
ACCC	ACGC	AC 1	CCGC	CTGCC	A CI	TATTO	CGCGI	GTA	TCGC	ATA	CTAG	GCGT	TT I	rcgcc	CAGTT	1583
SAAC	ATG	AGC (CATI	CTGI	c co	CAGI	GAAF	AAA A	AAAA	AAA	AAAF	AATT	rcc 1	CGCGG	CCGC	1642

447

11/51

(2)	INF	ORMA'	TION	FOR	SEQ	ID :	NO:	29:								
	<u>(</u> 1	()	A) L B) T C) S	engt: Ype : Tran	HARACH: 1. nuc. DEDNI OGY:	536 leic ESS:	base aci sin	pai d	rs							
	(11) MO:	LECU	LE T	YPE:	CDN	A									
	(vi		A) 0	RGAN		Tra			besc	ens						
	(ix		A) N	AME/	ION:		1407									
	(ix		A) N	AME/	KEY:											
	(ix		A) N	AME/	KEY:			tide								
	(xi) SE	QUEN	CE D	escr:	[PTI	ON:	SEQ	ID N	0: 2	9:					
GGA	TCCG	TAA	rcgc	ccca	AC A	rtcgʻ	TTCC	A TC	TTAG	CAGC	CGT	CCGC	GCC	CAGG:	rcttc	G 60
	TCCG.				ATG	GCC Ala	TTC	TCA		TTG	GCC	TCG	CTG	CTC	TTC	2G 60 111
ATA GTG		TAT Tyr	GCA	gact Tac	ATG Met -17 GCC	GCC Ala AGG	TTC Phe -15	TCA Ser	ATC Ile	TTG Leu CGT	GCC Ala GCA	TCG Ser -10	CTG Leu ATC	CTC Leu CCG	TTC Phe	
ATA GTG Val	ACCC TGT Cys	TAT Tyr	GCA Ala TCC	TAC Tyr GCG	ATG Met -17 GCC Ala	GCC Ala AGG Arg 1	TTC Phe -15 GCT Ala	TCA Ser GTG Val	ATC Ile	TTG Leu CGT Arg 5	GCC Ala GCA Ala	TCG Ser -10 CAT His	CTG Leu ATC Ile	CTC Leu CCG Pro	TTC Phe CTC Leu 10	111
GTG Val CGC Arg	TGT Cys -5 GAC	TAT Tyr ACC Thr	GCA Ala TCC Ser	TAC Tyr GCG Ala 15	ATG Met -17 GCC Ala TGT Cys	GCC Ala AGG Arg 1 CTA Leu	TTC Phe -15 GCT Ala GAT Asp	TCA Ser GTG Val GTA Val	ATC Ile CCC Pro ACA Thr 20 GCA	TTG Leu CGT Arg 5 CGC Arg	GCC Ala GCA Ala GAT Asp	TCG Ser -10 CAT His GTG Val	CTG Leu ATC Ile CAG Gln	CTC Leu CCG Pro CAG Gln 25	TTC Phe CTC Leu 10 AGC Ser	111
GTG Val CGC Arg	TGT Cys -5 GAC Asp	TAT Tyr ACC Thr ATG Met	GCA Ala TCC Ser TAC Tyr 30	TAC Tyr GCG Ala 15 TCT Ser	ATG Met -17 GCC Ala TGT Cys	GCC Ala AGG Arg 1 CTA Leu TAT Tyr	TTC Phe -15 GCT Ala GAT Asp TTC Phe	TCA Ser GTG Val GTA Val CCG Pro 35	ATC Ile CCC Pro ACA Thr 20 GCA Ala	TTG Leu CGT Arg 5 CGC Arg GCA Ala	GCC Ala GCA Ala GAT Asp ACT Thr	TCG Ser -10 CAT His GTG Val TAT Tyr	CTG Leu ATC Ile CAG Glñ GTG Val 40	CTC Leu CCG Pro CAG Gln 25 GCT Ala	TTC Phe CTC Leu 10 AGC Ser CCG Pro	111 159 207
GTG Val CGC Arg TGG Trp CCC Pro	TGT Cys -5 GAC Asp TCC Ser	TAT Tyr ACC Thr ATG Met AGT Ser 45	GCA Ala TCC Ser TAC Tyr 30 TGC Cys	TAC Tyr GCG Ala 15 TCT Ser CAG Gln	ATG Met -17 GCC Ala TGT Cys CCC Pro ATC Ile	GCC Ala AGG Arg 1 CTA Leu TAT Tyr AAT Asn	TTC Phe -15 GCT Ala GAT Asp TTC Phe CAG Gln 50 GCA	TCA Ser GTG Val GTA Val CCG Pro 35 GTC Val	ATC Ile CCC Pro ACA Thr 20 GCA Ala CAC His	CGT Arg 5 CGC Arg GCA Ala ATC Ile CGC	GCC Ala GCA Ala GAT Asp ACT Thr	TCG Ser -10 CAT His GTG Val TAT Tyr CAA Gln 55	CTG Leu ATC Ile CAG Glñ GTG Val 40 CGT Arg	CTC Leu CCG Pro CAG Gln 25 GCT Ala	TTC Phe CTC Leu 10 AGC Ser CCG Pro GGT Gly	111 159 207 255

GTT ACG AAC TAC ACC TAC AGC TTA GGT CAG GAC AGC CTC GTT GAA CTC Val Thr Asn Tyr Thr Tyr Ser Leu Gly Gln Asp Ser Leu Val Glu Leu 95 100 105

GT Gly	GCG Ala	ACT Thr	CAG Gln 110	TCC Ser	TCC (Ser (GAA (Glu)	Ala	GGC Gly 115	CAG Gln	GAG Glu	GCA Ala	TTC Phe	Thr 120	Arg	Tyr	4,55
TCA Ser	TCC Ser	CTC Leu 125	GTG Val	AGC Ser	GCG Ala	qeA	GAG Glu 130	CTT Leu	CCC Pro	TTC Phe	GTT Val	CGG Arg 135	GCG Ala	TCG Ser	GGC Gly	543
TCA Ser	GAT Asp 140	CGC Arg	GTC Val	GTT Val	GCG Ala	ACT Thr 145	GCC Ala	AAC Asn	DAA neA	TGG Trp	ACT Thr 150	GCA Ala	ggt Gly	TTC Phe	GCG Ala	591
CTT Leu 155	GCG Ala	AGC Ser	TCA Ser	AAC Asn	AGC Ser 160	ATC Ile	ACG Thr	CCC Pro	GTG Val	CTC Leu 165	TCA Ser	GTC Val	ATC Ile	ATT Ile	TCC Ser 170	639
GAA Glu	GCG Ala	GGC	TAA neA	GAC Asp 175	ACC Thr	CTC Leu	GAC Asp	GAC Asp	AAC Asn 180	ATG Met	TGC	CCC	GCT Ala	GCA Ala 185	GGC	687
GAT Asp	TCG Ser	GAT Asp	CCC Pro	Gln	GTC Val	AAT Asn	CAA Gln	TGG Trp 195	Leu	GCG Ala	CAG Gln	TTC Phe	GCA Ala 200	FIC	CCG Pro	735
ATG Met	ACT Thr	GCT Ala 205	Arg	CTC Leu	AAC Asn	GCA Ala	GGC Gly 210	Ala	CCC Pro	GGC Gly	GCG Ala	AAC Asn 215	Leu	ACG Thr	GAC Asp	783
ACG	GAC Asp 220	Thi	TAC	CAA :	CTG Leu	CTC Leu 225	Tha	CT?	A TGC 1 Cys	CCG	TTC Phe 230	יותם פ	ACT Thr	GTA	GCC Ala	831
ACC Thi	Glu	G CGC	G CGT	r AGT g Ser	GAP Glu	ı Phe	TGC Cys	CAD C	C ATO	TAC Ty:	GIL	G GAG	CTC	G CAC	G GCG Ala 250	· 879
GA! Glu	AS A	GCC Ala	C TTO	C GCG a Ala 255	ту:	CAA C	GCC Ala	C GA	T CTO p Len 260	ı Ası	D AAG	S TTO	TAC Ty	GG(G1) 26	C ACT y Thr 5	927
GGI Gl	A TAC y Ty:	c GG c Gl	C CA y Gl: 27	n Pro	C CTC	C GGI	A CC	C GT o Va 27	I GI	A GGC	C GT	C GGG	3 TAC y Ty: 28		C AAC e Asn	975
GA(G CT	C AT u Il 28	e Al	G CG a Ar	C CT	C AC u Th	C GC ± Al 29	a Gl	G AA n As	C GT	G TC 1 Se	C GA r As 29	b ur	OA O	G CAG r Gln	1023
AC Th	AA D eA r OE	n Se	C AC	A CT	C GA u As	C TC p Se 30	r Se	G CC	CC GA	G AC	G TT r Ph 31	e PI	G CT o Le	C AA u As	c ccc	1071
AC Th 31	r Le	C TA	C GC	G GA ea a.	C TT p Ph 32	e Se	G CF	C GI	AC AA	C CA in Gl 32	n Me	G GT	C GC	G AT	C TTC e Phe 330	1119
TC Se	G GC	C AT	rG GG	T CT Ly Le 33	u Ph	C AF	C CI	AG TO Ln So	er Al	CG CC La Pr	G CI	C GA	r Pr	G AC	G ACG ir Thr 15	116

CCC Pro	GAC Asp	CCC Pro	GCG Ala 350	CGC Arg	ACG Thr	TTC Phe	CTC	GTC Val 355	AAG Lya	AAG Lys	ATC Ile	GTG Val	CCG Pro 360	TTC Phe	TCC Ser	1215
GCG Ala	CGC Arg	ATG Met 365	GTC Val	GTC Val	GAG Glu	CGC Arg	CTC Leu 370	gac Asp	TGC Cys	GGC Gly	GGT Gly	GCG Ala 375	CAG Gln	AGC Ser	GTG Val	1263
CGC Arg	CTG Leu 380	CTC Leu	GTG Val	AAC Asn	GAC Asp	GCA Ala 385	GTG Val	CAG Gln	CCG Pro	CTG Leu	GCG Ala 390	TTC Phe	TGC Cys	GGG Gly	GCG Ala	1311
GAC Asp 395	ACG Thr	AGC Ser	GGG Gly	GTG Val	TGC Cys 400	ACĢ Thr	CTG Leu	GAC Asp	GCG Ala	TTT Phe 405	GTC Val	GAG Glu	AGC Ser	CAG Gln	GCG Ala 410	1359
TAC Tyr	GCG Ala	CGG Arg	neA	GAT Asp 415	GGC Gly	GAG Glu	GGC Gly	GAC Asp	TTC Phe 420	GAG Glu	AAG Lys	TGC Cys	TTC Phe	GCG Ala 425	ACA Thr	1407
TAGT	TCCA	GG I	'GTAG	ATAC	C CG	GGGA	AGAT	GTA	CTCI	CTA	GACA	CCTC	GC A	TGTA	CTTAT	1467
CGAT	TAGA	AA G	AGAC	CCTG	G CT	GCTC	TGCC	CTC	AAAA	AAA	AAAA	AAAA	AA A	AAAA	ATTCC	1527
TGCG	GCCG	C							٠.						•	1536

(2) INFORMATION FOR SEQ ID NO: 21:

	(i)	(A) (B) (C)	LENCE LEN TYP STF	IGTH: PE: 1 VANDI	: 150 Ducle EDNES	01 ba eic a SS: a	ase p acid sing:	pairs	ı								
((11)	MOL	ECULE	TY!	PE: (CDNA											
ı	(TV)	(A)	GINAI) ORC) STI	SAN I	SM: Z	Agro			Lades	1							
	(ix)	(A	TURE) NAI) LO	ME/K			375										
	(ix)	(A	TURE) NAI) LO	ME/K				ide			÷						
	(ix)	(A	TURE) NA) LO	ME/K				idė		•							
	(xi)	SEQ	UENC	E DE	SCRI	PTIC)N: S	EQ I	סמ ס	: 21	:						
GAT	CCGA	AT T	CACT	Me	G TC	r Le	C TI	C AT le Il	C GG e Gl	C GG y Gl -2	у Су	T TI	G CI	C GI	G 1	49	
rrr ?he -15	TTA Leu	CAG Gln	GCG Ala	AGC Ser	GCA Ala -10	TAC Tyr	GGC GLY	G] À GCC	GTC Val	GTG Val -5	CAG Gln	GCC Ala	ACA Thr	TTC Phe	GTG Val 1	97	
CAG Sln	CCG Pro	TTT Phe	TTC Phe 5	CCT Pro	CCA Pro	CAG Gln	ATT Ile	CAG Gln 10	GAC Asp	TCT Ser	TGG Trp	GCA Ala	GCT Ala 15	TAT Tyr	ACA Thr	145	
CCA Pro	TAT Tyr	TAT Tyr 20	CCT Pro	GTT Val	CAG Gln	GCG Ala	TAC Tyr 25	ACG Thr	CCT Pro	CCC Pro	Pro	AAG Lys 30	GAT Asp	TGC Cys	AAG Lys	193	,
ATC Ile	ACA Thr 35	CAA Gln	GTT Val	AAC Asn	ATT Ile	ATT Ile 40	CAA Gln	CGA Arg	CAT His	GGT Gly	GCC Ala 45	CGC Arg	TTT Phe	CCG Pro	ACA Thr	241	
TCG Ser 50	ejà egg	GCA Ala	GGC Gly	ACA Thr	AGG Arg 55	ATC Ile	CAA Gln	GCA Ala	GCT Ala	GTG Val 60	DAA Lys	AAG Lys	CTT	CAA Gln	TCA Ser 65	289)
GCT Ala	AAA EY3	ACC Thr	TAT Tyr	ACG Thr 70	GAT Asp	CCT Pro	CGT Arg	CTC	GAC Asp 75	Phe	CTG Leu	ACC	AAC Asn	TAT Tyr 80	ACC Thr	337	,
TAT Tyr	ACC Thr	CTT Leu	GGT Gly 85	CAC His	GAC Asp	GAT Asp	CTC Leu	GTA Val 90	CCG Pro	TTT Phe	GGA Gly	GCG Ala	CTT Leu 95	Gln	TCA Ser	385	3

									CGA Arg								433
									TCG Ser							•	481
									TTT Phe							!	529
									CTC Leu 155							!	577
									GCG Ala							ı	625
									ACG Thr							,	673
									ACA Thr							•	721
									ATA Ile							•	769
									GAG Glu 235							,	817
									ACA Thr								865
									TAA neA								913
ACA Thr	GAA Glu 275	ATG Met	CCA Pro	GTT Val	CGA Arg	GAT Asp 280	AAC Asn	ACC Thr	CAG Gln	ACG Thr	AAC Asn 285	AGG Arg	ACA Thr	CTC	GAC Asp		961
									CGC Arg							1	009
									TTT Phe 315							1	057
									TTC Phe							1	105

GG TP	GTC Val	ACC Thr 340	AGT Ser	CGG Arg	CTT	ACG Thr	CCT Pro 345	TTC Phe	AGC Ser	GCG Ala	AGA Arg	ATG Met 350	GTC Val	ACT Thr	GAG Glu	1153
:GG \rg	TTG Leu 355	CTG Leu	TGT Cys	CAA Gln	AGG Arg	GAT Asp 360	GGG	ACA Thr	GGG Gly	AGC Ser	GGT Gly 365	GGA GLy	CCA Pro	TCC Ser	AGG Arg	1201
ATC [le	ATG Met	CGG Arg	AAT Asn	GGA Gly	AAT Asn 375	GTG Val	CAG Gln	ACG Thr	TTT Phe	GTG Val 380	AGG Arg	ATT Ile	CTT Leu	GTC Val	AAC Asn 385	1249
TAE qe	GCT Ala	TTA Leu	CAG Gln	CCT Pro 390	TŤG Leu	AAG Lys	TTC Phe	TGC Cys	GGA Gly 395	GGG Gly	GAC qe <i>A</i>	ATG Met	TAD qeA	AGT Ser 400	TTG Leu	1297
rgr	ACT Thr	Leu	GAA Glu 405	Ala	TTC Phe	GTC Val	GAG Glu	AGC Ser 410	CAG Gln	AAG Lys	TAT Tyr	GCA Ala	CGA Arg 415	GAĞ Glu	GAT Asp	134
			Asp					Phe	GAT Asp		ATAT	TGC	agta	tgct	CA	139
GTG	AGTA	GAC	TACA	GTGC	AG G	CCCT	GTAA	C TC	TTGT	ATTG	TGT	TTCT	GGA	ATTC	CTCGGA	145
GCG	TAGT	TTG	TAGO	AAAA:	AA A	AAAA	AAAA	A AA	ATTC	CTGC	GGC	CGC				150

455

	17/51	
	(2) INFORMATION FOR SEQ ID NO: 23:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1593 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
	(vi) ORIGINAL SOURCE: (A) ORGANISM: Peniophora lycii (B) STRAIN: CBS 686.96	
	(ix) FEATURE: (A) NAME/KEY: sig_peptide (B) LOCATION:123212	
v	(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION:2131439	
	(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION:1231439	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:	
	GGATCCGAAT TCCATCTTCT GCTCTGACCT CCATCTCGCT GAGCGGCCGA CGAGAACCTA	60
	GGGGCTCTAA GTCCACGTAC TATCGCCGCG CCTGTGAAGG CCCCATACCA GCCCTTATCG	12
	AT ATG GTT TCT TCG GCA TTC GCA CCT TCC ATC CTA CTT AGC TTG ATG Met Val Ser Ser Ala Phe Ala Pro Ser Ile Leu Leu Ser Leu Met -30 -25 -20	16
·	TCG AGT CTT GCT TTG AGC ACG CAG TTC AGC TTT GTT GCG GCG CAG CTA Ser Ser Leu Ala Leu Ser Thr Gln Phe Ser Phe Val Ala Ala Gln Leu -15 -10 -5 1	21
	CCT ATC CCC GCA CAA AAC ACA AGT AAT TGG GGG CCT TAC GAT CCC TTC Pro Ile Pro Ala Gln Asn Thr Ser Asn Trp Gly Pro Tyr Asp Pro Phe 5 10 15	26:
,	TTT CCC GTC GAA CCG TAT GCA GCT CCG CCG GAA GGG TGC ACA GTG ACA Phe Pro Val Glu Pro Tyr Ala Ala Pro Pro Glu Gly Cys Thr Val Thr 20 25 30	31:
	CAG GTC AAC CTG ATT CAG AGG CAC GGC GCG CGT TGG CCC ACA TCC GGC Gln Val Asn Leu lle Gln Arg His Gly Ala Arg Trp Pro Thr Ser Gly 35 40 45	35
	GCG CGG TCG CGG GTC GCC GCC GTA GCG AAG ATA CAA ATG GCG CGA	40

Ala Arg Ser Arg Gln Val Ala Ala Val Ala Lys Ile Gln Met Ala Arg

CCA TTC ACG GAT CCC AAG TAT GAG TTC CTC AAC GAC TTC GTG TAC AAG Pro Phe Thr Asp Pro Lys Tyr Glu Phe Leu Asn Asp Phe Val Tyr Lys 70 75 80

F: 0 A

TC he	GGC	GTC Val	GCC Ala 85	GAT Asp	CTG (Leu)	CTA (Leu)	CCG ! Pro :	Phe 90	GGG (GCT A	AAC Asn	CAA Gln	Ser 95	eiH	Gl	n	003	
CC hr	GGC Gly	ACC Thr 100	GAT Asp	ATG Met	TAT Tyr	Thr	CGC Arg	TAC Tyr	AGT . Ser	ACA Thr	CTA Leu	TTT Phe 110	GAG Glu	GLY	G1 G1	ι G	551	
SAT Asp	GTA Val 115	CCC Pro	TTT Phe	GTG Val	CGC Arg	GCG Ala 120	GCT Ala	GGT Gly	GAC Asp	Gln	CGC Arg 125	GTC Val	GTT Val	GAC Asp	TC Se	c	599	
TCG Ser 130	ACG Thr	AAC Asn	TGG Trp	ACG Thr	GCA Ala 135	GGC	TTT Phe	GGC Gly	TAD	GCT Ala. 140	TCT Ser	GGC Gly	GAG Glu	ACT Thr	G1 V2 14		647	
CTC Leu	CCG Pro	ACG Thr	CTC Leu	CAG Gln 150	GTT Val	GTG Val	CTT Leu	CAA Gln	GAA Glu 155	GAG Glu	GGG Gly	AAC	TGC Cys	ACG Thr 160		en LC	695	
TGC Cys	TAA neA	AAT Asn	ATG Met 165	TGC Cys	CCG Pro	TAA neA	GAA Glu	GTG Val 170	GAT Asp	GGT Gly	gac Asp	GAA Glu	TCC Ser 175	ACA	A A	CG hr	743	
TGG Trp	CTG	GGG Gly 180	v Val	TTT Phe	GCG Ala	CCG Pro	AAC Asn 185	ATC Ile	ACC	GCG Ala	CGA	Leu 190	Man	GCT Ala	G A	CT la	791	
GCG Ala	CCG Pro	Sex	GCC Ala	AAC Asn	CTC Leu	TCA Ser 200	Asp	AGC Ser	GAC Asp	GCG	CTC Lev 205	Thi	CTC	ATO Mei	G G	at Sp	839	1
ATO Met	: Cys	CCC Pro	TTC Phe	GAC Asp	ACT Thr 215	Leu	AGC Ser	TCC	: GGG	AAC Asn 220	ALE	AGC A Se	CCC Pro	TT(-	GT Ys 125	887	•
GA0 Asp	CTA Lev	A TT	r ACC	GCG Ala 230	Glu	GAG Glu	TAT Tyr	GTC Val	TCG Sex 235	: Tyr	GA(TA L Ty:	TAC	TA' Ty 24		AC Asp	935	3
CTO	C GAG	C AA	G TAC S Ty: 24:	C TAT	GEC	ACG Thr	GLy	250	o GT?	OAA E IEA Y	GC:	T CT a Le	C GG: u Gl; 25	Y	T (TC /al	983	3
CA(G GGG	C GT y Va 26	1 G1	A TAC y Ty:	GTC Val	C AAT L Ast	GAG Glu 265	Lei	G CTT	r GCI	A CG A Ar	C TT g Le 27	U 111	c GG r Gl	y (CAA Gln	103:	1
GC	C GT a Va 27	l Ar	A GA	C GA(p Gl	F ACC	G CAC	Thi	AA As:	C CGG	C AC	G CT r Le 28	U AS	c AG p Se	C GA	C Sp	CCT Pro	107	9
GC Al 29	a Th	A TT r Ph	c cc	G CT	G AA(u As) 29	n Ar	r ACC g Th	TT r Ph	C TA e Ty	C GC r Al 30	a As	C TI	C TC	G CI		GAT Asp 305	112	7
ממ	ר זר	C AT	G GT	G CC	o Il	C TT	r GCO e Al	G GC a Al	G CT a Le 31	n GT	G CI	C Ti	C AA	111 Tr.	CC la 20	ACC Thr	117	!5

GCC Ala	CTC Leu	GAC Asp	CCG Pro 325	Leu	AAG Lys	CCC	GAC Asp	GAG Glu 330	AAC Asn	AGG Arg	TTG Leu	TGG Trp	GTG Val 335	GAC Asp	TCT Ser	122:
AAG Lys	CTG Leu	GTA Val 340	CCG Pro	TTC Phe	TCT Ser	GGA Gly	CAT His 345	ATG Met	ACG Thr	ĠTC Val	GAG Glu	AAG Lys 350	CTG Leu	GCA Ala	TGT Cys	1271
TCT Ser	GGG Gly 355	AAG Lys	GAG Glu	GCG Ala	GTC Val	AGG Arg 360	GTG Val	CTC Leu	GTG Val	AAC Asn	GAC Asp 365	GCG Ala	GTG Val	CAG Gln	CCG Pro	1319
CTG Leu 370	GAG Glu	TTC Phe	TGC Cys	GGA Gly	GGT Gly 375	GTT Val	GAT Asp	GLY	GTG Val	TGC Cys 380	GAG Glu	CTT Leu	TCG Ser	GCT Ala	TTC Phe 385	1367
GTA Val	GAG Glu	AGC Ser	Gin	ACG Thr 390	TAT Tyr	GCG Ala	CGG Arg	GAG Glu	TAA Asn 395	GGG Gly	CAA Gln	GGC Gly	GAC Asp	TTC Phe 400	GCC Ala	1415
AAG Lys	TGC Cya	G17 GGC	TTT Phe 405	GTT Val	CCG Pro	TCG Ser	GAA Glu	TAGO	GGGA	GA C	CGTC	TATG	C TA	CACA	GTAA	1469
TTGT	GTAC	TC T	ATAG	CACT	G TA	GCTG	TACT	TAC	aagt	CGT	AGGG	TACG	AT C	GTAC	TTACG	1529
CTCG	TTTA	TT G	ATCC	TTCC	т тт	AAAA	AAAA	ААА	AAAA	AAA .	AAAA	AAAA	AA A	TTCC	TGCGG	1589
CCGC		•											•			1593

	ccg.																						6
C	149	329	ac	32	200	, ==	cet	:50	555	csc	550	:550	355		-55	559	, , ,	gee	چوو	754	:45	tga	12
5	gagg	333	às	6 5	550	:=9	550	:52:	tga	tça	cgg	TAC	gaz	LEG	:52	acg	152	CZC	499	CC	get	329	18
c	355	355	gt	tg	cgt	:20	taa	te	EEE	CĒE	200	9	7001	عت	TE 2	cat	CE	700	tot	ot	a to	ĒŒ	24
E	tgg:	300	Č.	-	ai:	-		: Ear	ort:a	att	ato	É	ćŧ,	CCC		ctt			- , -			-6-	30
_	- 	300	33	3 7 ~+	3 P 1				,				~~										36
3	tte		-3	3-				, -7		ב- כ			,		177.		.23	-59	905	154	-99		
	322		94	45	GAC	39	a 5 4	lag	gaa	444	959	gaa	Laca	1663	LAC	acg	350	300	ctg	132	-23	=3=	421
3	ccg	1 90	τg	ga.	550	:tt	gac	:50	623	tet	320	222	cat	:221	CE	262	יבבי	cag	CAC	cc	776	EEA	48
7	LAC	ttg	at	25	a 5:	gt	ttc	55	320	caç	409	gti	tg	gct	gt	gt:	:==:	t ac	CZZ	tc	LCC	326	54
t	g G F (yct	AC.	tz	CEA	にてこ	att	50	995	tgt	tga	tge	age	cgr	gt	100	22	442	tgo	:cg	-55	zat	60
	LCC																						66
	SEE																						72
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1	AGATTCAACGACGGAGGAATCGCAACCCTAATTGTCGGTATCATGGTGAC M V T	50 3
Sl	TCTGACTTTCCTGCTTTCGGCCGCGTATCTGCTTTCTGGGtgagtgggctt L T F L L S A A Y L L S G	
101	ggatctattgctcggatagggctgtggtgctgattctgaaacggagTAGA R	150
151	GTGTCTGCGGCACCTAGTTCTGCTGGCTCCAAGTCCTGCGATACGGTAGA V S A A P S S A G S K S C D T V D	200
201	CCTCGGGTACCAGTGCTCCCCTGCGACTTCTCATCTATGGGGCCAGTACT L G Y Q C S P A T S H L W G Q Y S	
251	CGCCATTCTTTCGCTCGAGGACGAGCTGTCCGTGTCGAGTAAGCTTCCC F F F S L E D E L S V S S K L F	300 67
301	AAGGATTGCCGGATCACCTTGGTACAGGTGCTATCGCGCCATGGAGCGCG K D C R I T L V Q V L S R K G A R	350 84
351	GTACCCAACCAGCTCCAAGAGCAAAAAGTATAAGAAGCTTGTGACGGCGA Y P T S S K S K K Y K K L V T A I	
401	TCCAGGCCAATGCCACCGACTTCAAGGGCAAGTTTGCCTTTTTGAAGACG Q A N A T D F K G K F A F L K T +	450 11
451	TACAACTATACTCTGGGTGCGGATGACCTCACTCCCTTTGGGGAGCAGCA Y N Y T L G A D D L T P F G E Q Q	
501	GCTGGTGAACTCGGGCATCAAGTTCTACCAGAGGTACAAGGCTCTGGCGC L V N S G I K F Y Q R Y K A L A R	
SSL	GCAGTGTGGCGGTTATTCGCGCCTCAGGCTCGGACCGGGTTATTGCT S V V P F I P 2 S G S D R V I A	

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601	TCGGGAGAGAAGTTCATCGAGGGGTTCCAGCAGGGGAAGCTGGCTG	650
		930
	S G E K F I E G F Q Q A K L A ·D 9	194
6=1	TGGCGCGACGAACCGCGCCGCTCCGGCGATTAGTGTGATTATTCCGGAGA	~~~
93T		
	GATNRAAPAISVIIPES	201
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701		
701	GCGAGACGTTCAACAATACGCTGGACCACGGTGTGTGCACGAAGTTTGAG	
	ETFNNTLDEGVCTKFE	217
	*	
751	GCGAGTCAGCTGGGAGATGAGGTTGCGGCCAATTTCACTGCGCTCTTTGC	0.00
	ASQLGDEVAANFTALFA	234
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801	ACCCGACATCCGAGCTCGCGCGAGAAGCATCTTCCTGGCGTGACGCTGA	950
991	ACCOMPATICATION TO TO CONTRACT TO THE TOTAL TO THE TOTAL TO THE	830
	PDIRARAEKHLPGVTLT	251
851	CAGACGACGACGTTGTCAGTCTAATGGACATGTGTTCGTTTGA <u>TACGGTA</u>	800
971	CHARLEST OF CHARLEST OF THE TOTAL OF THE CONTROL OF	300
	DEDVVSLMDMCSFDTV	257
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901	GCGCGCACCAGCGAAGTCAGCTGTCACCGTTCTGTCAACTCTTCAC	950
202	DEGEGERACE ACCEPTAGE TO LACEUTE TO TEACTET TO ACC	330
	ARTSDASQLSPFCQLFT	284
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951	TCACAATGAGTGGAAGAAGTACAACTACCTTCAGTCCTTGGGCAAGTACT	1000
	TERCHAPTED THE THE TRACE TO THE TOTAL CONTROL OF THE TOTAL CONTROL OT THE TOTAL CONTROL OF TH	1000
	H N E W K K Y N Y L Q S L G K Y Y	301
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1001	ACGGCTACGGCGCAGCCAACCCTCTGGGACCGGCTCAGGGGATAGGGTTC	1050
7007		
	GYGAGNPLGPAQGIGF	317
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1061	ACCAACGAGCTGATTGCCCGGTTGACTCGTTCGCCAGTGCAGGACCACAC T N E L I A R L T R S P V Q D H T	1100
TOST	ACCACCAGE FOAT FOCECOGE FOACECOE FOCACIACIACIA	TIGG
	TNELIARLTRSPVODET	334
		11=0
TIGI	CAGCACTAACTCGACTCTAGTCTCCAACCCGGCCACCTTCCCGTTGAACG	
	STNSTLVSNPATFPLNA	351
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	+	
1121	CTACCATGTACGTCGACTTTTCACACGACAACAGCATGGTTTCCATCTTC	1200
	TMYVDFSHDNSMVSIF	
		23/
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1201	TTTGCATTGGGCCTGTACAACGGCACTGAACCCTTGTCCCGGACCTCGGT	1250
1201	TTTGCATTGGGCCTGTACAACGGCACTGAACCCTTGTCCCGGACCTCGGT F A L G L Y N G T E P L S R T S V	1250

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1251	GGAAAGCGCCAAGGAATTGGATGGGTATTCTGCATCCTGGGTGGTGCCTT	1300
	ESAKELDGYSASWVVPF	
	•	
1301	TCGGCGCGCGAGCCTACTTCGAGACGATGCAATGCAAGTCGGAAAAGGAG	1750
7707		
	GARAYFETMQCKSEKE	41/
1721	CCTCTEGTTCGCGCTTTGATTAATGACCGGGTTGTGCCCACTGCATGGCTG	
	PLVRALINDRVVPLKGC	4.34
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1401	CGATGTGGACAAGCTGGGGCGATGCAAGCTGAATGACTTTGTCAAGGGAT	
	DVDKLGRCKLNDFVKGL	451
1451	TGAGTTGGGCCAGATCTGGGGGAGAGTGCTTTAGTTGAGAT	
	SWARSGGNWGECFS	46
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1501	GTCATTGTTATGCTATACTCCAATAGACCGTTGCTTAGCCATTCACTŢCA	1550
	•	
1551	CTTTGCTCGAACCGCCTGCCG	1573

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2	cerreus 9A-1	_				50
	cettens cps	MGVETVIS.	VallfrsTSG iatlfgsTSG	Talegre N	hencesyn-c	YOCEPELSHK
	niger var. awamori		lYLLagVTSG			
	niger T213					
	niger NRRL3135		lyLLagVTSG			
	fumigacus 13073		LYLLsgVTSG			
	fumigacus 32722		AYLLsgVSAA			
	fumigacus 58128		AYLLsgVSAA			
	fumigatus 26906		AYLLSGVSAA			
	fumigatus 32239		AYLLsgVSAA			
	nidulans		mYLLsgVAGA			
	chermophilus	MASSEVALSL	yYLLsrVSAQ	ArvvQ	NKSCNTADEG	IQCE PRVSRV
	chermophila	MATERIAL SA	GLValyVS=N	PRV	SUSCULATION SIG	FOC-TRIBE
•••		MAGSTATASE			arscorbord	EQCG LAISHE
	Consensus	MCTT _VTTST	GYLLVSAG	22VCN	SHECOTYPES	AUCESEIGHI
	Conphys		IATLEGSTAG			
	oonput 6	PRIVITA ATTES.	TALLE GOLAG	IALGENGN	3436014000	Idestaroim
		51				100
Α.	terreus 9A-1		QCESPFPLOV	PEDCETTEVO	VT.ARHGARSP	
	cerreus cbs		QDESPEPLDV			
A.	niger var. awamori					
Α.	niger T213	MCUAPTEE	ANESVISPOV	PAGCAVIEND	AT 25 ACT BAD	TESKEKKYSA
	niger NRRL3135	MCOATBLLCI	ANESVISPEV	PAGCRVIERO	VI SAHGARYD	TDSKcKkYSA
\overline{A} .	fumigatus 13073	MCOVEREESI	EDELSVSSKL	PKDCRITTIVO	VI SARCARYS	TSSKSKKYKK
A.	fumigacus 32722		EDEISVSSKL			
A.	fumigatus 58128		EDELSVSSKL			
Α.	fumigatus 26906		EDELSVSSKL			
	fumigatus 32239	WGOYSPEESI.	EDELSVSSDL	PKDCRVTEVO	VI.SRHGARYP	TASKSKKYKK
	nidulans	WCOYSPYEST	EQESAISeDV	PHECEVTEVO	VT.SRHGARYD	TESKSKAYSG
	thermophilus	WCOYSPEES	ADQSEISPDV	PONCKITEVO	LLSRHGARYP	TSSKEELYSO
M.	thermophila	WGOYSPYESV	pSELDaSI	PODCEVTEAO	VLSRHGARAP	TLKRaaSYvD
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	Consensus	WGOYSPYFSL	EDESAISPOV	PDDCRVTEVO	VLSRHGARYP	TSSK-KAYSA
	Conphys	WGOYSPYESL	EDESAISPDV	PDDCRVTEVO	VLSRHGARYP	TSSKSKAYSA
		•				
		101				150
	terreus 9A-1	tIAAIQKSAT	afpGKYAFLQ	SYNYSLOSEE	LTPFGrNQLr	DlGaQFYeRY
	terreus cbs	tIAAIQKNAT	alpGKYAFLK	SYNYSMGSEN	LTPFGFNQLq	DLGaQFYRRY
A.	niger var. awamori	LIEEICONVT	LFDGKYAFLK	TYNYSLGADD	LTPFGEQELV	NSGIKEYQRY
A.	niger T213	LIEEICONVT	t FDGKYAFLK	TYNYSLGADD	LTPFGEQELV	NSGIKEYQRY
<u>A.</u>	niger NRRL3135	LIEEICONAT	t FDGKYAFLK	TYNYSLGADD	LTPFGEQELV	NSGIKEYQRY
	fumigatus 13073	LVTAIOANAT	dfKGKFAFLK	TYNYTLGADD	LTPFGEQQLV	NSGIKEYORY
	fumigatus 32722	LVTAIOANAT	dekckeaflk	TYNYTLGADD	LTPFGEQQLV	NSGIKEYQRY
A.	fumigacus 58128	LVTAIOaNAT	dfkgkfaflk	TYNYTLGADD	LTPFGEQQLV	NSGIKEYQRY
A.	fumigacus 26906	LVTAIOANAT	dekgkeaflk	TYNYTLGADD	LTAFGEOOLV	NSGIKEYQRY
<u>.A</u> .	fumiçacus 32239	LVTAIOKNAT	eFKGKFAFLE	TYNYTLGADD	LTPEGEQOMV	NSGIKFYOKY
	nidulens -	LIFATOKNAT	SEWGQYAFLE	SYNYTIGADO	LTIFGENOMV	DSGaKFYRRY
	Chermophilus	LISTICKTAT	aYKGYYAFLK	DYEYGLGAND	LTPEGENOMI	QLGIKEYNHY
	chermophila	LIDEISHGAT	sygPgYEFLR	TYDYTLGADE	LTREGCOOMV	NSGIKEYRRY
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	Consensus	LIZATORNAT	-FKGKYAFLK	TYNYTLGADO	LTPFGENOMY	NSGIKFYRRY
	Conphys	LIEATOWNE	AFKGKYAFIK	TYNYTLGADD	LTPEGENCMV	NSGIKEYRRY

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Α.	tarreus 9A-1		VRATDASRVh	FSAEKEVEGE	GTARGODENA	
	terreus cbs		VRAADSSRVh			
	niger var. awamori					
	niger T213		IRSSGSSRVI			
Α.		TOT TOWELDS	IRSSGSSRVI	ASCENTIFGE	CSTKT. LOP-A	ded0225rkrd
	fumigatus 13073	FORTWILLE	IRASGSDRVI	ACCENETECE	Oct XT ADRCA	TNEADBATE
	fumigacus 32722	ペイドサイフへんろこ	LRASGSURVI	ASGENITEGE	O-MAC ADECA	TNOARCALS
	fumigatus 58128	KALAKSVVEE	IRASGSDRVI	ASGERILEGE	CCVUTADECY	TWOARTALS
		KALAKSVV95	IRASGSDRVI	ADGENITEGE	O-FICE FORCE	TNOARTALS
	fumigatus 26906	RALARSVVDE	IRASGSDRVI	ASGERILLGE	QGANLADEGA	.INCAACAIS
	fumigacus 32239	KALAGSVVPF	IRSSGSDRVI	ASGEK: LEGE	QQANVADEGA	TURAAPVIS
	nidulans	KNILARKNIPE	IRASGSDRVV	ASAEKFINGF	REAQLADEGS	gQATPVVn
	thermophilus	KSLARNavef	VRCSGSDRVI	ASGILFIEGE	QSAKVIDPhS	dKHDAPPTIn
M.	Chermophila	RALARKSIPF	VRTAGGDRVV	hsaenftqgf	RSALLADRGS	EAKLITEACU
	_					
	Consensus	KALARKIVPE	IRASGSDRVI	ASAEKFIEGE	QSAKLADPGS	-PHQASPVI-
	Conphys	KALARKIVPF	IRASGSDRVI	ASAEKFIEGF	QSAKLADPGS	QPHQASPVID
		•				250
		201	_			250
	cerreus 9A-1	Valfegsayn	NTLEHSLCTA	FESSEVG	DDAVANETAV	FAPALAQREE
	terreus cbs	VVIPEGTAYN	NTLEHSICTA	FEASEVG	DAAaDNETAV	FAPATAKRLE
Α.	niger var. awamori	VVISZASS _S N	NTLDPGTCTV	FEDSELA	DTVEANETAT	FAPSTRORLE
	niger T213	VVISEASSEN	NTLDPGTCTV	FEDSELA	DIVEANETAT	FAPSIRQRLE
	niger NRRL3135	VVISEASSEN	NTLDPGTCTV	FEDSELA	DIVEAMETAT	FVPSIRQRLE
	fumigatus 13073	VIIPESETEN	NTLDHGVCTk	FEASQLG	DEVAANFTAL	FAPDIRARAE
	fumigatus 32722	VIIPESETFN	NTLDHGVCTk	FEASQLG	DEVAANFTAL	FAPDIRARAE
	fumigatus 58128	VIIPESETEN	NTLDHGVCTK	FEASQLG	DEVAANFTAL	FAPDIRARAE
	fumigatus 26906	VIIPESETFN	NTLDHGVCTk	FEASQLG	DEVAANFTAL	FAPDIRARAK
	fumigatus 32239	VIIPESETYN	NTLDHSVCTN	FEASELG	DEVEANETAL	FAPAIRARIE
Α.	nidulans	VITERIDGEN	NTLOHSTOVS	FEN. DELA	DELEANFTAI	MGPPIRKRLE
	thermophilus	VITAEGPSYN	NTLDEGSCPV	FEDSSaG	HDAQEKFAKq	FAPALLEKIK
M.	thermophila	VVIPETAGAN	NTLHNDLCTA	FEEgpyStIG	DDAQDTYLST	FAGPILARVN
	Consensus	VIIPEGSGYN	NTLOHGTCTA	FEDSELG	DOAEANFTAT	FAPAIRARLE
	Conphys	VIIPEGSGYN	NTLDHGTCTA	FEDSELG	DOVEANETAL	FAPAIRARLE
						222
		251				300
A.	terreus 9A-1	ADLPGVqLST	DDVVnLMAMC	PFETVSLTD.	• • • • • • • • •	DANTLSPECD
<u>A.</u>	terreus cbs	ADLPGVGLSA	DOVVILMANC	PFETVSLTD.		DANTESPECE
A.	niger var. awamori	NDI.SCVTI.TD	TEVTYLMDMC	SEDTISEST.		ADLKT25FCD
A.	niger T213	NOT.SGVTT.TD	TEVTVIMONC	SEDTISEST.		ADIKT2 SECD
A.	niger NRRL3135	NDLSGVTLTD	TEVTYLMDMC	SEDTISTST.		ADLKT255CD
Ā.	fumigatus 13073	KHT.PGVTT.TD	EDVV91.MDMC	SEDTVARTS.		DASQLSPECQ
A.	fumigatus 32722	WHT.PGVTT.TO	FOVUSLADAC	SEDTVARTS.		DASQLSPECQ
	fumigatus 58128	WHI.PGVTT.TD	FDVVgLMDMC	SEDTVARTS.		DWZGTZSECG
	fumigatus 26906	WHI.PGVTI.TO	FOUVELMONC	SEDTVARTS.		DYZÖTZ SECÖ
	fumigatus 32239	kHLPGVal.TD	DDVVsLMDMC	SEDTVARTA.		DWZFTZEECV
	nidulans	MOLDCIKE TN	FNVTVLMDMC	SEDTMARTA.		MGIFFRECY
	thermophilus	DHT.PGVDT.Av	SDVDVLMDLC	PFETLARNh.		TUT. LSPECA
	thermophila	ANT PCANT TO	ADTVaLMDLC	PEETVASSSS	dpatadaggg	NGrpLSPECr
•••						
	Consensus	ADT. PGVTT. TO	. EUAA-I WUWC	PEETVARTS-		DATELSPECA
	Conphys	Ant.pcvmr.mn	EDUCYTMOMC	PEETVARTS		DATELSPECA
	hrila	THE STATE	THE THE STATE OF T			

		•••				•
A		30:				350
	cerreus 9A-l cerreus cos	LiTacEWtQY	NATTZTOKAA	Gygggnplgp	VQGVGWaNEL	MARLTRAPVH
		LETAAEWEQY	NYLISLOKYY	GYGGGNPLG?	VQGVGWaNEL	IARLTRSPVH
Α.	niger var. awamori niger T213					
	niger 1213 niger NRRL3135					IARLTHSPVH
					TQGVGYANEL	
	fumigacus 13073					IARLTRSPVQ
	fumigatus 32722					IARLTRSPVQ
	fumigatus 58128					IARLTRSPVQ
	fumigacus 26906					IARLTRSPVQ
	fumigacus 32239					IARLThSPVQ
	nidulans					IARLTQSPVQ
	chermophilus					IARMTHSPVQ
М.	Chermophila	LFSEsEWraY	DYLQSVGXWY	GYGPGNPLGP	TQGVGFVNEL	LARLAGVPVR
	Consensus					IARLTRSPVQ
	Conphys	LFTHDEWRQY	DYLQSLGKYY	GYGAGNPLGP	AQGYGFANEL	LARLTRSPVQ
		351				400
	terreus 9A-1				NLVSIFWALG	
	terreus cbs	DHTCVNNTLD	ANPATEPLNA	TLYADESHOS	NLVSIFWALG	LYNGTkPLSq
A.	niger var. awamori	DDTSSNHTLD	SNPATEPLNS	TLYADESHON	GIISILFALG	LYNGTkPLST
	niger T213				GIISILFALG	
	niger NRRL3135				GIISILFALG	
	fumigatus 13073	DHTSTNsTLv	SWPATERLNA	TMYVDESHON	SMVSIFFALG	LYNGTEPLSE
	fumigatus 32722	DHTSTNsTLv	SNPATFPLNA	TMYVDESHDN	SMVSIFFALG	LYNGTGPLST
	fumigatus 58128	DHTSTNsTLv	SNPATFPLNA	TMYVDFSHDN	SMVSIFFALG	LYNGTEPLS:
А.	fumigatus 26906				SMVSIFFALG	
	fumigatus 32239	DHTSTNsTLD	SDPATFPLNA	TIYVDESHDN	GMIPIFFAMG	LYNGTEPLSq
	nidulans	ONTSTNHTLD	SNPATEPLOR	KLYADESHON	SMISIFFAMG	LYNGTQPLSm
T.	thermophilus	DYTTVNHTLD	SNPATEPLNA	TLYADESHON	TMTSIFaALG	Lyngtaklst
M.	thermophila	DGTSTNRTLD	GDPTTFPLGr	PLYADESHON	DMMGVLgALG	aydgyppldk
	_		•			
	Consensus	DHTSTNHTLD	SNPATFPLNA	TLYADFSHDN	SMISIFFALG	LYNGTAPLST
	Conphys	DHTSTNHTLD	SNPATFPLNA	TLYADESHDN	SMISIFFALG	LYNGTAPLST
		*				
				•		
		401				. 450
	terreus 9A-1	TSVESVSQTD	GYAAAWTVPF	AARAYVEMMQ	<u>c</u>	RAEKEP
	terreus cbs	TTVEDITETD	GYAAAWTVPF	AARAYIEMMQ	c	RAERQE
Α.	niger var. awamori	TTVENITQTD	GESSAWTVPE	ASRLYVEMMQ	C	QAEQEP
	niger T213	TTVENITQTD	GESSAWTVPF	ASRLYVEMMQ	C	QAEQEP
	niger NRRL3135	TTVENITQTD	GESSAWTVPE	ASR1YVEMMQ	c	QAEQEP
A.	fumigacus 13073	TSVESaKELD	GYSASWVVPF	GARAYFELMQ	C	KSEKEP
Α.	fumigatus 32722	TSVESaKELD	GYSASWVVPF	GARAYFELMQ	C	KSEKEP
Α.	fumigatus 58128	TSVESaKELD	GYSASWVVPF	GARAYFELMQ	C	KSEKES
Α.	fumigacus 26906	TSVESaKELO	GYSASWVVPF	GARAYFELMQ	C	KSEKEP
	fumigatus 32239	TSeESTKESN	GYSASWAVPE	GARAYFELMQ	C	KSEKEP
	nidulans	DSVESIQEMD	GYAASWTVPF	GARAYFELMQ	C	E.KKEP
T.	thermophilus	TEIKSIEETO	GYSAAWTVPF	GGRAYIEMMQ	C	DOSDEP
M.	thermophila	TArrOpEELG	GYAASWAVPE	AARLYVEKMR	Csgggggggg	geg≍QEKDEa
	Consensus	TSVESIEETO	GYSASWTVPF	GARAYVEMMQ	C	Qaekep
	Conphys	TSVESIEETD	GYSASWTVPF	GARAYVEMMQ	c	QAEKEP

		451				500
Α.	cerreus 9A-l	LVRVLVNDRV	MPLHGCPTOK	LGRCK:DAFV	AGLS FAQAGG	NWADCE
<u>A.</u>	terreus cos	LVRVLVNDRV	MPLHGCAVDN	LGRCK-DDFV	EGLSTARAGG	NWAECE
A.	niger var. awamori	LVRVLVNDRV	VPLHGCPIDa	LGRCT=DSEV	rGLSFARSGG	DWAECSA
Α.	niger T213	LVRVLVNDRV	VPLHGCPIDa	LGRCTEDSEV	rglsfarsgg	DWAECFA
Α.	niger NRRL3135	LVRVLVNDRV	.VPLHGCPVDa	LGRCTTDSTV	rGLSFARSGG	DWAECEA
A.	fumigacus 13073	LVRALINDRY	VPLHGCDVDK	LGRCKLNDEV	KGLSWARSGG	NWGECES-~~
Α.	fumigacus 32722	LVRALINDRV	VPLHGCDVDK	LGRCKLNDFV	KGLSWARSGG	NWGECES~~~
A.	fumigatus 58128	LVRALINDRV	VPLHGCDVDK	LGRCKLNDEV	KGLSWARSGG	NWGECES
A.	fumigatus 26906	LVRALINDRV	VPLHGCDVDK	LGRCKLNDFV	KGLSWARSGG	NWGECFS
<u>A.</u>	fumigatus 32239	'LVRALINDRV	VPLHGCAVDK	LGRCKLKDEV	KGLSWARSGG	NSEQSES
A.	nidulans	LVRVLVNDRV	VPLHGCAVDK	FGACTLDDWV	EGLNTARSGG	NW LTCFTL
T.	thermophilus	VVRVLVNDRV	VFLHGCEVDS	LGRCKEDDFV	rGLS₹AR⊂GG	NWEGCYAase
М.	thermophila	MYRYLYNDRY	MTLKGCGADE	rGMCTLErFI	ESMAFARGNG	KWD1CFA
	Consensus	LVRVLVNDRV				NWAECEA
	Corphys	LVRVLVNDRV	VPLHGCAVDK	LCRCKRODEV	EGLSFARSGG	NWAECEA

	C9-1	
	TATATGAATTCATGGGCGTGTTCGTCGTGCTACTGTCCATTGCCACCTTGTTCGGTTCCA	
	1	
	ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGT	
	CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTG	
6.	1	
	GTAGGCCATGGCGGAACCCAGGAGCACCATTAAGAGTGAGAACACTGTGACAACTGCCAC CP-2	:
	CP-3	
121	GTTACCAATGTTTCCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATACTTCTCTT	
	CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAGTTATGAGAGGTATGAAGAGAA	
101	TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCGTTC	
181		
	ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG	
	CP-4	
	CP-5	
	AAGTTTTGTCTAGACACGGTGCTAGATACCCAACTTCTTCTAAGTCTAAGGCTTACTCTG	
241		300
	TTCAAAACAGATCTGTGCCACGATCTATGGGTTGAAGAAGATTCAGATTCCGAATGAGAC	
301	CTTTGATTGAAGCTATTCAAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA	260
	GAAACTAACTTCGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTCATGCGAAAGAACT	360
	CP-6	
	CP-7	
	AGACTTACAACTACACTTTGGGTGCTGACGACTTGACTCCATTCGGTGAAAACCAAATGG	
361	·	420
	TCTGAATGTTGATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACC	
421	TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT	
421		480
	AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA	
	CP-8	
	CP-9	
481	TCATTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTT	
	AGTAATCTCGAAGACCAAGACTGTCTCA A TA A CCA A CA CCA CTTTTTCA A CTA A CTTTCCA A	-+ 5

541	TCCATC IGC TAAGITGSCTGACCCAGGITCTCAACCACACCAAGCTTCTCCAGTTATTG	500
	AGGTTAGACGATTCAACCGACTGGGTCGAAGAGTTGGTGGTTCGAAGAGGTCAATAAC	
	CP-11 ACGTTATTATTCCAGAAGGaTCcGGTTACAACAACACTTTGGACCACGGTACTTGTACTG	
601	TGCAATAATAAGGTCTTCCtAGgCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGAC	660
661	CTTTCGAAGACTCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTCGCTC	720
	GAAAGCTTCTGAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAG CP-12	
721	CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGAC	790
121	GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAAACTGACTG	700
781	TIGITTACTIGATGGACATGTGTCCATTCGAAACTGTTGCTAGAACTTCTGACGCTACTG	840
	AACAAATGAACTACCTGTACACAGGTAAGCTTTGACAACGATCTTGAAGACTGCGATGAC	
841	AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGACAATACGACTACTTGC	900
	TTAACAGAGGTAAGACAAGAACAAGTGAGTGCTGCTTACCTCTGTTATGCTGATGAACG CP-14 CP-15	
901	PATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG+ TTAGAAACCCATTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCAC	960
961	TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT	.1020
	AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA	
	CP-17	
021	CTACTAACCACACTTTGGACTCTAACCCAGCTACTTTCCCATTGAACGCTACTTTGTACG	1080
	GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGC	

1081	CTGACTTCTCACGACAACTCTATGATTTCTATTTTCTTCGCTTTGGGTTTGTACAACG										
	GACTGAAGAGAGTGCTGTTGAGATAAAGATAAAAGAAGCGAAACCCAAACATGTTGC CP-18										
	CP-19										
1141	GTACTGCTCCATTGTCTACTTCTGTTGAATCTATTGAAGAAACTGACGGTTACTCTG										
	CATGACGAGGTAACAGATGATGAAGACAACTTAGATAACTTCTTTGACTGCCAATGAGAC	1200									
1201	CTTCTTGGACTGTTCCATTCGGTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG										
	GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACAGTTCGAC	1200									
	CP-20										
	CP-21										
1261	AAAAGGAACCATTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG	1 2 2 2 2									
	TTTTCCTTGGTAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC	T350									
1321	CTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTA	1 300									
•	GACAACTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT	T290									
	CP-22										
	GATCTGGTGGTAACTGGGCTGAATGTTTCGCTTAAGAATTCATATA										
	CTAGACCACCATTGACCCGACTTACA & CCCA ATTCTTA & CTATAT										

F: 00

1	TCTGTAACCGATAGCGGACCGACTAGGCATCGTTGATCCACAATATCTCA
Sl	GACAATGCAACTCAGTCGAATATGAAGGGCTACAGCCAGC
101	GGCCGTCTAGGTCGGGCTCCGGGGATGAGGAGGAGCAGCTCGTGTTCAT
151	TTCGGTCATGGCTTTTTTCACGGTCGCTCTTTCGCTTTATTACTTGCTAT M A F F T V A L S L Y Y L L S
201	CGAGGTGAGATCCCTACAACACCTGTCCTGCTCTAGTTGAATTGGTACTTAT
251	CEGEACAGAGTCTCTGCTCAGGCCCCAGTGGTCCAGAATCATTCAT
301	TACGGCGGACGGTGGATATCAATGCTTCCCCAATGTCTCTCATGTTTGGG T A D G G Y Q C F P N V S H V W G +
351	GTCAGTACTCGCCGTACTTCTCCATCGAGCAGGAGTCAGCTATCTCTGAG Q Y S P Y F S I E Q E S A I S E
4 0 1	GACGIGCCTCATGGCTGAGGTTACCTTTGTGCAGGTGCTCTCGCGGCA
451	TGGGGCTAGGTATCCGACAGAGTCGAAGAGTAAGGCGTACTCGGGGTTGA
501	TTGAAGCAATCCAGAAGAATGCTACCTCTTTTTGGGGACAGTATGCTTTT SE A I Q K N A T S F W G Q Y A F S +
551	CTGGAGAGTTATAACTATACCCTCGGCGCGGGATGACTTGACTATCTTCGG L E S Y N Y T L G A D D L T I F G :
501	CGAGAACCAGATGGTTGATTCGGGTGCCAAGTTCTACCGACGGTATAAGA 6 E N Q M V D S G A K F Y R R Y K N
551	ATCTCGCCAGGAAAATACTCCTTTTATCCGTGCATCAGGGTCTGACCGT L A R K N T P F I R A S G S D R

701	GTCGTTGCGGAGAGTTCATTAATGGATTTCGCAAGGCTCAGCT V V A S A E K F I N G F R K A Q L	750 180
35.		
/51	CCACGACCATGGCTCCAAACGTGCTACGCCAGTTGTCAATGTGATTATCC	800
	H D H G S K R A T P V V N V I I P	197
801	CTGAAATCGATGGGTTTAACAACACCCTGGACCATAGCACGTGCGTATCT	050
	E I D G F N N T L D H S T C V S	850 213
	*	
851	TTTGAGAATGATGAGCGGGGGATGAAATTGAAGCCAATTTCACGCAAT	500
	FENDERADE I EANFTAI	230
	.	
901	TATGGGACCTCCGATCCGCAAACGTCTGGAAAATGACCTCCCTGGCATCA	950
	MGPPIRKRLENDLPGIK	247
951	AACTTACAAACGAGAATGTAATATATTTGATGGATATGTGCTCTTTCGAC	1000
	_	263
	•	
1001	ACCATGGCGCACCGCCCACGGAACCGAGCTGTCTCCATTTTGTGCCAT	1050
		280
	•	
1051	<u>CTTCACTGAAAAGGAGTGGCTGCAGTACGACTACCTTCAATCTCTATCAA</u>	1100
	FTEKEWLQYDYLQSLSK	
	·	
1101	AGTACTACGGCTACGGTGCCGGAAGCCCCCTTGGCCCAGCTCAGGGAATT	1150
	YYGYGAGSPLGPAQGI	
LLSL	GGCTTCACCAACGAGCTGATTGCCCGACTAACGCAATCGCCCGTCCAGGA	1200
	G F T N E L. I A R L T Q S P V Q D	330
	-	
1201	CAACACAAGCACCAACCACACTCTAGACTCGAACCCAGCCACATTTCCGC	1250
	N T S T N H T L D S N P A T F P L	
	+ +	
1361		
-431	TCGACAGGAAGCTCTACGCCGACTTCTCCCACGACAATAGCATGATATCG DRKLYADF.SHDNJSMLIS	
T301	ATATTCTTCGCCATGGGTCTGTACAACGGCACCCAGCCGCTGTCAATGGA	
	IFFAMGLYNGTQ9LSMD	3 9 (

1351	TTCCGTGGAGTCGATCCAGGAGATGGACGGTTACGCGGCGTCTTGGACTG	1400
	SVESIQEMDGYAASWTV	207
		331
1401	TTCCGTTTGGTGCGAGGGCTTACTTTGAGCTCATGCAGTGCGAGAAGAAG	
1401		
	PFGARAYFELMQCEKK	413
	·	
	• • • • •	
1451	GAGCCGCTTGTGCGGGTATTAGTGAATGATCGCGTTGTTCCTCTTCATGG	1500
	EPLVRVLVNDRVVPLHG	430
		430
	•	
1501		
	CTGCGCAGTTGACAAGTTTGGACGGTGCACTTTGGACGATTGGGTAGAGG	
	CAVDKFGRCTLDDWVEG	447
	•	
	• • • • • • • • • • • • • • • • • • • •	
1551	GCTTGAATTTTGCAAGGAGCGGCGGGGAACTGGAAGACTTGTTTTACCCTA	1600
	LNFARSGGNWKTCFTL	463
	·	
1601	TARAGGGCGTTTGCTCATTCATAAGTGTTGTGCAGGTATAGGAAGGTTAG	1650
TabT	GGRATTAGCTGTTTGGCTTTACTCTTATTAGACCAAGAATGATTTGTTTG	1700
1701	TTCTCAAGGCCTTCTAGCATATCGTCAAGTGGGATAAATCACCTATCCTC	1750
	• • • • • • • • • • • • • • • • • • • •	
1751	CRIGIGIAGGIGAACCCGCTCTTGCATCLACCTCTTGTGTTTCAGAGTAG	1800
TROI	TTTCACCAAACATATCCTCGTGTCCTCTCTCTGCTCTTCGGTCTCATAT	1850
1851	TACACTGTTCTCTATCTATATCGTCAACAAAACTACCACCCAAACACCAA	1900
-434		7340
	• •	
1901	ATGTCACACTTTCCAGCACGAAAFTTCTTCG 1911	
	· · · · · · · · · · · · · · · · · · ·	

- 1 ATGGGCGTCTCTGCTGTTCTACTTCCTTTTGTATCTCCTGTCTGGAGTCACCTCCGGACTG
 -23 M G V S A V L L P L Y L L S G V T S G L
- 61 GCAGTCCCCGCCTCGAGAAATCAATCCAGTTGCGATACGGTCGATCAGGGGTATCAATGC A V P A S R N Q S S C D T V D Q G Y Q C -1 +1
- 121 TTCTCCGAGACTTCGCATCTTTGGGGTCAATACGCACCGTTCTTCTCTCTGGCAAACGAA
 18 F S E T S H L W G Q Y A P F F S L A N E
- 181 TCGGTCATCTCCCCTGAGGTGCCCGCGGATGCAGGTCACTTTCGCTCAGGTCCTCTCC 38 S V I S P E V P A G C R V T F A Q V L S
- 241 CGTCATGGAGCGCGGTATCCGACCGACTCCAAGGGCAAGAAATACTCCGCTCTCATTGAG 58 R H G A R Y P T D S K G K K Y S A L I E
- 301 GAGATCCAGCAGAACGCGACCACCTTTGACGGAAAATATGCCTTCCTGAAGACATACAAC 78 E I Q Q N- A T T F D G K Y A F L K T Y H
- 361 TACAGCTTGGGTGCAGATGACCTGACTCCCTTCGGAGAACAGGAGCTAGTCAACTCCGGC 98 Y S L G A D D L T P F G E Q E L V N S G
- 421 ATCAAGITCTACCAGCGGTACGAATCGCTCACAAGGAACATCGTTCCATTCATCCGATCC
 118 I K F Y Q R Y E S L T R N I V P F I R S
- 481 TCTGGCTCCAGCCGCGTGATCGCCTCCGGCAAGAAATTCATCGAGGGCTTCCAGAGCACC 138 S G S S R V I A S G K K F I E G F Q S T
- 541 AAGCTGAAGGATCCTCGTGCCCAGCCCGGCCAATCGTCGCCCAAGATCGACGTGGTCATT 158 K L K D P R A Q P G Q S S P K I D V V I
- 601 TCCGAGGCCAGCTCATCCAACAACACTCTCGACCCAGGCACCTGCACTGTCTTCGAAGAC 178 S E A S S S N N T L D P G T C T V F E D
- 661 AGCGAATTGGCCGATACCGTCGAAGCCAATTTCACCGCCACGTTCGTCCCTCCATTCGT 198 S E L A D=T V E A N F T A T F V P S I R
- 781 ATGGACATGTGCTCCTTCGACACCATCTCCACCAGCACCGTCGACACCAAGCTGTCCCCC 238 M D M C S F D T I S T S T V D T K L S P

841	TTC	TGI	GAC	CIG	TIC	ACC	CAI	GAC	GAA	LTGC	ATC	:AAC	TĄC	GAC	TAC	CIC	CAC	TCC	TTG	IAA A
258	F	C	D	L	F	T	H	D	E	W	I	·N	Y	D	Y	L	Q	S	L	K
901	ÅAG	TAT	TAC	GGC	CAI	GGI	'GCA	GGI	'AAC	CCC	CTC	GGC	CCG	ACC	CAG	GGC	GTC	GGC	TAC	GCI
278	K	Y	Y	G	H	G	A	G	N	P	L	G	P	T	Q	G	V	G	Y	A
					•															
961	AAC	GAG	CTC	ATTC	GCC	ייי		ACC	CAC	rtee	CCT	GTC	CAC	GAT	'GAC	ACC	AGT	TCC	AAC	CAC
	N																			
-,2-		_		_			_	-	_	_		•		_	_	_	_	_	•-	
1021	A ~~T	-T-17-C	GAC	TOO	A CC	-	·	'A CC	-	~~~	بخليب	440	بلخاباء	ابن ∆د	ناملماء	TAC	ccc	CAC	لملمك	TCC
318	T	L	ממט	S	age S	- -	A	ACC T	F	P	L.	N	S	T	L	¥	A	D	F	S
J	_	_	_	_	_	~		-	-	-	_		_	_		_				
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	1 TTCCACGCTGAAGCCTGACTGCGATTTCCAAGCTGCATGCA	50
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	ANCTECCTECTTATCTTCATCAGACGCAGATACACAACCTGGTCTGTAGA	100
101	TGCACCCATGACGGACGAACGCACCGCTCTCTTGGCCTCCAGGGACCCGG	
		150
151	AGGTCGAGGGCGATGAGGTCGCGGCCTCGACGGCCTCCCAGTCCCTGTTG	700
201	•	200
- 201	CAGTTGAGATCTCGCTGCGAACGTCGACCGCAGATATGGTTGTCTTCGAC	250
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401	ATCCGCATGTTGATAGCCACTCTTGCAATACAGTGGAAGGAGGGTATCAG	450
	P H V D S H S C M T V E G G Y Q	36
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451·	TGTCGTCCAGAAATCTCCCACTCCTGGGGCCAGTATTCTCCATTCTTCTC	
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501	Company	
201	CCTGGCAGACCAGTCGGGAGATCTCGCCAGATGTCCCACAGAACTGCAAGA	550
	LADQSEISPDVPQNCKI	70
551	TTACGTTTGTCCAGCTGCTTCTCGTCACGGCGCTAGATACCCTACGTCT	
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601	TOCALCACTOR	-
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651	GACTGCGTACAAAGGCTACTATGCCTTCTTGAAAGACTACAGATACCAGC	700
	TAYKGYYAFLKDYRYQL	
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147	TGGGAGCGAACGACCTGACGCCCTTTGGGGAAAACCAGATGATCCAGTTG	750
	G A N D L T P F G E N Q M I Q L	136

751	GGCATCAAGTTTTATAACCATTACAAGAGTCTCGCCAGGAATGCCGTCCC	800
	G I K F Y N H Y K S L A R N A V P	153
801	ATTCGTTCGTTGCTCCGGCTCTGATCGGGTCATTGCCTCGGGGAGACTTT	850
	FVRCSGSDRVIASGRLF	
851	TCATCGAAGGTTTCCAGAGGCGCCAAAGTGCTGGATCCTCATTCAGACAAG	900 186
901	CATGACGCTCCTCCCACGATCAACGTGATCATCGAGGAGGGTCCGTCC	
	HDAPPTINVILEEGPSY	203
951	CAATAACACGCTCGACACCGGCAGCTGTCCAGTCTTTGAGGACAGCAGC	1000
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1001	GGGGACATGACGCACAGGAAAGTTCGCAAAGCAATTCGCACCAGCTATC	1050
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1051	CTGGAAAAGATCAAGGACCATCTTCCCGGCGTGGACCTGGCCGTGTCGGA	1100
	LEKIKDHLPGVDLAVSD	
1101	TGTACCGTACTTGATGGACTTGTGTCCGTTTGAGACCTTGGCTCGCAACC	1150
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1151	<u>ACACAGACACGCTGTCTCCGTTCTGCGCTCTTTCCACGCAAGAGGAGTGG</u>	1200
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1201	<u>CAAGCATATGACTACTACCAAAGTCTGGGGAAATAC</u> TATGGCAATGGCGG	1250
	Q A Y D Y Y Q S L G K Y Y G N G G	
1251	GGGTAACCCGTTGGGGCCAGCCCAAGGCGTGGGGTTTGTCAACGAGTTGA G N P L G P A Q G V G F V N E L I	1300
1301	TTGCTCGCATGACCCATAGCCCTGTCCAGGACTACACCACGGTCAACCAC	1350
	ARMTHSPVQDYTTVNH	336
1351	ACTOTTGACTOGAATOCGGCGACATTCCCTTTGAACGCGACGCTGTACGC	1400
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TAGI	AGATTTCAGCCACGACAACACAATGACGTCAATTTTCGCGGCCTTGGGCC	1450
	DFSHDNTMTSIFAALGL	T420
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1451	TGTACAACGGGACCGCGAAGCTGTCCACGACCGAGATCAAGTCCATTGAA	
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1501	GAGACGGACGGCTACTCGGCGGCGCGAGC	
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	ETDGYSAAWTVPFGGRA	403
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1551	CTATATCGAGATGATGCAGTGTGATGATTCGGATGAGCCAGTCGTTCGGG	1600
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1601	TGCTGGTCAACGACCGGGTGGTGCCACTGCATGGCTGCGAGGTGGACTCC	1650
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F037	CTGGGGCGATGCAAACGAGACGACTTTGTCAGGGGACTGAGTTTTGCGCG	1700
	LGRCKRDDFVRGLSFAR	457
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L701	ACAGGTGGGAACTGGGAGGGGTGTTACGCTGCTTCTGAGTAGGTTTATT	1750
	Q G G N W E G C Y A A S E -	466
		400
1751	CACCCACTTCCACCTTCACCTT	
	CAGCGAGTTTCGACCTTTCTATCCTTCAAACACTGCACAAAGACACACTG	1800
7201	CATGAAATGGTAACAGGCCTGGAGCGTTTTAGAAGGAAAAAAGTT	1845

06	GCCGATTTTGCCTTGTCCCAGACTATGCCTATCTCCCTGCCCATAATTTCACCCCCCTTTGCCCAGCAGCAGCATGAATCGCCCCCCCC
81(CGCCACGCACCTCGGTATCCTACTGCTCACAGGTGTACGCCGAGTTGCTTCAAAGGATCCAGGACGCGCCGACCGA
720	TACTCCCCCTTCTTCTCCCTGCCCGAGGTCTCTGAATCTCGCCTGCCGTGCCCAAGGGCTGTCGTGTCGAGGTTTGTGCAGGTGCTGTCC Y S P F F S L A E V S E I S P A V P K G C R V E F V Q V L S
630	OGTICGCCATTATTGACGCCCTCCCCCCCCATTCCTCCTTTCTGGAGGAGGAGCATCCCAACGTGGACATTGCCCGCCACTGGGGCCAG S A L L T A S P A I P P F W R K K H P N V D I A R H W G Q
540	TAGGTTTGGGGTCCTTCTGGTCCTGCTGCAATTgtacgcattcttctagaccctaattatagaggtctgttgctgatattctgact
450	CSCATCGTGCTGATATAAAAGACTGCCAAATGCCGAAGACGAAATGCAGCAACGTTCAGCCCGCAGAGTGATTGCCGTCATGGCGGGA M A G
360	CTGCAAGTACTATTGAATAGTGCTTCAATGCTACCATGATCGGACACCAACACTCATGGAAGCCCGCCC
270	ACCAGGGGATTGATTTTCAATGCGTTGTTGTTGTTCATCCGATTCATGAACAAGTGGACATTATTATTATGATTGCACGTGTCCTAAG
180	TTGCCACACCACCTGCATGAGAATCCATCGATCCATGATCGCTCAGGATGATCTGATCATCTCGCGTTGGAAGAGTCCACTTTATG
6	AATTACGGAGTAGTTGCCATTCGATGTTCATTGATCAACAGTCAACCCCAAGTTTCGTAGTATTTCCAAACTCCTCCACTGGCCGTGCG

Fig. 14A

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1890 AATGATCGGGTTGTGCCGCTGCATGGTTGTCGGTTGATCGGCGGGGTGTCGCGGGGGTGGGGTTAAGGGCACTCACGTTTGCT RWGRCRRDEWIKGLTFA C R < D NDRVVPLHG

1980 CGACAGGGTGGGCATTGGGATCGCTGTTTGATTAGATGCTCATAGACATAGCCCATGATTCCGAATTGATGTTTTTAGATACAATCA WDRCF 2070 CTGCGGAAAGGGAAATGATCCAAAAAGCGCCAGTCTAGTATAACTTTGCGAATCCGTTGACTTGTTCAGTCCTTGGTGTCGGTGTCGACCAACC AGCCTGCCACAGGTCCAATGTTCCCGCTCTACATGGAGTCCGTCGTCGCCGAGATCATCCACCCCAGCCCAGCGAGGAGCTGTTCCGTTG

AGGETATCTGCCGTGGTTGACCCCCGTGCTCACAGTCACA 2200

Fig. 14C

45/51

 $\underline{\textbf{AAGCTT}} \textbf{GGGCAAACTCATCATCATCATGATGATTCCACTGTTCAGCTACCTGGCTGCTTCTCT} \underline{\textbf{GT}} \textbf{GGGTTCATC}$ 80 HindIII MLILMIPLFSYLAAASL $\tt CTTTGCCCCTGTCTCGATGTTAAAATACTAAACATATTTCACC\underline{AG}{ACG}TGTACTCTCCCCTCAGCCAGTGTCCTGTGACA$ 160 RVLSPQPVSCD SPELGYQCDQQTTHTWGQYSPFFSVPS E I S P S V P D G C R L T F A Q V L S R H G A R F P T ${\tt CCCGGGTAAAGCCGCCGCCATCTCCGCTGTCCTCACCAAAATCAAAACCTCTGCCACCTGGTACGGTTCCGACTTTCAGT}$ PGKAAAISAVLTKIKTSATWYGSDFQ TCATCAAGAACTACGACTATGTACTTGGCGTAGACCACCTGACCGCGTTCGGCGAGCAAGAAATGGTCAACTCCGGCATC FIKNYDYVLGVDHLTAFGEQEMVNSGI ${\tt AAGTTCTACCAGCCTACTCCTCATCCAGACAGAAGACTCGGATACGCTCCCCTTCGTCCGCCCTCTGGCCAGGA}$ 560 K F Y Q R Y S S L I Q T E D SD F V R A S G Q E Т L P 640 R V I A S A E N F T T G F Y S A L S A D K N P S TACCAAGACCAGAAATGGTCATCATTTCTGAGGAGCCAACAGCCAACAACACCCATGCACCACGGCCTCTTCTTT 720 LPRPEMVIISEEPTANNTMHHGLCR E D S T T G D Q A Q A E F I A A T F P P I T A R L N A ${\tt CCAAGGTTTCAAAGGCGTCACCCTCTCCAACACCGACGTCCTATCACTAATGGACCTCTGCCCCTTTGACACCGTCGCCT}$ 880 Q G F K G V T L S N T D V L S L M D L C P F D T V 960 Y P L S S L T T T S S V S G G G K L S P F S L F T A ${\tt AGCGACTGGACAATCTACGATTACCTCCAGTCCCTAGGGGAAATACTACGGTTTCGGCCCCGGTAATTCCCTAGCTGCCAC}$ 1040 S D W T I Y D Y L Q S L G K Y Y G F G P G N S L A A ${\tt CCAGGGGGTAGGGTACGTCAACGAGCTTATCGCCCGCTTGATCCGTGCTCCGTCGTAGATCACACGACGACCAACTCTA}$ 1120 QGVGYVNELIARLIRAPVVDHTT

C: AFA

T L D G D E K T F P L N R T V Y A D F S H D N D M M N

 $\tt CTCTTGATGGCGACGAAAAAACGTTTCCGTTGAACAGAACGGTGTATGCGGATTTTTCCCATGATAATGATATGATGAAT$

ATCCTGACTGCTTTGCGGATATTCGAGCATATCAGTCCGATGGATAACACCACTATCCCGACCAACTATGGCCAGACAGG 1280

I L T A L R I F E H I S P M D N T T I P T N Y G Q T G

 ${\tt AGATGACGGGGTGAAGGAAAGGGATTTGTTCAAGGTTAGTTGGGCGGTGCCCTTTGCTGGGAGGGTGTACTTTGAGAAAA}\\ {\tt 1360}$

D D G V K E R D L F K V S W A V P F A G R V Y F E K

 ${\tt TGGTTTGTGATGGGGATGGGAAGATTGATAGTGATGAGGGCTCAGAAAGAGTTGGTGAGGATTTTGGTTAATGAT\\1400}$

M V C D A D G D G K I D S D E A Q K E L V R I L V N D

CGGGTGATGAGATTGAATGGGTGTGATGAACAGGGTAGGTGTGGATTGGAGAAGTTTGTGGAGAGTATGGAGTT 1520

RVMRLNGCDADEQGRCGLEKFVESMEF

ARRGGEWEERCFV XbaI

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	MLILMIPLFSYLAAASLRVLSPQPVSCDSPELGYQCDQQTTHTWGQYS	48
-	MTGLGVMVVMVGFLAIASLQSESRPCDTPDLGFQCGTAISHFWGQYS	47
49	PFFSVPSEISPSVPDGCRLTFAQVLSRHGARFPTPGKAAAISAVLTKIKT	98
48	: :.: : : :	97
99	SATWYGSDFQFIKNYDYVLGVDHLTAFGEQEMVNSGIKFYQRYSSLIQTE	148
98	: .::: ::. . . : : . . GAISYGPGYEFLRTYDYTLGADELTRTGQQQMVNSGIKFYRRYRAL	143
149	DSDTLPFVRASGOERVIASAENFTTGFYSALSADKNPPSSLPRP.EMVII	197
144	ARKSIPFVRTAGQDRVVHSAENFTQGFHSALLADRGSTVRPTLPYDMVVI	193
198	SEEPTANNTMHHGLCRSFEDSTTGDQAQAEFIAATFPPITARLNAQG	244
*	. .:. : :: . : . : : : . PETAGANNTLHNDLCTAFEEGPYSTIGDDAQDTYLSTFAGPITARVNA.N	
245		
243		
294	TASDWTIYDYLQSLGKYYGFGPGNSLAATQGVGYVNELIARLIRAPVVDH	343
	: : : : . :: : : . SESEWRAYDYLQSVGKWYGYGPGNPLGPTQGVGFVNELLARLAGVPVRDG	
344		393
343	. . :: :. : TSTNRTLDGDPRTFPLGRPLYADFSHDNDMMGVLGALGAYDGVPPLD	389
94		442
90	. : : : :. : : ::::::::::::	433
43	. EAQKELVRILVNDRVMRLNGCDADEQGRCGLEKFVESMEFARRGGEWE	490
34	:. : : : : : : : : : : : :	483
91	ERCFV 495	
84	L.CFA 487	

P_involtus_A1 P_involtus_A2 C_pubescens A_pediades P_lyoii A_fundigatus C_onsphyA A_midulans A_ficium_NRR13135 A_ferreus MAFSILASLI F_VENGASYGG A_midulans A_ficium_NRR13135 A_ferreus MAFSILASLI MAGFICIAPLIAL SGRYSAAPS A_MOSSCOTTV DLGYQCSPAT MASSILASTLYLLSGRYSAAPS SAGSKSCOTV DLGYQCSPAT MAGFICIAPLIAL SGRYSAAPS SAGSKSCOTV DLGYCCSPAT MAGFICIAPLIAL SGRYSAAPS SAGSKSCOTV DLGYQCSPAT MAGFICIAPLIAL SGRYSAAPS SAGSCOTV DLGYQCSPAT MAGFICIAPLIAL SGRYSAAPS SAGSKSCOTV DLGYQCSPAT MAGFICIAPLIAL SGRYSAAPS SAGSKSCOTV DLGYQCSPAT MAGFICIAPLIAL SGR	Peniophora numbers	1				37
P_involtus_A1 P_involtus_A2 ML GGFVALACLI SISEVLATSV PKNT APFFFIPESE T_pubescens A_pediades P_lycii A_fumigatus ConsphyA A_ficum_NRL3135 A_ficum_NRL3135 A_ferreus MMAFSTLASLL FVCYAYARAV PRAHIPLEDT SACLDVTRDV MSSAMPSFILL SURSALAST GFFFVAQUESCATA MMAFSTLASLL SGRVSAAPS SAGSKSCTV DLGYQCSPBL A_nidulans A_ficum_NRRL3135 A_ferreus MMAFF TALSLAYLL .SGRVSAAPS SAGSKSCTV DLGYQCSPBL A_nidulans A_ficum_NRRL3135 A_ferreus MMAFF TALSLYYLL .SGRVSAAPS SAGSKSCTV DLGYQCSPBL A_terreus MMAFF TALSLYYLL .SGRVSAAPS SAGSKSCTV DLGYQCSPBL A_terreus MGFL ATVISVALLE SGTVSGLAVVA SKNQSCDTV DLGYQCSPBL MAGICLGSFL VILLIGESELL TASPAIPPFW RKKHSUD I MAGICLGSFL VILLIGESELL TASPAIPPFW RKKHSUD I MAGICLGSFL VILLIGESELL TASPAIPPFW RKKHSUD I MAGICLGSFL VILLIGESELL TASPAIPPFW RVLSSCCTV DLGYQCSPBL MAGICLGSFL VILLIGESELL TASPAIPPFW RVLSSCCTV DGGYQCFBL MAGICLGSFL VILLIGESELL STRAIPPFW RVLSSCCTV DGGYQCFBL MAGICLGSFL VILLIGESELL TASPAIPPFW RVLSSCCTV DGGYCFBL MAGICLGSFL VILLIGESELL TASPAIPPFW	Alignment numbers	1		•		50
T_pubescens	P_involtus_A1	ML	FGFVALACLL	SLSEVLATSV	PKNT	
A_pediades	P_involtus_A2					
A_pediades P_lycii A_fumigatus A_fumigatus ConsphyA A_nidulans A_ficuum_NRRL3135 A_ferreus T_thermo T_lanuginosa M_thermophila C_foecundissimum A_pediades P_lycii A_nidulans A_ficuum_NRRL3135 A_terreus T_bermo T_lanuginosa M_thermophila C_foecundissimum A_ficund_Salus A_pediades A_pediades A_pediades A_pediades A_pediades A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_fumigatus A_fumigatus A_ficuum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_ficum_NRRL3135 A_ficum_NRRL3135 A_fumigatus A_ficum_NRRL3135 A_fumigatus A_fumigatus A_fumigatus A_fumigatus A_ficum_NRRL3135 A_ficum_NRRL3135 A_ficum_NRRL3135 A_ficum_NRRL3135 A_fumigatus A_fumi	T_pubescens					
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P_involtus_A1 P_involtus_A2 P_involtus_A2 T_pubescens A_pediades P_lycii A_fumigatus ConsphyA A_nidulans A_ficuum_MRRL3135 A_ficuum_MRRL3135 A_terreus T_lanuginosa M_thermophila P_involtus_A1 P_involtus_A2 T_pubescens A_pediades A_nidulans A_ficuum_MRRL3135 A_ficuum_MRRL3135 A_terreus T_lanuginosa A_terreus A_pediades A_nidulans A_ficuum_MRRL3135 A_terreus T_thermo T_lanuginosa A_nidulans A_ficuum_MRRL3135 A_ficuum_MRRL3135 A_terreus T_thermo T_lanuginosa A_terreus A_pediades A_nidulans A_ficuum_MRRL3135 A_ficuum_MRRL3135 A_terreus A_terreus A_nidulans A_ficuum_MRRL3135 A_terreus A_nidulans A_ficuum_MRRL3135 A_terreus A_nidulans A_terreus A_nidulans A_ficuum_MRRL3135 A_terreus A_nidulans A_ficuum_MRRL3135 A_terreus A_nidulans A_nidulans A_terreus A_nidulans A_terreus A_pediades A_nidulans A_terreus A_ter		-				
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P_involtus_A2 T_pubescens A_pediades A_pediades P_lycii A_fumigatus ConsphyA A_nidulans A_ficuum_NRRL3135 A_terreus T_thermo T_lanuginosa M_thermophila P_involtus A2 IKAGLSKLQS VQNFTDPKFD FIKSFTYDLG TSDLVPFGAA QSFDAGLEVF FUNDTYYSLG QDSLVELGAT QSSEAGQEAF FUNDTYYTLG HDDLVPFGAL QSSQAGEETF FUNDTYKFG VADLLPFGAN QSHQTGTDMY VALUET FLINDFVYKFG VADLLPFGAN QSHQTGTDMY A_fumigatus VKKLVTAIQA NATDFKGKFA FLKTYNYTLG ADDLTPFGEQ QLVNSGIKFY YSALIEAIQK NATSFWGQYA FLESYNYTLG ADDLTPFGEN QMVDSGAKFY A_terreus T_thermo T_lanuginosa M_thermophila VAELLQRIQD TATEFKGDFA FLRDYAYHLG ADDLTPFGEN QMIQLGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY		101				150
T_pubescens A_pediades A_pediades P_lycii QVAAVAKIQM ARPFTDPKYE FLNDFVYKFG VADLLPFGAN QSHQTGTDMY A_fumigatus ConsphyA A_nidulans A_ficuum_NRRL3135 A_terreus A_terreus T_thermo T_lanuginosa M_thermophila T_lanuginosa A_nidulans A_solieAlQK NATSFKGDFA T_solieAlQK NATSFKGDFA T		IKAGLTKLQG	VQNFTDAKFN	FIKSFKYDLG	NSDLVPFGAA	QSFDAGQEAF
T_pubescens A_pediades A_pediades P_lycii QVAAVAKIQM ARPFTDPKYE FLNDFVYKFG VADLLPFGAN QSHQTGTDMY A_fumigatus ConsphyA A_nidulans A_ficuum_NRRL3135 A_terreus A_terreus T_thermo T_lanuginosa M_thermophila T_lanuginosa A_nidulans A_solieAlQK NATSFKGDFA T_solieAlQK NATSFKGDFA T	P_involtus_A2	IKAGLSKLQS	VQNFTDPKFD	FIKSFTYDLG	TSDLVPFGAA	QSFDAGLEVF
P_lycii QVAAVAKIQM ARPFTDPKYE FLNDFVYKFG VADLLPFGAN QSHQTGTDMY A_fumigatus YKKLVTAIQA NATDFKGKFA FLKTYNYTLG ADDLTPFGEQ QLVNSGIKFY ConsphyA YSALIEAIQK NATAFKGKYA FLKTYNYTLG ADDLTPFGEN QMVNSGIKFY YSGLIEAIQK NATSFWGQYA FLESYNYTLG ADDLTIFGEN QMVDSGAKFY YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY YAATIAAIQK SATAFPGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY T_thermo YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY		IQTAVAKLKA	ASNYTDPLLA	FVTNYTYSLG	QDSLVELGAT	QSSEAGQEAF
P_lycii QVAAVAKIQM ARPFTDPKYE FLNDFVYKFG VADLLPFGAN QSHQTGTDMY A_fumigatus YKKLVTAIQA NATDFKGKFA FLKTYNYTLG ADDLTPFGEQ QLVNSGIKFY YSALIEAIQK NATAFKGKYA FLKTYNYTLG ADDLTPFGEN QMVNSGIKFY YSGLIEAIQK NATSFWGQYA FLESYNYTLG ADDLTIFGEN QMVDSGAKFY A_ficuum_NRRL3135 YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY YAATIAAIQK SATAFPGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY T_thermo YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YSQLISRIQK TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY	A pediades	IQAAVKKLQS	AKTYTOPRLD	FLTNYTYTLG	HDDLVPFGAL	QSSQAGEETF
A_fumigatus YKKLVTAIQA NATDFKGKFA FLKTYNYTLG ADDLTPFGEQ QLVNSGIKFY ConsphyA YSALIEAIQK NATAFKGKYA FLKTYNYTLG ADDLTPFGEN QMVNSGIKFY YSGLIEAIQK NATSFWGQYA FLESYNYTLG ADDLTIFGEN QMVDSGAKFY YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YSQLISRIQK TATAYKGYYA FLKDYRYQLG ADDLTPFGEN QMIQLGIKFY YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY	P_lycii					
CONSPHYA A nidulans A_nidulans A_ficuum_NRRL3135 A_terreus T_thermo T_lanuginosa M_thermophila M_thermophila CONSPHYA YSALIEAIQK NATSFWGQYA FLESYNYTLG ADDLTIFGEN QMVDSGAKFY YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY YAATIAAIQK SATAFPGKYA FLQSYNYSLD SEELTPFGRN QLRDLGAQFY YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY	A_fumigatus					
A_nidulans A_ficuum_NRRL3135 A_ficuum_NRRL3135 A_terreus T_thermo T_lanuginosa M_thermophila M_thermophila XSGLIEAIQK NATSFWGQYA FLESYNYTLG ADDLTIFGEN QMVDSGAKFY YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY YAATIAAIQK SATAFPGKYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY						
A_ficuum_NRRL3135 YSALIEEIQQ NATTFDGKYA FLKTYNYSLG ADDLTPFGEQ ELVNSGIKFY A_terreus YAATIAAIQK SATAFPGKYA FLKDYNYSLD SEELTPFGRN QLRDLGAQFY T_thermo YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY						
A_terreus YAATIAAIQK SATAFPGKYA FLQSYNYSLD SEELTPFGRN QLRDLGAQFY T_thermo YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY M_thermophila YVDLIDRIHH GAISYGFGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY	A ficuum NRRL3135					
T_thermo YSQLISRIQK TATAYKGYYA FLKDYRYQLG ANDLTPFGEN QMIQLGIKFY T_lanuginosa YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY M_thermophila YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY						
T_lanuginosa YAELLQRIQD TATEFKGDFA FLRDYAYHLG ADNLTRFGEE QMMESGRQFY M_thermophila YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY						
M_thermophila YVDLIDRIHH GAISYGPGYE FLRTYDYTLG ADELTRTGQQ QMVNSGIKFY ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY						
ISAVLTKIKT SATWYGSDFQ FIKNYDYVLG VDHLTAFGEQ EMVNSGIKFY						
134						
774	•	134				176
151 200						
P_involtus A1 ARYSKLVSKN NLPFIRADGS DRVVDSATNW TAGFASASHNTVQ	P_involtus Al	aryskluskn	NLPFIRADGS	DRVVDSATNW	TAGFASA	
P_involtus_A2 ARYSKLVSSD NLPFIRSDGS DRVVDTATNW TAGFASASRNAIQ	P_involtus_A2					

Fig 174

		49/	51		
T_pubescens	TRYSSLVSAI	TO/		w magnaza	SSNSIT
A_pediades		NI.DEUDAGG	C MEMBERS OF THE STATE OF THE S	W TAGFALA	· · · · · SSNSIT
P lycii		DUDEUDAAC	S NEVVOSAINV	W TEGFSAA	SHHVLN
A_fumigatus		DVFFVRAAG	D QRVVDSSTNV S DRVIASGEKI		SGETVL
consphyA		VVPFIRASG	S DRVIASGEKI		DPGA.TNRAA
A nidulans		IVPFIRASG	S DRVIASAEKI	_	A DPGSQPHQAS
A_ficuum_NRRL3135	OBVECT MON	NTPFIRASG	S DRVVASAEKI		H DHGSKRAT
A_terreus	QRIESH. TRI	IVPFIRSSG	SRVIASGKKI	FIEGFQSTKLE	C DPRAQPGQSS
T thermo		INPEVRATO	A SRVHESAEKE		DHHANPHQPS
T_lanuginosa	MRIKSL.ARN	AVPFVRCSGS	DRVIASGRLE	, iegeosakvi	DPHSDKHDAP
M_thermophila	HRYREQ.ARE	IVPFVRAAGS	ARVIASAEFF	NRGFQDAKDE	DPRSNKDQAE
M_chermophila	RRYRAL, ARK	SIPFVRTAGO	DRVVHSAENF	TOGFHSALLA	DRGSTVRPTL
	OKISSTIDED	TLPFVRASGO) ERVIASAENF	TTGFYSALSA	DKNPPSSLPR
	QTE				
	177				
	201				217
P_involtus A1	··· - 				250
P_involtus_A1	PKTWTTTPÖT.	G. NOTLEDN	MCPAAGD	SDPQVNA	WLAVAFPSIT
T_pubescens	PKTDLTTPQT	G. NOTLEDN	MCPAAGE	SDPQVDA	WLASAFPSVT
A_pediades	PVESVIISEA	GNDTLDDN	MCPAAGD	SDPQVNQ	WLAQFAPPMT
——	PILFVILSES	LNDTLDDA	MCPNAGS	SDPQTGI	WTSIYGTPIA
P_lycii A_fumigatus	PILOVVLQEE	GNCTLCNN	MCPNEVD	GD. ESTT	WLGVFAPNIT
	PAISVIIPES	ETFNNTLDHG	VCTKFEA	SQLGDEVAAN	FTALFAPDIR
consphyA A_nidulans	PAIDAILEG	SGYNNTLDHG	TCTAFED	SELGDDVEAN	FTALFAPAIR
A_ficuum_NRRL3135	PAANATIBET	DGFNNTLDHS	TCVSFEN	DERADEIEAN	FTAIMGPPIR
	PKIDVVISEA	SSSNNTLDPG	TCTVFED	SELADTVEAN	FTATFVPSIR
A_terreus	PRVDVAIPEG	SAYNNTLEHS	lctafes		FTAVFAPAIA
T_thermo			SCPVFED	-	FAKQFAPAIL
T_lanuginosa			TCPAAEE		FLQVFGPRVL
M_thermophila	PYDMVVIPET	AGANNTLHND	LCTAFEEGPY	STIGDDAQDT	YLSTFAGPIT
	P.EMVIISEE	PTANNTMHHG	LCRSFED	STTGDQAQAE	FIAATFPPIT
	218				
	251		•		252
P_involtus_A1		\T MD-mo-2			300
P_involtus_A2	ARLNAAAPSV	MUTDIDAFNL	VSLCAFLTVS	KEKK	s
T_pubescens	AQLNAAAPGA	NLTDADAFNL	VSLCPFMTVS	KEQK	s
A_pediades	ARLNAGAPGA	NLTDTDTYNL	LTLCPFETVA	TERR	s
	NRLNQQAPGA	NITAADVSNL	IPLCAFETIV	KETP	s
P_lycii A_fumigatus	ARLNAAAPSA	NLSDSDALTL	MDMCPFDTLS	SGNA	· · · · ·
consphyA	ARAEKHLPGV	TLTDEDVVSL	MDMCSFDTVA	RTSDASQ.	Es
A_nidulans	ARLEADLPGV	TLTDEDVVYL	MDMCPFETVA	RTSDATE.	····LS
A_ficuum_NRRL3135	KRLENDLPGI	KLTNENVIYL	MDMCSFDTMA	RTAHGTE.	····LS
	QRLENDLSGV				
A_terreus					LS .
T_thermo	EKIKDHLPGV				
T_lanuginosa	KKITKHMPGV	NLTLEDVPLF	MDLCPFDTVG	SDPVLFPRQ.	····LS
M_thermophila	ARVNANLPGA	NLTDADTVAL	MDLCPFETVA	SSSSDPATAD	AGGGNGRPLS
	ARLNAGFKGV	TLSNTDVLSL	MDLCPFDTVA	YPLSSLTTTS	SVSGGGK LS
	Q				
	252				
	253 .				300
P_involtus_A1	301				350
P_involtus_A1 P_involtus_A2	DECELERATE	GSFEAFAYGG	DLDKFYGTGY	GQELGPVQGV	GYVNELIARL
_	DFCTLFEGIP	JSFEAFAYAG	DLDKFYGTGY	GQALGPVQGV	GYINELLARL
T_pubescens	EFCDIYEELQ	AE.DAFAYNA	DLDKFYGTGY	GQPLGPVQGV	GYINELIARL
A_pediades	PFCNLFTP	EEFAQFEYFG	DLDKFYGTGY	GQPLGPVQGV	GYINELLARL
P_lycii	PFCDLFTA	EEYVSYEYYY	DLDKYYGTGP	GNALGPVQGV	GYVNELLARL
\mathtt{A} _fumigatus	PFCQLFTH I	MEMKKANATÖ	SLGKYYGYGA	GNPLGPAQGI	GFTNELIARL

Fig 17P

WO 99/49022 PCT/DK99/00153

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consphyA PFCALFT..H DEWRQYDYLQ SLGKYYGYGA GNPLGPAQGV GFANELIARL

COMPHYA		DEMKČIDITO			
A_nidulans		KEWLQYDYLQ			
A_ficuum_NRRL3135	PFCDLFTH	DEWINYDYLQ	SLKKYYGHGA	GNPLGPTQGV	GYANELIARL
A_terreus	PFCDLFTA	TEWTQYNYLL	SLDKYYGYGG	GNPLGPVQGV	GWANELMARL
T_thermo	PFCALSTQ	EEWQAYDYYQ	SLGKYYGNGG	GNPLGPAOGV	GFVNELIARM
T_lanuginosa		DDWMAYDYYY			
M_thermophila		SEWRAYDYLQ			
		SDWTIYDYLQ	•		
	I CODII A	DDMITIDIDG	DIGRETOTOL	CHOTHWIGGA	GIVNELLAKLI
	201				
e de la companya de	301				349
B 4	351				400
P_involtus_A1		QTNRTLDASP			
P_involtus_A2		QTNRTLDAAP			
T_pubescens		QTNSTLDSSP			
A_pediades		QTNRTLDSSP			
P_lycii	TGQ.AVRDET	QTNRTLDSDP	ATFPLNRTFY	ADFSHDNTMV	PIFAALGLFN
A_fumigatus	TRS.PVQDHT	STNSTLVSNP	ATFPLNATMY	VDFSHDNSMV	SIFFALGLYN
consphyA	TRS. PVQDHT	STNHTLDSNP	ATFPLNATLY	ADFSHDNSMI	SIFFALGLYN
A_nidulans	TQS.PVQDNT	STNHTLDSNP	ATFPLDRKLY	ADFSHDNSMI	SIFFAMGLYN
A_ficuum_NRRL3135	THS.PVHDDT	SSNHTLDSSP	ATFPLKSTLY	ADFSHDNGII	SILFALGLYN
A terreus	TRA. PVHDHT	CVNNTLDASP	ATFPLNATLY	ADFSHDSNLV	SIFWALGLYN
T thermo	THS. PVQDYT	TVNHTLDSNP	ATFPLNATLY	ADFSHDNTMT	SIFAALGLYN
T_lanuginosa		TVNHTLDDNP			
M_thermophila		STNRTLDGDP			
_		TTNSTLDGDE			
	350	*		383	
	401	•			450
P_involtus A1		NPWRT	WRTSSLVPFS	GRMVVERLSC	
P_involtus A2	-	DPNRT			
T_pubescens		DPART			
A pediades		NPKRT			
P_lycii		DENRL			
A_fumigatus		ESAKELDG			
consphyA		ESIEETDG			
A nidulans		ESIQEMDG			
A_ficuum_NRRL3135		ENITQTDG			
A terreus		ESVSQTDG			
T thermo		KSIEETDG			
T_lanuginosa		QPPTGAAADG			
M_thermophila		RDPEELGG			
,	HISPMDQTGD		FKVSWAVPFA	GRVYFEKMVC	DADGDGKIDS
	NTTIP	INYG			
					425
	384				425
_ ,	451				500
P_involtus_A1	FGT	TKVRVLVQDQ	VQPLEFCGGD	RNGLCTLARF	VESQTFARSD
P_involtus_A2		TKVRVLVQDQ			
$ exttt{T_pubescens}$		QSVRLLVNDA			
A_pediades		TFVRILVNDA			
P_lycii		EAVRVLVNDA			
A_fumigatus		PLVRALINDR			
consphyA		PLVRVLVNDR			
A_nidulans		PLVRVLVNDR			
A_ficuum_NRRL3135		PLVRVLVNDR			
A_terreus		PLVRVLVNDR	VMPLHGCPTD	KLGRCKRDAF	VAGLSFAQAG
T_thermo		PVVRVLVNDR			

IKELTFARQG IESMAFARGN VESMEFARRG

51/51

T_lanuginosa M_thermophila	E GRQEKDE	PFVRVLVNDR EMVRVLVNDR ELVRILVNDR	VMTLKGCGAD	ERGMCTLERF	
A_fumigatus consphyA A_nidulans A_ficuum_NRRL3135 A_terreus	D EAQK 426 501 GAGDFEKCFA GAGDFEKCFA GEGDFEKCFA GQGDFEKCFD GQGDFAKCGFGNWGECFSGNWAECFAGNWKTCFTGDWAECFAGNWADCFGNWADCF.	### ### ##############################			•
M_thermophila	GKWDLCFA GEWEECFV				

R

SEQUENCE LISTING

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5		L> in	ntro: 71).	n . (12)	5)												
10	<220 <220 <220	L> CI		. (70)									,				
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20		l> s:		eptio .(64)													
	<400 aago	-	ggc a	aaaci	cato	Me				ı Met						c tac r Tyr	52
25				gct Ala 15			tgg	gttc	atc (ettt	geee	et gt	ctcg	gatgt	=		100
30	taaa	aatao	cta a	aacat	catti	cc a	ccaga				ı Se					g tcc l Ser	153
35	tgt Cys															cac His	201
40	Thr	Trp		caa Gln	Tyr	Ser	Pro		Phe		Val		Ser				249
45				cct Pro													297
				gcc Ala													345
50	gct Ala			acc Thr													393
55	ttt Phe			atc Tle													441

115

120

5	Ini	: gcg	y tto A Phe 125	; GTZ	gaç Glu	g caa u Glr	a gaa n Glu	a ato 1 Met 130	: Va]	aad Asi	tco Sei	ggc Gly	ato Ile 135	Lys	tto Phe	tac Tyr	489
	cag Glr	a Arg	TAL	Ser	tcc Ser	Lev	ato Ile 145	Gln	aca Thr	gaa Glu	a gad 1 Asp	tcg Ser 150	qaA	acg Thr	ctc Leu	ccc Pro	537
10	Phe 155	Val	cgc Arg	gcc	tct Ser	ggc Gly 160	GIn	gaa Glu	cgc Arg	gto Val	ato Ile 165	: Ala	tcc Ser	gcc Ala	gag Glu	aac Asn 170	585
15	ttc Phe	acc	acc Thr	ggc	ttc Phe 175	Tyr	tcg Ser	gcc Ala	ctc Leu	tca Ser 180	Ala	gac Asp	aag Lys	aac Asn	cct Pro 185	cct Pro	633
20	SET	tcc Ser	tta Leu	cca Pro 190	aga Arg	cca Pro	gaa Glu	atg Met	gtc Val 195	atc Ile	att Ile	tct Ser	gag Glu	gag Glu 200	cca Pro	aca Thr	681
25	АІА	Asn	205	Thr	atg Met	His	His	Gly 210	Leu	Сув	Arg	Ser	Phe 215	Glu	qaA	Ser	729
	acc Thr	acc Thr 220	Gly	gac Asp	caa Gln	gcc Ala	caa Gln 225	gcg Ala	gaa Glu	ttc Phe	atc Ile	gcc Ala 230	gcc Ala	acc Thr	ttc Phe	cca Pro	777
30	ccc Pro 235	atc Ile	acc Thr	gcc Ala	cgt Arg	ctc Leu 240	aac Asn	gcc Ala	caa Gln	ggt Gly	ttc Phe 245	aaa Lys	ggc Gly	gtc Val	acc Thr	ctc Leu 250	825
35	tcc Ser	aac Asn	acc Thr	gac Asp	gtc Val 255	cta Leu	tca Ser	cta Leu	atg Met	gac Asp 260	ctc Leu	tgc Cys	ccc Pro	ttt Phe	gac Asp 265	acc Thr	873
40	gtc Val	gcc Ala	tac Tyr	ccc Pro 270	ctt Leu	tcc Ser	tcc Ser	ctc Leu	acc Thr 275	acc Thr	acc Thr	tct Ser	tcc Ser	gtt Val 280	tct Ser	gga Gly	921
45	gly	Gly	aag Lys 285	tta Leu	tcc Ser	ccc Pro	ttc Phe	tgc Cys 290	tct Ser	ctt Leu	ttc Phe	act Thr	gcc Ala 295	agc Ser	gac Asp	tgg Trp	969
	aca Thr	atc Ile 300	tac Tyr	gat Asp	tac Tyr	ctc Leu	cag Gln 305	tcc Ser	cta Leu	gjà aaa	aaa Lys	tac Tyr 310	tac Tyr	ggt Gly	ttc Phe	ggc Gly	1017
50	ccc Pro 315	ggt Gly	aat Asn	tcc Ser	ьeи	gct Ala 320	gcc Ala	acc Thr	cag Gln	Gly	gta Val 325	gly aaa	tac Tyr	gtc Val	aac Asn	gag Glu 330	1065
55	ctt Leu	atc Ile	gcc Ala	Arg	ttg Leu 335	atc Ile	cgt Arg	gct Ala	Pro	gtc Val 340	gta Val	gat Asp	cac His	Thr	acg Thr 345	acc Thr	1113

5														aac Asn 360			1161
								_		-		_		atc Ile	_		1209
10	gct Ala													acc Thr			1257
15	ccg Pro 395													gaa Glu			1305
20											Ala			gtg Val			1353
25							-							gat Asp 440		_	1401
														cgg Arg			1449
30					-		_		_	_	_			gga Gly			1497
35	aag Lys 475													gag Glu			1545
40			tgt Cys		_	tag	ctci	taga									1570
	<212	l> 49 2> PI		crhi	num :	foect	undi:	ssim	ım								
50		0> 2 Leu	Ile	Leu	Met 5	Ile	Pro	Leu	Phe	Ser 10	Tyr	Leu	Ala	Ala	Ala 15	Ser	
	Leu	Arg	Val	Leu 20	Ser	Pro	Gln	Pro	Val 25	Ser	Cys	Asp	Ser	Pro 30	Glu	Leu	
55	Gly	Tyr	Gln 35	Сув	Asp	Gln	Gln	Thr 40	Thr	His	Thr	Trp	Gly 45	Gln	Tyr	Ser	

	Pro	Phe 50	Phe	e Ser	. Val	Pro	Ser 55		Ile	Ser	Pro	Ser 60		Pro	Asp	Gly
5	Cys 65	arg	J Lev	Thr	Phe	Ala 70	Gln	Val	Leu	Ser	Arg		Gly	Ala	Arg	Phe 80
10		Thr	Pro	Gly	Lys 85	Ala	Ala	Ala	Ile	Ser 90		Val	Leu	Thr	Lys 95	
	Lys	Thr	: Ser	100	Thr	Trp	Tyr	Gly	Ser 105	Asp	Phe	Gln	Phe	Ile 110	Lys	Asn
15	Tyr	Asp	115	Val	Leu	Gly	Val	Asp 120	His	Leu	Thr	Ala	Phe 125	Gly	Glu	Gln
	Glu	Met 130	Val	Asn	Ser	Gly	Ile 135	Lys	Phe	Tyr	Gln	Arg 140	Tyr	Ser	Ser	Leu
20	11e 145	Gln	Thr	Glu	Asp	Ser 150	Asp	Thr	Leu	Pro	Phe 155	Val	Arg	Ala	Ser	Gly 160
25	Gln	Glu	Arg	Val	Ile 165	Ala	Ser	Ala	Glu	Asn 170	Phe	Thr	Thr	Gly	Phe 175	Tyr
	Ser	Ala	Leu	Ser 180	Ala	Asp	Lys	Asn	Pro 185	Pro	Ser	Ser	Leu	Pro 190	Arg	Pro
30	Glu	Met	Val 195	Ile	Ile	Ser	Glu	Glu 200	Pro	Thr	Ala	Asn	Asn 205	Thr	Met	His
	His	Gly 210	Leu	Сув	Arg	Ser	Phe 215	Glu	Asp	Ser	Thr	Thr 220	Gly	Asp	Gln	Ala
35	Gln 225	Ala	Glu	Phe	Ile	Ala 230	Ala	Thr	Phe	Pro	Pro 235	Ile	Thr	Ala	Arg	Leu 240
40	Asn	Ala	Gln	Gly	Phe 245	Lys	Gly	Val	Thr	Leu 250	Ser	Asn	Thr	Asp	Val 255	Leu
	Ser	Leu	Met	Asp 260	Leu	Сув	Pro	Phe	Asp 265	Thr	Val	Ala	Tyr	Pro 270	Leu	Ser
45	Ser	Leu	Thr 275	Thr	Thr	Ser	Ser	Val 280	Ser	Gly	Gly	Gly	Lys 285	Leu	Ser	Pro
	Phe	Cys 290	Ser	Leu	Phe	Thr	Ala 295	Ser	Asp	Trp	Thr	Ile 300	Tyr	Asp	Tyr	Leu
50	Gln 305	Ser	Leu	Gly	Lys	Tyr 310	Tyr	Gly	Phe	Gly	Pro 315	Gly	Asn	Ser	Leu	Ala 320
55.	Ala	Thr	Gln	Gly	Val 325	Gly	Tyr	Val	Asn	Glu 330	Leu	Ile	Ala	Arg	Leu 335	Ile
	Arg	Ala	Pro	Val	Val	Asp	His	Thr	Thr	Thr	Asn	Ser	Thr	Leu	Asp	Gly

				340					345					350		
5	Asp	Glu	Lys 355	Thr	Phe	Pro	Leu	Asn 360	Arg	Thr	Val	Tyr	Ala 365	Asp	Phe	Ser
	His	Asp 370	Asn	Asp	Met	Met	Asn 375	Ile	Leu	Thr	Ala	Leu 380	Arg	Ile	Phe	Glu
10	His 385	Ile	Ser	Pro	Met	Asp 390	Asn	Thr	Thr	Ile	Pro 395	Thr	Asn	Tyr	Gly	Gln 400
	Thr	Gly	Asp	Asp	Gly 405	Val	Lys	Glu	Arg	Asp 410	Leu	Phe	Lys	Val	Ser 415	Trp
15	Ala	Val	Pro	Phe 420	Ala	Gly	Arg	Val	Tyr 425	Phe	Glu	Lys	Met	Val 430	Cys	Asp
20	Ala	qaA	Gly 435	Asp	Gly	Lys	Ile	Asp 440	Ser	Asp	Glu	Ala	Gln 445	Lys	Glu	Leu
	Val	Arg 450	Ile	Leu	Val	Asn	Asp 455	Arg	Val	Met	Arg	Leu 460	Asn	Gly	Cys	Asp
25	Ala 465	Asp	Glu	Gln	Gly	Arg 470	Сув	Gly	Leu	Glu	Lys 475	Phe	Val	Glu	Ser	Met 480
	Glu	Phe	Ala	Arg	Arg 485	Gly	Gly	Glu	Trp	Glu 490	Glu	Arg	Cys	Phe	Val 495	

5618-KaPe

POI	Original (for SU	5618-KaPe BMISSION) - printed on 22.03.1999 10:06:53 AM
0-1	Form - PCT/RO/134 (EASY) Indications Relating to Deposited Microorganism(s) or Other Biological Material (PCT Rule 13bis)	
0-1-1	Prepared using	PCT-EASY Version 2.83
		(updated 01.03.1999)
0-2	International Application No	(424464 01:03:1939)
0-3	Applicant's or agent's file reference	5618-KaPe
1	The indications made below relate to the deposited microorganism(s) or other biological material referred to in the description on:	
1-1	page	6
1-2	line	18
1-3	Identification of Deposit	
1-3-1	Name of depositary institution	Centraalbureau voor Schimmelcultures
1-3-2	Address of depositary institution	Oosterstraat 1, Postbus 273, NL-3740 AG
		Baarn, Netherlands
1-3-3	Date of deposit	23 January 1997 (23.01.1997)
1-3-4	Accession Number	CBS 427.97
1-4	Additional Indications	NONE
1-5	Designated States for Which Indications are Made	all designated States
1-6	Separate Furnishing of Indications	NONE
	These indications will be submitted to the International Bureau later	
2	The indications made below relate to the deposited microorganism(s) or other biological material referred to in the description on:	
2-1	page	6
2-2	line	20
2-3	Identification of Deposit	
2-3-1	Name of depositary institution	DSMZ-Deutsche Sammlung von
		Mikroorganismen und Zellkulturen GmbH
2-3-2	Address of depositary institution	Mascheroder Weg 1b, D-38124
		Braunschweig, Germany
2-3-3	Date of deposit	17 March 1999 (17.03.1999)
2-3-4	Accession Number	DSMZ 12742
2-4	Additional Indications	NONE
2-5	Designated States for Which Indications are Made	all designated States
2-6	Separate Furnishing of Indications	NONE
•	These indications will be submitted to the International Bureau later	

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5618-KaPe

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FOR RECEIVING OFFICE USE ONLY

0-4	This form was received with the international application: (yes or no)	yes
0-4-1	Authorized officer	Brown Steeluner
	FOR INT	ERNATIONAL BUREAU USE ONLY
0-5	FOR INT	ERNATIONAL BUREAU USE ONLY

International application No.

PCT/DK 99/00153

A. CLASSIFICATION OF SUBJECT MATTER

IPC6: C12N 9/16, C12N 15/55, A23K 1/165
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: C12N, A23K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Box 5055, S-102 42 STOCKHOLM

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

	JMENTS CONSIDERED TO BE RELEVANT	
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,	 '	
Ρ,Χ	EP 0897985 A2 (F. HOFFMANN-LA ROCHE AG), 24 February 1999 (24.02.99)	1-47
	~-	
x	WO 9735016 A1 (NOVO NORDISK BIOTECH, INC.), 25 Sept 1997 (25.09.97), page 10, line 22 - page 11, line 18, claim 11	1-47
x	EP 0420358 A1 (GIST-BROCADES N.V.), 3 April 1991 (03.04.91), page 10, line 6 - line 14; and the claims	1-47
		

•	Special categories of cited documents:	"T"	later document published after the international filing date or priority
"A"	document defining the general state of the art which is not considered to be of particular relevance		date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"E"	erlier document but published on or after the international filing date	"X"	document of particular relevance: the claimed invention cannot be
"L"	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other		considered novel or cannot be considered to involve an inventive step when the document is taken alone
o	special reason (as specified) document referring to an oral disclosure, use, exhibition or other	"Y"	document of particular relevance: the claimed invention cannot be
	means		considered to involve an inventive step when the document is combined with one or more other such documents, such combination
"P"	document published prior to the international filing date but later than the priority date claimed		being obvious to a person skilled in the art
		~& . "	document member of the same patent family
Date	of the actual completion of the international search	Date	of mailing of the international search report
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6 .	July 1999		1 3 -07- 1999
Nam	e and mailing address of the ISA.	Autho	rized officer
Swe	dish Patent Office		

Carolina Palmcrantz/Els

International application No.

PCT/DK 99/00153 _

C (Continu	ation). DOCUMENTS CONSIDERED TO BE RELEVANT	
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X	WO 9748812 A2 (HER MAJESTY THE QUEEN IN RIGHT OF CANADA), 24 December 1997 (24.12.97), page 13, line 7 - line 12	1-47
A	WO 9114782 A1 (GIST-BROCADES N.V.), 3 October 1991 (03.10.91)	1-47
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Patent document Publication Patent family Publication cited in search report date member(s) date EP 0897010 A2 07/02/99 ΑU 5954398 A 01/10/98 CA 2231948 A 25/09/98 JP 10276789 A 20/10/98 EP 0897985 **A2** 24/02/99 NO 983364 D 00/00/00 WO 9735016 A1 25/09/97 AU 2077197 A 10/10/97 AU 2539197 A 10/10/97 CA 2248980 A 25/09/97 EP 0904383 A 31/03/99 US 5866118 A 02/02/99 WO 9735017 A 25/09/97 EP 0420358 A1 03/04/91 **AT** 180014 T 15/05/99 AU 636673 B 06/05/93 AU 6501190 A 28/04/91 BG 60108 A 15/10/93 CA 2042054 A 28/03/91 CN 1051058 A 01/05/91 DE 420358 T 12/10/95 EP 0779037 A 18/06/97 ES 2072834 T 01/08/95 FI 912530 D 00/00/00 HU 215179 B 28/10/98 JP 4506007 T 22/10/92 LT 1527 A 26/06/95 LT 3957 B 27/05/96 10310 A,B LV 20/10/94 NO 303988 B 05/10/98 PL 167790 B 30/11/95 PL 168470 B 29/02/96 PT 95447 A,B 22/05/91 US 5436156 A 25/07/95 US 5863533 A 26/01/99 WO 9105053 A 18/04/91 WO 9748812 A2 24/12/97 ΑU 3021697 A 07/01/98 CA 2257101 A 24/12/97 EP 0904385 A 31/03/99 NO 985804 D 00/00/00

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				AU		B	26/05/94
				AU	7765691	Α	21/10/91
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